

Artículo Original / Original Article

Antinociceptive and anti-inflammatory effect of *Lippia pedunculosa* Hayek essential oil and its β -cyclodextrin inclusion complex[Efecto antinociceptivo y antiinflamatorio del aceite esencial de *Lippia pedunculosa* Hayek y su complejo de inclusión de β -ciclodextrina]Renan Marinho Braga¹, Humberto Hugo Nunes de Andrade¹, Humberto de Carvalho Aragão Neto¹, Ryldene Marques Duarte da Cruz¹, Davidson Barbosa Assis¹, Luiz Henrique Agra Cavalcante-Silva¹, Sandra Rodrigues-Mascarenhas¹, Liana Clébia de Moraes Pordeus¹, Lucindo José Quintans-Júnior² & Reinaldo Nóbrega de Almeida¹¹Institute of Research in Drugs and Medicines, Federal University of Paraíba, João Pessoa, Brazil²Department of Pharmacy, Federal University of Sergipe, Aracaju, BrazilReviewed by:
Daniela Russo
Basilicata University
ItalyCaroline Gribner
Federal University of Paraná
BrazilCorrespondence:
Reinaldo Nóbrega DE ALMEIDA
reinaldoan@uol.com.brSection **Biological activity**Received: 29 November 2019
Accepted: 27 March 2020
Accepted corrected: 7 May 2020
Published: 30 March 2021**Citation:**Braga RM, de Andrade HHN, Neto HCA, da Cruz RMD, Assis DB, Silva LHAC, Rodrigues-Mascarenhas S, Pordeus LCM, Júnior LJQ, de Almeida RN. Antinociceptive and anti-inflammatory effect of *Lippia pedunculosa* Hayek essential oil and its β -cyclodextrin inclusion complex. **Bol Latinoam Caribe Plant Med Aromat** 20 (2): 162 - 176 (2021). <https://doi.org/10.37360/blacpma.21.20.2.13>**Abstract:** *Lippia pedunculosa* Hayek (EOLp) presents tripanocid and amebicid effects. However essential oil needs to be further studied in experimental models of analgesia and inflammation once the prevalence of pain in the population generates great suffering and disability and the drugs most often used have undesirable side effects. We also evaluated whether the inclusion complex formulation EOLp/ β -cyclodextrin (β -CD) was able to improve the antinociceptive activity of the EOLp alone. Data were evaluated by analysis of variance (ANOVA), followed by Tukey's test. Differences were considered significant if $p < 0.05$. EOLp presented better antinociceptive effect when compared to the EOLp/ β -CD inclusion complex. Thus, cyclodextrins appear not to be efficient for essential oils with peroxide substances. However, in peritonitis, EOLp reduced total leucocyte migration and IL-1 β levels in the peritoneal fluid, which confirmed its anti-inflammatory effect. The observed effects suggest that EOLp is the best promising option for the treatment of inflammation and pain-related disorders.**Keywords:** Essential oil; *Lippia pedunculosa*; Cyclodextrin; Pain; Inflammation.**Resumen:** *Lippia pedunculosa* Hayek (EOLp) presenta efectos tripanocidas y amebicidas. En este trabajo se estudia su aceite esencial en modelos experimentales de analgesia e inflamación una vez que la prevalencia del dolor en la población genera un gran sufrimiento y discapacidad, y los medicamentos que se usan con mayor frecuencia tienen efectos secundarios indeseables. También se evalúa si la formulación del complejo de inclusión EOLp/ β -ciclodextrina (β -CD) fue capaz de mejorar la actividad antinociceptiva de la EOLp sola. Los datos se evaluaron mediante análisis de varianza (ANOVA), seguido de la prueba de Tukey. Las diferencias se consideraron significativas si $p < 0,05$. EOLp presentó un mejor efecto antinociceptivo en comparación con el complejo de inclusión EOLp/ β -CD. De esta manera, las ciclodextrinas parecen no ser eficientes para aceites esenciales con sustancias de peróxido. Sin embargo, en peritonitis, EOLp redujo la migración total de leucocitos y los niveles de IL-1 β en el líquido peritoneal, lo que confirma su efecto antiinflamatorio. Los efectos observados sugieren que EOLp es una buena y prometedora opción para el tratamiento de la inflamación y los trastornos relacionados con el dolor.**Palabras clave:** Aceite esencial; *Lippia pedunculosa*; Ciclodextrina; Dolor; Inflamación

INTRODUCTION

In most diseases such as autoimmune, gastrointestinal, neurodegenerative, and respiratory diseases, cancers, and infections; inflammation and pain are both cause and consequence. To relieve pain and to turn the treatment of disease into a less traumatic process is critical, and in the pharmaceutical search for adequate solutions, has a long history. The drugs most often used to treat pain and inflammation, despite their well-known adverse effects, are the non-steroidal anti-inflammatory (NSAIDs), and opioids (Pang *et al.*, 2012; Lenardão *et al.*, 2016). This situation has inspired researches with essential oils and their components which could become new analgesic medicines to treatment for painful disorders (De Sousa, 2011; Guimarães *et al.*, 2013), including orofacial pain (Damasceno *et al.*, 2016; Tomaz-Morais *et al.*, 2017; Melo *et al.*, 2017; Siqueira-Lima *et al.*, 2017).

Essential oils (EOs) and monoterpenes present a series of limitations like low solubility in water, slow dissolution rate, instability, and short half-life (Siqueira-Lima *et al.*, 2014). Yet, formation of cyclodextrin (CD) complexes has been successfully used to improve the therapeutic properties of non-polar natural products (Pinho *et al.*, 2014; de Oliveira *et al.*, 2015; Rodrigues *et al.*, 2017). CDs are cyclic oligosaccharides composed of either: 6, 7, 8, or 9 glucopyranose units, respectively (α -, β -, γ - or δ -CD). They present a relatively hydrophilic surface and a lipophilic central cavity (Li & Loh, 2008; Hwang *et al.*, 2012; Andrade *et al.*, 2017). CDs are an important new class of pharmaceutical excipients, which promotes the formation of inclusion complexes with these molecules, which has improved stability, solubility and bioavailability of nonpolar substances, as well as transforming essential oils (liquids) into powders of easy dispersion in water, facilitating the control of its volatility (Siqueira-Lima *et al.*, 2014).

β -cyclodextrins (β -CD(s)) have been extensively used to form inclusion complexes with EOs from: *O. basilicum* (Nascimento *et al.*, 2014), *Lavandula viridis*, *Lavandula pedunculata*, *Thymus lotocephalus* (Costa *et al.*, 2015), *Hyptis pectinata* (Menezes *et al.*, 2015), *Cymbopogon winterianus* (Santos *et al.*, 2015), and *Lippia grata* (Siqueira-Lima *et al.*, 2014), and thus improve their analgesic profile.

The genus *Lippia* (Verbenaceae) is distributed across the South and Central Americas and tropical Africa (Terblanché & Kornelius, 1996).

There are almost 200 species, some of which are already used to promote analgesic and anti-inflammatory effects (Pascual *et al.*, 2001; Siqueira-Lima *et al.*, 2014). *Lippia pedunculosa* Hayek, popularly known as “Pai-pedro”, is a shrub found in the northeast (Alagoas and Sergipe), and southeastern (São Paulo) regions of Brazil (Santos, *et al.*, 2009; Silva *et al.*, 2013). Certain biological properties of the essential oil from the leaves of *Lippia pedunculosa* (EOLp) have already been described. *In vitro* assays have demonstrated promising trypanocidal (Menezes *et al.*, 2014) and amoebicidal activity (de Amorim Santos *et al.*, 2016), this due to the presence of the monoterpenes rotundifolone (71.7%) and (R)-limonene (21.8%); the principal compounds in EOLp (Santos, *et al.* 2014). Several studies have attempted to improve the pharmacological profile of *Lippia* species essential oils using a β -CD inclusion complexes (Fernandes *et al.*, 2004; Marreto *et al.*, 2008; Arana-Sánchez *et al.*, 2010; Siqueira-Lima *et al.*, 2014). Our collaborators started the evaluation of the analgesic profile of EOLp with a preliminary formalin test which indicated a possible antinociceptive effect (Menezes *et al.*, 2018).

So, the aim of this study was to confirm the antinociceptive and anti-inflammatory effect of EOLp and a β -CD inclusion complex containing *L. pedunculosa* leaf essential oil (EOLp/ β -CD) in experimental protocols of pain and inflammation in mice and investigate whether EOLp/ β -CD complexing improves the pharmacological activity of isolated EOLp as well as evaluating potential pain modulation interactions involving important neurotransmitter pathways.

MATERIALS AND METHODS

Plant material and essential oil

Fresh *L. pedunculosa* leaves were collected in October 2013 in *Poço Redondo*, Sergipe, in northeastern Brazil (S 09° 40' 46" W 37° 39' 41"). *L. pedunculosa* was identified by Dr. Ana Paula do Nascimento Prata (Federal University of Sergipe - Herbarium, HFUS); a voucher specimen has been deposited (HFUS 23159).

Pharmacological Procedures

Animals

Male Swiss mice (25–35 g), 2–3 months of age were used. They were randomly kept in appropriate cages at $21 \pm 2^\circ\text{C}$ in a 12-hr light/dark cycle (light from 06:00 am to 06:00 pm) with free access to food

(Purina®, São Paulo, SP, Brazil), and water. All experiments were carried out between 01:00 pm and 05:00 pm, nociception tests were performed by the same visual observer in a double-blind study.

Ethical approval

All applicable international, national, and/or institutional guidelines for the care and use of animals were followed. All procedures performed in studies involving animals were in accordance with the ethical standards of the institution or practice at which the studies were conducted. Experimental protocols were approved by the Animal Care and Use Committee of the Federal University of Paraíba (0159/2015).

Evaluation of motor activity

The motor activity was evaluated in a rota-rod apparatus (Dunham & Miya, 1957). Initially, mice were selected 24 h previously by eliminating animals unable to remain on the rota-rod apparatus (Insight ©, Ribeirão Preto, SP, Brazil) for a period of 60s (7 rpm). Then, the selected animals were then divided into four groups (n = 6, per group) and treated p.o. with vehicle, EOLp, or EOLp/β-CD (400 mg/kg, p.o.). At 1, 1.5, and 2 h each animal was placed on a rota-rod apparatus, and the time (in seconds) completed on the bar was measured for up to 180 sec.

Acetic acid-induced writhing

This study was performed according to Koster & Anderson (1959), Mice (n=6, per group) were pre-treated with EOLp (200, 300, 400 mg/kg, p.o.), EOLp/β-CD (200, 300, 400 mg/kg, p.o.), morphine (6 mg/kg, i.p.), or the EOLp dilution vehicle (Tween-80 5%, p.o.). After 1 h, the mice received a 1% acetic acid injection (i.p.), which caused writhing characterized by extension of the hind limbs and contraction of the abdominal muscles. The number of abdominal writhes was observed for a period of 15 min; after a latency period of 5 min.

Formalin-induced nociception

The procedure described by Hunskaar & Hole (1987) was used. Nociception was induced by injecting (s.c.) 20 μL of 2% formalin in the subplantar region of the right hind paw. The mice (n=6, per group) had previously received EOLp (200, 300, 400 mg/kg, p.o.), EOLp/β-CD (200, 300, 400 mg/kg, p.o.), morphine (6 mg/kg, i.p.), or vehicle 1h prior to the injection of formalin. They were individually placed in mirrored chambers (25 X 25 X 25 cm), and the

time (in seconds) spent licking their injected paws was immediately recorded. Nociception was quantified for 5 min after formalin injection (first phase, neurogenic) and again at 15-30 min (second phase, inflammatory).

Formalin, glutamate, and capsaicin-induced orofacial nociception

Orofacial nociception protocols were performed on the mice by s.c. injection of 20 μL 2% formalin, 40 μl of glutamate (25 mM), or capsaicin (20 μL, 2.5 μg) into the right upper lip (perinasal area) using a 27-gauge needle. Mice (n = 6, per group) were treated with either vehicle, EOLp, EOLp/β-CD (300 mg/kg, p.o.), or morphine (6 mg/kg, i.p.) at 1h before specific algogen administrations. Formalin test: The neurogenic phase occurs within 0–5 min after the administration of formalin and is followed by a latency period of about 10 min. An inflammatory phase occurs within 15-40 min after formalin administration. Glutamate test: Mice were observed individually for 15 min following glutamate injection. Capsaicin test: Animals were assessed during the 10-20 min after injection of capsaicin. The animals were observed individually in mirrored chambers (25 X 25 X 25 cm) to allow an unobstructed view of the orofacial region. For the three tests, the nociceptive behavior was assessed during the period of time in which the animals continued rubbing the orofacial region.

Hot Plate Test

The animals were pre-selected, and those considered suitable presented a pain response time of less than 10s when placed on the hot plate apparatus (Insight ©, Ribeirão Preto, SP, Brazil) at 55 ± 1°C (Eddy & Leimbach, 1953). The selected mice were pre-treated with vehicle, EOLp, EOLp/β-CD (400 mg/kg, p.o.), or morphine (6 mg/kg, i.p.); and they were individually placed on the hot plate at 0.5, 1, and 2h after initial treatment. The registered parameter was the latency time to jump or lick the hind paws. In order to minimize animal paw tissue destruction, time on the plate did not exceed 30s.

Carrageenan-induced peritonitis test

Leukocyte migration was induced by intraperitoneal administration of carrageenan (1%, 300 μL) in mice (n=6) treated 1h earlier with either vehicle, EOLp (200, 300, and 400 mg/kg, p.o.), or dexamethasone (2 mg/kg, i.p.). Four hours after administration of the stimulus, the mice were euthanized, and the

peritoneal cavity was washed with 2 mL of saline. From the peritoneal fluid an aliquot was removed for a total leukocyte count. The results were expressed as the number of leukocytes/mL.

Dosage of TNF- α and IL-1 β in the peritoneal fluid

Four hours after carrageenan administration to groups pre-treated with vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), or dexamethasone (2 mg/kg, i.p.), peritoneal liquid was removed and centrifuged at 1500 rpm/5 min/4°C. The supernatant was removed for measurement of TNF- α and IL-1 β using sandwich cytokine ELISA performed according to the manufacturer's instructions (Boster Biological Technology CO., Ltd, Pleasanton, CA, USA). The cytokine levels were calculated from standard curves and expressed as total quantity per milliliter (pg/mL).

Statistical analysis

The data obtained were evaluated using the GraphPad Prism (v 4.00) software (San Diego, CA, USA); by one and two-way analysis of variance (ANOVA); followed by Tukey's test. Differences were considered significant if $p < 0.05$.

RESULTS

EOLp effect on motor coordination

Treatment with EOLp (200, 300, and 400 mg/kg, p.o.) did not alter the rotating bar permanence times: observed at 60 (178.8 \pm 0.8; 146.3 \pm 19.3; 157.5 \pm 14.7s), 120 (174.5 \pm 3.6; 152.0 \pm 11.4; 173.0 \pm 3.5s), and 180 (176.3 \pm 2.4; 176.0 \pm 1.4; 150.5 \pm 11.1s) minutes from initial treatment; as compared to the controls (180.0 \pm 0.0; 176.0 \pm 1.6; 171.0 \pm 3.4s)(Figure N° 1).

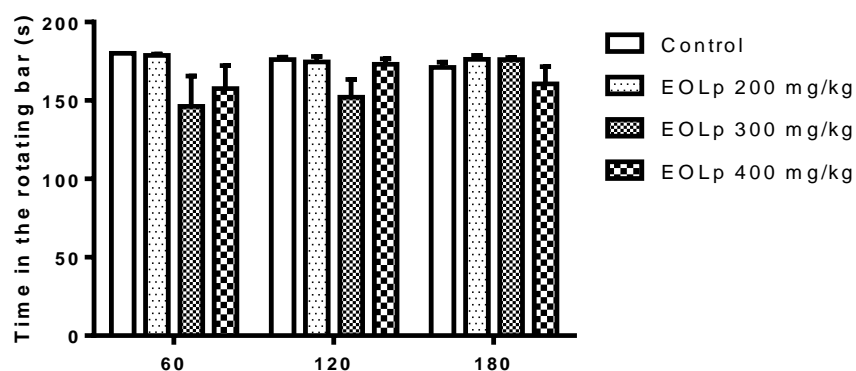


Figure N° 1

Effect of EOLp administration on motor coordination. Mice were pretreated with vehicle, or EOLp (200, 300 and 400 mg/kg, p.o.); and at 60, 120 and 180 min later were placed on a rotating rod. Each column represents the mean \pm S.E.M. (n = 6). Statistical analysis: ANOVA “one-way”, followed by Tukey's test

EOLP and EOLp/ β -CD effects on acetic acid test induced contortions

EOLp in doses of 300 and 400 mg/kg presented a significant reduction (11.3 \pm 1.7; 5.7 \pm 2.2) in the number of writhings induced by acetic acid as compared to the control group (34.5 \pm 2.0). EOLp/ β -CD did not reduce contortions at any of the tested doses. Morphine, as expected decreased the number of writhes (2.0 \pm 0.5) (Figure N° 2).

EOLP and EOLp/ β -CD effects on formalin-induced nociception

The EOLp in doses of 300 (47.1 \pm 4.7) and 400 (21.1

\pm 7.2) mg/kg presented a significant reduction in the paw licking time for the first phase when compared to the control group (74.5 \pm 3.1). Morphine (6 mg/kg), as expected decreased the same parameter (21.7 \pm 2.4) (Figure N° 3.1). During the second phase of the test, EOLp in doses of 300 (20.1 \pm 7.2) and 400 (0.2 \pm 0.1) mg/kg also reduced paw licking time significantly when compared to the control group (123.9 \pm 8.4). Morphine (6 mg/kg), as expected decreased the same parameter (26.0 \pm 5.1) (Figure N° 3.2). The animals treated with EOLp (200 mg/kg), and EOLp/ β -CD (at all doses) did not present reduced paw licking times.

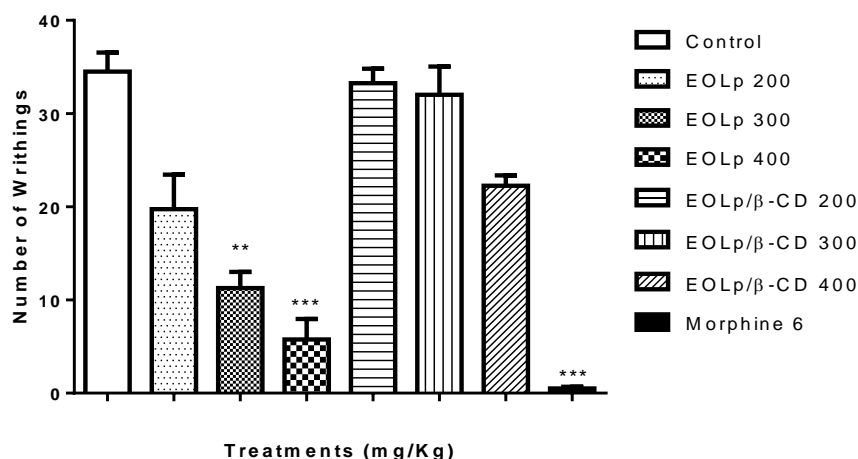


Figure N° 2

Effect of EOLp and EOLp/β-CD treatment on abdominal contortions induced by acetic acid tests. Mice were pretreated with either vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), EOLp/β-CD (200, 300 and 400 mg/kg, p.o.), or morphine (6 mg/kg, i.p.). Each column represents mean ± S.E.M. (n = 6) (ANOVA “one-way”–Kruskal-Wallis followed by Dunn's test) ** $p < 0.01$, *** $p < 0.001$ vs control

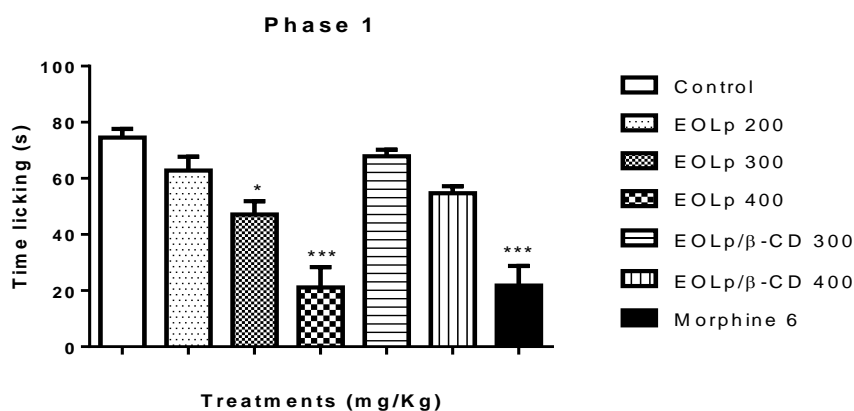


Figure N° 3.1

Effect of EOLp and EOLp/β-CD treatments on the first phase of the formalin-induced nociception test. Mice were pretreated with vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), EOLp/β-CD (200, 300 and 400 mg/kg, p.o.), or morphine (6mg/kg, i.p.). Each column represents mean ± S.E.M. (n = 6) (ANOVA “one-way”–Kruskal-Wallis followed by Dunn's test) * $p < 0.05$, *** $p < 0.001$ vs control

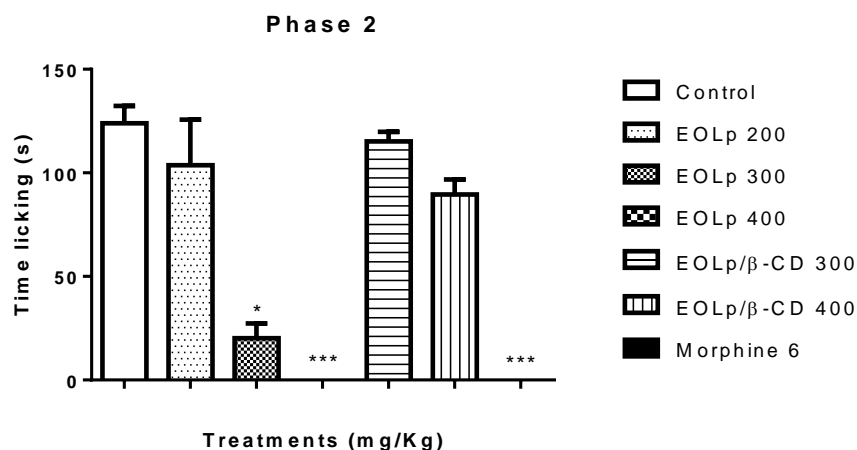


Figure N° 3.2

Effect of EOLp and EOLp/β-CD treatment on the second phase of the formalin-induced nociception test. Mice were pretreated with vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), EOLp/β-CD (200, 300 and 400 mg/kg, p.o.), or morphine (6 mg/kg, i.p.). Each column represents mean ± S.E.M. (n = 6) (ANOVA “one-way”– Kruskal-Wallis followed by Dunn's test) * $p < 0.05$, *** $p < 0.001$ vs control

Formalin, glutamate, and capsaicin-induced orofacial nociception

Administration of EOLp (300 mg/kg) produced a reduction in face-rubbing behavior induced by formalin in both phases ($p < 0.05$; $p < 0.001$), glutamate ($p < 0.01$), and capsaicin ($p < 0.001$), when compared

with the control groups (Table N° 1). Conversely, these outcomes were not observed for EOLp/β-CD; which was unable to inhibit formalin, glutamate, or capsaicin-induced orofacial nociception behavior. As expected, morphine was able to reduce nociceptive behavior in all analyses.

Table N° 1

Effect of EOLp, EOLp/β-CD, and morphine on formalin, glutamate, and capsaicin-induced orofacial nociception.

| Treatment | Dose (mg/kg) | Formalin test | | Glutamate test ¹ | Capsaicin test ¹ |
|-----------|--------------|-------------------------|-------------------------|-----------------------------|-----------------------------|
| | | 0-5min ¹ | 15-40min ¹ | | |
| Control | - | 59.1 ± 3.9 | 101.8 ± 9.4 | 70.9 ± 3.5 | 103.0 ± 3.6 |
| EOLp | 300 | 15.6 ± 5.2 ^a | 27.0 ± 5.0 ^c | 18.4 ± 4.2 ^b | 5.1 ± 1.4 ^c |
| EOLp/β-CD | 300 | 23.1 ± 6.4 | 85.1 ± 12.3 | 40.6 ± 7.3 | 57.2 ± 8.6 |
| Morphine | 6 | 5.5 ± 3.2 ^c | 2.1 ± 0.5 ^c | 7.6 ± 0.8 ^c | 3.4 ± 1.0 ^c |

¹Values represent mean ± S.E.M.

^a $p < 0.05$; ^b $p < 0.01$ ^c $p < 0.001$ Kruskal-Wallis followed by Dunn's test), significantly different from control group

EOLP and EOLp/β-CD effect - hot plate test

After treatment with EOLp the results at doses of 200, 300, and 400 mg/kg did not increased latency

when compared to the control. As expected, morphine increased the reaction time for all times tested (Figure N° 4).

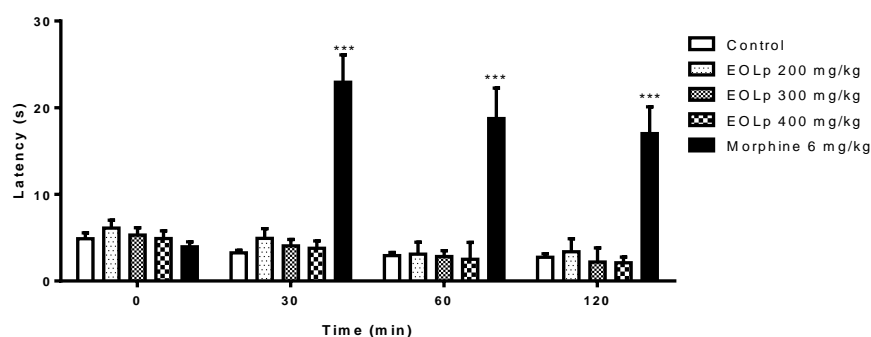


Figure N° 4

Effect of EOLp administration during the hot plate test. Mice were pretreated with vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), or morphine (6 mg/kg, i.p.). Measurements were performed at 30, 60, and at 120 min after treatment. Each column represents mean \pm S.E.M. (n = 6). Statistical analysis: ANOVA “one-way” followed by Tukey's test. *** $p < 0.001$ vs control group

EOLp effect on carrageenan-induced peritonitis test

Treatment with EOLp 1h before carrageenan administration (1%, 300 μ L) inhibited leukocyte migration at doses of 300 and 400 mg/kg (4.3 ± 0.8 ;

2.3 ± 0.2) when compared to the control (12.0 ± 1.1). As expected, dexamethasone reduced leukocyte migration (4.5 ± 1.0) (Figure N° 5).

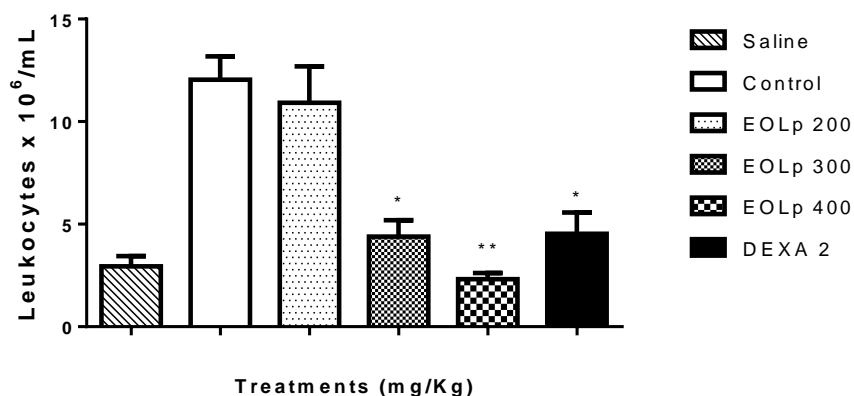


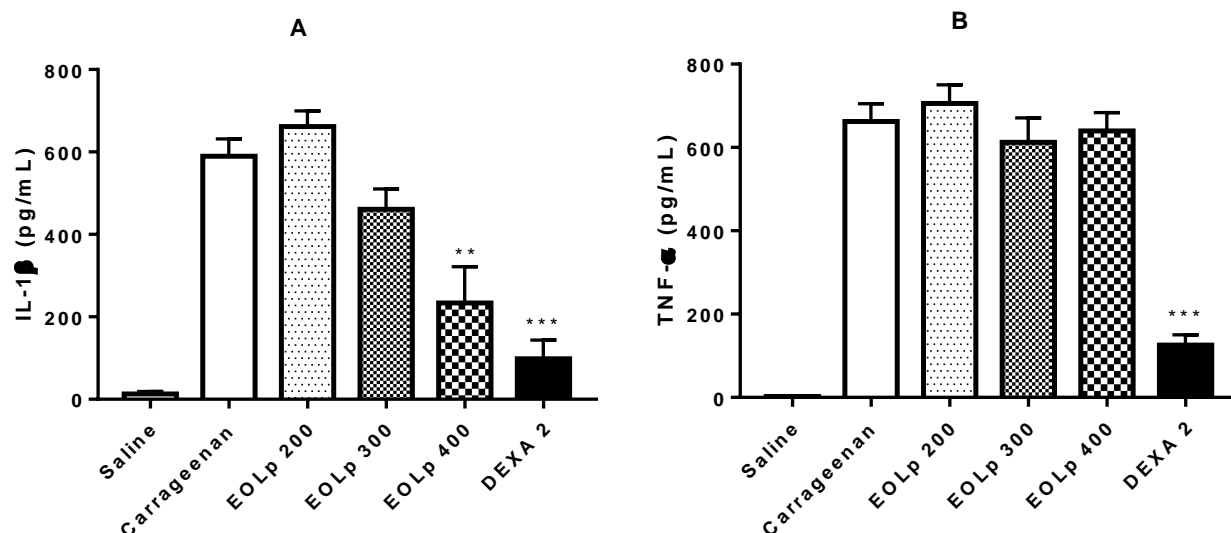
Figure N° 5

Effect of EOLp on total leukocyte migration in carrageenan-induced peritonitis, by injection of carrageenan (1%, 300 μ L) into the mice peritoneal cavities at 1 hour after administration of either vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), or dexamethasone (DEXA, 2 mg/kg i.p.). Each column represents mean \pm S.E.M. (n = 6). Statistical analysis: ANOVA “one-way” followed by Dunnett's test. * $p < 0.05$, ** $p < 0.01$ versus control group

EOLp effect on TNF- α and IL-1 β

Four hours after carrageenan administration, EOLp at a dose of 400 mg/kg (233.6 ± 87.9 pg/mL) reduced ($p < 0.01$) IL-1 β levels compared to the control group (589.4 ± 42.1 pg/mL). However, when evaluating TNF- α levels, we observed that EOLp (at doses of

200, 300, and 400 mg/kg) was unable to promote reductions (705.4 ± 45.0 ; 612.3 ± 58.4 ; 639 ± 44.0 pg/mL) when compared to the control group (662.5 ± 42.4 pg/mL). Dexamethasone reduced both IL-1 β (98.0 ± 45.7 pg/mL) and TNF- α (125.7 ± 24.3 pg/mL) levels (Figures No. 6a & 6b).



Figures No. 6a & 6b

EOLp effect on levels of IL-1 β (A) and TNF- α (B) in the peritoneal cavity 4h after intraperitoneal administration of carrageenan. Mice were pretreated with vehicle, EOLp (200, 300 and 400 mg/kg, p.o.), and dexamethasone (DEXA, 2 mg/kg i.p.), 1h before the injection of a phlogistic stimulus. Each column represents mean \pm S.E.M. (n = 6). Statistical analysis: ANOVA “one-way” followed by Dunnett’s test.

** $p < 0.01$, *** $p < 0.001$ vs control group

DISCUSSION

Previous studies have suggested that central nervous system (CNS) depression and non-specific muscle relaxation effects can reduce motor response coordination and invalidate behavioral test results (de Almeida *et al.*, 2004; Quintans Júnior *et al.*, 2010). We therefore evaluated the effect of EOLp with a rota-rod apparatus and no significant alteration in mice motor coordination was observed. Thus, the activity observed for EOLp in the nociception tests cannot be attributed to inhibitory effects on the CNS or muscle relaxation.

In order to solubilize nonpolar molecules or complex mixtures such as essential oils, the formation of inclusion complexes with CDs has been extensively used (Santos *et al.*, 2015). Besides protect physicochemical properties of the essential oils from the oxidation, high temperatures, light degradation and volatilization (Marreto *et al.*, 2008), in natural products, β -CD complexation can improve pharmacological properties; whether analgesic or anti-inflammatory (Quintans-Júnior *et al.*, 2013; Siqueira-Lima *et al.*, 2014; Rodrigues *et al.*, 2017). Therefore, whether β -CD complexation might improve the antinociceptive profile of the EOLp was evaluated. We initially performed the acetic acid-induced writhing test; a classical model of visceral

pain used as a screening tool for evaluation of analgesic or anti-inflammatory drugs (Mohamad *et al.*, 2010). When administered by intraperitoneal injection, acetic acid promotes activation of cationic channels expressed in peripheral fibers and also stimulates the release of inflammatory mediators like bradykinin, histamine cytokines, and eicosanoids (Deraedt *et al.*, 1980). These mediators can increase vascular permeability, reduce the nociception threshold, and stimulate nociceptive fiber nerve terminals (Le Bars *et al.*, 2001).

Our results indicated that EOLp (300 and 400/mg/kg, p.o.) reduced the number of writhes induced by acetic acid (Figure No. 2). Similar to species of the same genus such as *L. alba* (Costa *et al.*, 1989; Viana *et al.*, 1998), *L. multiflora* (Abena *et al.*, 2003), *L. nodiflora* (Ahmed *et al.*, 2004), and *L. gracilis* (Mendes *et al.*, 2010; Guilhon *et al.*, 2011; Guimarães *et al.*, 2012). Rotundifolone, the major constituent of EOLp (71.7%) (Santos, *et al.*, 2014), also presents efficacy equivalent to EOLp in acetic acid-induced writhings (Sousa *et al.*, 2009). However, EOLp/ β -CD presented no effect in any of the doses we tested.

It is already described in the literature the possibility of CDs to increase physicochemical properties of apolar substances, also pharmacological

properties in relation to nociception and inflammation, (de Oliveira *et al.*, 2015; Brito *et al.*, 2015; Lima *et al.*, 2016; Santos *et al.*, 2017). Oil extracts (EO) from *Lippia grata* (Siqueira-Lima *et al.*, 2014), *Ocimum basilicum* (Nascimento *et al.*, 2015) and *Hyptis pectinata* (Menezes *et al.*, 2015), in their inclusion complex with CDs, have all exhibited superior antinociceptive effect when compared to their isolated form. Surprisingly, this was not found in our study; EOLp tested was more effective in reducing antinociceptive behavior. The results presented in this study clearly demonstrate that EOLp/ β -CD lost its analgesic properties when compared with equivalent doses of EOLp alone. This is the first essential oil that presented minor analgesic profile when complexed with β -CD, which usually improves it (Nascimento *et al.*, 2017). Araujo-Filho *et al.*, (2017) recently demonstrated that β -CD improves the analgesic effect of D-limonene. Thus, the weak effect found for EOLp/ β -CD may be due to stereo-chemical difficulties in rotundifolone cavity or surface complexing with β -CD (Lima *et al.*, 2016; Menezes *et al.*, 2018).

Proceeding with antinociceptive evaluation, yet due to the low specificity of the acetic acid-induced writhing test, it is not possible to us confirm if the antinociception produced was central or peripheral. We therefore used more specific nociception models, such as the formalin test, in order to more precisely characterize the antinociceptive activity exhibited. The formalin-induced nociception test it is a biphasic experimental pain model. The first phase (neurogenic) is related to the activation of TRPA1 receptors (McNamara, *et al.*, 2007) at the nociceptive nerve endings, which promote pain by the release of substance P, glutamate, and bradykinin (Hunskar & Hole, 1987; Tjølsen *et al.*, 1992). After the first and before de second phase, there is a moment known as 'interphase' or 'quiescent period' which nociceptor excitability is decreased (Tjølsen *et al.*, 1992). The second (inflammatory) phase occurs due to spinal cord stimulus, after sensitization of nociceptors, with the release of serotonin, histamine, prostaglandins (PGE₂), nitric oxide (NO), glutamate, aspartate, and bradykinin (Henry *et al.*, 1999; Silva *et al.*, 2016). Diminished paw licking time in both phases of the formalin test is characteristic of drugs acting at central levels, such as opioid analgesics. However, peripheral non-steroidal anti-inflammatory and corticosteroid drugs cause inhibition only at the second phase (Mansouri *et al.*, 2014).

Acute treatment with EOLp (300 and 400 mg/kg, p.o.) produces significant inhibition of nociceptive behavior in both phases of the formalin test (Figure No. 3A); similar to *L. gracilis* essential oil at the same dose (400 mg/kg, p.o.), but different from rotundifolone (200 mg/kg, p.o.) (Sousa *et al.*, 2009) and R-(+)-limonene (50 mg/kg, i.p.) (do Amaral *et al.*, 2007), which inhibited only the second phase; indicating that the antinociceptive effects of these drugs may originate in anti-inflammatory events. Similarly, we also evaluated orofacial nociception by upper lip injection of formalin (s.c.) to induce a biphasic nociceptive response with ipsilateral forepaw face-rubbing episodes, sometimes using the hind paw, directed to the perinasal area (Luccarini *et al.*, 2006). The administration of EOLp (300 mg/kg, p.o.) decreased face-rubbing behavior during both phases of formalin-induced orofacial nociception (Table No. 1), potentially suggesting central antinociceptive activity.

It is already known that glutamate participates of the nociception signals from the nociceptive nerve ending to the dorsal horn of the spinal cord and the trigeminal sub nucleus caudalis. Blockage of glutamate reuptake has been indicated as a new method to treat chronic inflammatory pain (Yang *et al.*, 2015). Glutamate injection evokes pronounced nociceptive response which is mediated by neuropeptides (Substance P) released from C fibers, and activation of glutamate ionotropic receptors such as methyl-D-aspartate (NMDA). This promote the production of NO, and pro-inflammatory cytokines, such as TNF- α and IL-1 β , which act in neuronal excitability (Millan, 1999; Pereira *et al.*, 2010). Since pretreatment with EOLp (300 mg/kg, p.o.) significantly protected against glutamate induced orofacial nociception (Table No. 1), we suggest that antinociceptive effect of EOLp may be related to glutamate receptors, probably NMDA, which can limit mediator production (Ribas *et al.*, 2008; Bonjardim *et al.*, 2011).

Sakurada *et al.* (2003) proposed using the capsaicin-induced nociception test for studying compounds which act on neurogenic pain. Extracted from the red pepper, capsaicin injected into animals, produces inflammation and increases the excitability of spinal and trigeminal nociceptive neurons (Pelissier *et al.*, 2002), and in animals, promotes nociceptive behavior (Hu *et al.*, 2005; Lam *et al.*, 2009). Capsaicin activates the sensitive transient receptor potential of vanilloid 1 (TRPV1), which is an important receptor in pain transduction, increasing

the expression of TNF receptor 1 (TNFR1) (Waning *et al.*, 2007; Ma *et al.*, 2009), and the release of neuropeptides such as tachykinins (neurokinin A), substance P, calcitonin, glutamate, aspartate, oxide nitric, and other pro-inflammatory substances (Honda *et al.*, 2008). As demonstrated in Table No. 1, pretreatment with EOLp (300 mg/kg, p.o.) produced a significant reduction in neurogenic inflammatory nociception when induced by capsaicin injection into the right upper lip (perinasal area). As this effect was similar to that of a glutamate model; recently Ro *et al.* (2014) showed that there is an interaction between NMDA and TRPV1 receptors in the trigeminal ganglion, and they concluded that TRPV1 is an integrator of glutamate receptor signaling in trigeminal muscle nociceptors (Damasceno *et al.*, 2016); thus, we suggest that EOLp could antagonize the TRPV1 channel leading to glutamate release reductions.

In an attempt to confirm that EOLp presents central analgesic effects we performed hot plate testing, which is a behavioral model consisting of exposing the animal to a noxious thermal stimulus. The nociceptive parameters evaluated: the time to animal to jump or lick the hind leg represent integrated supra-spinal responses. Thermal stimulation is associated with activation of TRPV1 receptors at A δ and C fibers, which transmit nociceptive signals to the dorsal horn of the spinal cord, and finally to the somatosensory cortex (Yi *et al.*, 2014). Due to its activation of the CNS, this model is used to evaluate opioid, sedative, and hypnotic drug effects (Yeung *et al.*, 1977). Our data showed (Figure No. 3B), that the animals pretreated with EOLp do not present increased hot plate response latency, confirming that the analgesic effects presented by EOLp are not mediated at CNS levels.

Since EOLp presented analgesic effect in the second phase of both formalin tests, and during glutamate and capsaicin induced orofacial nociception, it seems to involve anti-inflammatory events. Since mediators produced in inflammation sites can promote pain by activation or sensitization of nociceptors adjacent to the injured tissue, the anti-inflammatory activity of EOLp was therefore also investigated. IL-1 β , IL-6, and TNF- α are pro-inflammatory cytokines found to increase the production of substance P and PGE₂ in a number of neuronal and glial cells, contributing to the development of pain behavior following a peripheral nerve injury (Carvalho *et al.*, 2013; de Almeida *et al.*,

2017).

In order to evaluate possible anti-inflammatory effect, carrageenan-induced peritonitis was promoted. Carrageenan is a linear sulfated polysaccharide derived from red algae with robust inflammatory properties, and is often used in models to screen for molecules with anti-inflammatory activity. Carrageenan-induced peritonitis techniques allow quantification of leukocytes migrating into the peritoneal cavity through the activity of chemotactic agents. After injection of carrageenan, EOLp (300 and 400 mg/kg) significantly prevented peritoneal cavity leukocyte recruitment. The results are consistent with a previous study with *Protium heptaphyllum* essential oil (Amaral *et al.*, 2009) which contains limonene as one of the major compounds. Another study has also reported that (+)-limonene epoxide reduces cell migration (de Almeida *et al.*, 2017). However, there are no reports in the literature concerning rotundifolone (a major constituent of EOLp) anti-inflammatory effect.

Peritonitis involves exudation and production of mediators such as nitric oxide and cytokines (IL-1 β and TNF- α) (Lima *et al.*, 2012). Thus, the effect of EOLp was evaluated from levels of IL-1 β and TNF- α in the peritoneal exudate of mice with carrageenan-induced peritonitis. EOLp reduced concentrations of IL-1 β , however TNF- α levels were not altered. The same has been reported for (+)-limonene epoxide (de Almeida *et al.*, 2017). Upon cell injury, infection, and inflammation, IL-1 β is released: primarily by monocytes, macrophages and mast cells, and as well as by non-immune cells, like neuronal and glial cells (Schwann cells, microglia, and astrocytes). Overproduction of IL-1 β is implicated in the pathophysiological changes which occur in inflammatory diseases (Zhang & An, 2007; Ren & Torres, 2009).

CONCLUSION

The results of this paper allow us to conclude that EOLp has better antinociceptive effect in mice when compared to EOLp/ β -CD complex. Thus, though CDs are efficient as drug delivery system with EOs inclusion complex, but not with EOs rich in peroxide substances. EOLp is a promising option for treating inflammation and pain-related disorders, significantly inhibiting acetic acid-induced writhing in both phases of formalin-induced nociception in mice, and in orofacial nociceptive behavior induced by formalin, glutamate, and capsaicin. Additionally, it reduced leucocyte migration to the peritoneal cavity, while

reducing IL-1 β levels, which together confirm its anti-inflammatory effect.

ACKNOWLEDGMENTS

The authors are grateful to the National Council for Scientific and Technological Development (CNPq) - Brazil for the financial support.

REFERENCES

- Abena AA, Diatwa M, Gakosso G, Gbeassor M, Hondi-Assah TH, Ouamba JM. 2003. Analgesic, antipyretic and anti-inflammatory effects of essential oil of *Lippia multiflora*. **Fitoterapia** 74: 231 - 236. [https://doi.org/10.1016/s0367-326x\(03\)00029-7](https://doi.org/10.1016/s0367-326x(03)00029-7)
- Ahmed F, Selim MST, Das AK, Choudhuri MSK. 2004. Anti-inflammatory and antinociceptive activities of *Lippia nodiflora* Linn. **Pharmazie** 59: 329 - 330.
- Amaral MP, Braga FA, Passos FF, Almeida FR, Oliveira RC, Carvalho AA, Chaves MH, Oliveira FA. 2009. Additional evidence for the anti-inflammatory properties of the essential oil of *Protium heptaphyllum* resin in mice and rats. **Latin Am J Pharm** 28: 775 - 782.
- Andrade TA, Freitas TS, Araújo FO, Menezes PP, Dória GAA, Rabelo AS, Quintans-Júnior LJ, Santos MRV, Bezerra DP, Serafini MR, Menezes IRA, Nunes PS, Araújo AAS, Costa MS, Campina FF, Santos ATL, Silva ARP, Coutinho HDM. 2017. Physico-chemical characterization and antibacterial activity of inclusion complexes of *Hyptis martiusii* Benth essential oil in β -cyclodextrin. **Biomed Pharmacother** 89: 201 - 207. <https://doi.org/10.1016/j.biopha.2017.01.158>
- Arana-Sánchez A, Estarrón-Espinosa M, Obledo-Vázquez EN, Padilla-Camberos E, Silva-Vázquez R, Lugo-Cervantes E. 2010. Antimicrobial and antioxidant activities of Mexican oregano essential oils (*Lippia graveolens* HBK) with different composition when microencapsulated in β -cyclodextrin. **Lett Appl Microbiol** 50: 585 - 590. <https://doi.org/10.1111/j.1472-765X.2010.02837.x>
- Araújo-Filho HG, Pereira EW, Rezende MM, Menezes PP, Araújo AA, Barreto RS, Martins AOBPB, Albuquerque TR, Silva BAF, Alcantara IS, Coutinho HDM, Menezes IRA, Quintans-Júnior LJ, Quintans JSS. 2017. D-limonene exhibits superior antihyperalgesic effects in a β -cyclodextrin-complexed form in chronic musculoskeletal pain reducing Fos protein expression on spinal cord in mice. **Neuroscience** 358: 158 - 169. <https://doi.org/10.1016/j.neuroscience.2017.06.037>
- Bonjardim LR, Silva AM, Oliveira MGB, Guimarães AG, Antonioli AR, Santana MF, Serafini MR, Santos RC, Araújo AAS, Estevam CS, Santos MRV, Lyra A, Carvalho R, Quintans-Júnior LJ, Azevedo EG, Botelho MA. 2011. *Sida cordifolia* leaf extract reduces the orofacial nociceptive response in mice. **Phytother Res** 25: 1236 - 1241. <https://doi.org/10.1002/ptr.3550>
- Brito RG, Araujo AA, Quintans JS, Sluka KA, Quintans-Junior LJ. 2015. Enhanced analgesic activity by cyclodextrins, a systematic review and meta-analysis. **Exp Opin Drug Deliver** 12: 1677 - 1688. <https://doi.org/10.1517/17425247.2015.1046835>
- Carvalho V, Fernandes L, Conde T, Zamith H, Silva R, Surrage A, Frutuoso V, Castro-Faria-Neto HA, Mendoeira F. 2013. Antinociceptive activity of *Stephanolepis hispidus* skin aqueous extract depends partly on opioid system activation. **Marine Drugs** 11: 1221 - 1234. <https://doi.org/10.3390/md11041221>
- Costa M, Di Stasi LC, Kirizawa M, Mendaçolli SL, Gomes C, Trolin G. 1989. Screening in mice of some medicinal plants used for analgesic purposes in the state of Sao Paulo. Part II. **J Ethnopharmacol** 27: 25 - 33. [https://doi.org/10.1016/0378-8741\(89\)90074-3](https://doi.org/10.1016/0378-8741(89)90074-3)
- Costa P, Medronho B, Gonçalves S, Romano A. 2015. Cyclodextrins enhance the antioxidant activity of essential oils from three Lamiaceae species. **Ind Crop Prod** 70: 341 - 346. <https://doi.org/10.1016/j.indcrop.2015.03.065>
- Damasceno MB, de Melo Júnior JM, Santos SA, Melo LT, Leite LH, Vieira-Neto AE, Moreira RA, Monteiro-Moreira AC, Campos AR. 2016. Frutalin reduces acute and neuropathic nociceptive behaviours in rodent models of orofacial pain. **Chem Biol Interact** 256: 9 - 15. <https://doi.org/10.1016/j.cbi.2016.06.016>
- de Almeida AA, Silva RO, Nicolau LA, de Brito TV, de Sousa DP, Barbosa AL, de Freitas RM, Lopes LD, Medeiros JR, Ferreira PM. 2017. Physio-pharmacological investigations about the anti-inflammatory and antinociceptive efficacy of (+)-limonene epoxide. **Inflammation** 40: 511 - 522. <https://doi.org/10.1007/s10753-016-0496-y>
- de Almeida RN, Motta SC, de Brito Faturi C, Catallani B, Leite JR. 2004. Anxiolytic-like effects of rose oil

- inhalation on the elevated plus-maze test in rats. **Pharmacol Biochem Behav** 77: 361 - 364.
<https://doi.org/10.1016/j.pbb.2003.11.004>
- de Amorim Santos IG, Scher R, Rott MB, Menezes LR, Costa EV, de Holanda Cavalcanti SC, Blank AF, Aguiar JS, da Silva TG, Dolabella SS. 2016. Amebicidal activity of the essential oils of *Lippia* spp. (Verbenaceae) against *Acanthamoeba polyphaga* trophozoites. **Parasitol Res** 115: 535 - 540.
<https://doi.org/10.1007/s00436-015-4769-4>
- de Oliveira MG, Guimarães AG, Araújo AA, Quintans JS, Santos MR, Quintans-Júnior LJ. 2015. Cyclodextrins: improving the therapeutic response of analgesic drugs: a patent review. **Expert Opin Ther Pat** 25: 897 - 907. <https://doi.org/10.1517/13543776.2015.1045412>
- De Sousa DP. 2011. Analgesic-like activity of essential oils constituents. **Molecules** 16: 2233 - 2252.
<https://doi.org/10.3390/molecules16032233>
- Deraedt R, Jouquey S, Delevallée F, Flahaut M. 1980. Release of prostaglandins E and F in an algogenic reaction and its inhibition. **Eur J Pharmacol** 61: 17 - 24. [https://doi.org/10.1016/0014-2999\(80\)90377-5](https://doi.org/10.1016/0014-2999(80)90377-5)
- do Amaral JF, Silva MIG, de Aquino Neto MRA, Neto PFT, Moura BA, de Melo CTV, Araújo FLO, de Sousa DP, Vasconcelos PF, Vasconcelos SM, de Sousa FCF. 2007. Antinociceptive effect of the monoterpene R-(+)-limonene in mice. **Biol Pharmaceut Bull** 30: 1217 - 1220. <https://doi.org/10.1248/bpb.30.1217>
- Dunham NW, Miya TS. 1957. A note on a simple apparatus for detecting neurological deficit in rats and mice. **J Am Pharmaceut Assoc** 46: 208 - 209.
- Eddy NB, Leimbach D. 1953. Synthetic analgesics. II. Dithienylbutenyl- and dithienylbutylamines. **J Pharmacol Exp Ther** 107: 385 - 393.
- Fernandes LP, Éhen Z, Moura TF, Novák C, Sztatisz J. 2004. Characterization of *Lippia sidoides* oil extract- β -cyclodextrin complexes using combined thermoanalytical techniques. Journal of thermal analysis and calorimetry. **J Thermal Anal Calorim** 78: 557 - 573.
<https://doi.org/10.1023/b:jtan.0000046119.33278.9f>
- Guilhon CC, Raymundo LJ, Alviano DS, Blank AF, Arrigoni-Blank MF, Matheus ME, Cavalcanti SC, Alviano CS, Fernandes PD. 2011. Characterisation of the anti-inflammatory and antinociceptive activities and the mechanism of the action of *Lippia gracilis* essential oil. **J Ethnopharmacol** 135: 406 - 413.
<https://doi.org/10.1016/j.jep.2011.03.032>
- Guimaraes AG, Gomes SV, Moraes VR, Nogueira PC, Ferreira AG, Blank AF, Santos AD, Viana MD, Silva GH, Júnior LJQ. 2012. Phytochemical characterization and antinociceptive effect of *Lippia gracilis* Schauer. **J Natural Med** 66: 428 - 434. <https://doi.org/10.1007/s11418-011-0601-3>
- Guimarães AG, Quintans JS, Quintans-Júnior LJ. 2013. Monoterpenes with analgesic activity, a systematic review. **Phytother Res** 27: 1 - 15. <https://doi.org/10.1002/ptr.4686>
- Henry JL, Yashpal K, Pitcher GM, Chabot JG, Coderre TJ. 1999. Evidence for tonic activation of NK-1 receptors during the second phase of the formalin test in the rat. **J Neurosci** 19: 6588 - 6598.
<https://doi.org/10.1523/jneurosci.19-15-06588.1999>
- Honda K, Kitagawa J, Sessle BJ, Kondo M, Tsuboi Y, Yonehara Y, Iwata K. 2008. Mechanisms involved in an increment of multimodal excitability of medullary and upper cervical dorsal horn neurons following cutaneous capsaicin treatment. **Mol Pain** 4: 59 - 70. <https://doi.org/10.1186/1744-8069-4-59>
- Hu JW, Fiorentino PM, Cairns BE, Sessle BJ. 2005. Capsaicin-induced inflammation within temporomandibular joint involves VR-1 receptor mechanisms. **Oral Biosci Med** 4: 241 - 248.
- Hunskar S, Hole K. 1987. The formalin test in mice: dissociation between inflammatory and non-inflammatory pain. **Pain** 30: 103 - 114. [https://doi.org/10.1016/0304-3959\(87\)90088-1](https://doi.org/10.1016/0304-3959(87)90088-1)
- Hwang YY, Shin DC, Nam YS, Cho BK. 2012. Characterization, stability, and pharmacokinetics of sibutramine/ β -cyclodextrin inclusion complex. **J Ind Enginee Chem** 18: 1412 - 1417.
- Koster R, Anderson M, De Beer EJ. 1959. Acetic acid for analgesic screening. **Fed Proceed** 18: 412 - 417.
- Lam DK, Sessle BJ, Hu JW. 2009. Glutamate and capsaicin effects on trigeminal nociception II: activation and central sensitization in brainstem neurons with deep craniofacial afferent input. **Brain Res** 1253: 48 - 59.
<https://doi.org/10.1016/j.brainres.2008.11.056>
- Le Bars D, Gozariu M, Cadden SW. 2001. Animal models of nociception. **Pharmacol Rev** 53: 597 - 652.
- Lenardão EJ, Savegnago L, Jacob RG, Victoria FN, Martinez DM. 2016. Antinociceptive effect of essential oils and their constituents: an update review. **J Braz Chem Soc** 27: 435 - 474.

- <https://doi.org/10.5935/0103-5053.20150332>
Li J & Loh XJ. 2008. Cyclodextrin-based supramolecular architectures: syntheses, structures, and applications for drug and gene delivery. **Adv Drug Deliver Rev** 60: 1000 - 1017.
<https://doi.org/10.1016/j.addr.2008.02.011>
- Lima DF, Brandão MS, Moura JB, Leitão JM, Carvalho FA, Miúra LM, Leite JR, Sousa DP, Almeida FR. 2012. Antinociceptive activity of the monoterpene α -phellandrene in rodents: possible mechanisms of action. **J Pharm Pharmacol** 64: 283 - 292. <https://doi.org/10.1111/j.2042-7158.2011.01401.x>
- Lima PS, Lucchese AM, Araujo-Filho HG, Menezes PP, Araujo AA, Quintans-Junior LJ, Quintans JS. 2016. Inclusion of terpenes in cyclodextrins: Preparation, characterization and pharmacological approaches. **Carbohydr Polym** 151: 965 - 987. <https://doi.org/10.1016/j.carbpol.2016.06.040>
- Luccarini P, Childeric A, Gaydier AM, Voisin D, Dallel R. 2006. The orofacial formalin test in the mouse: a behavioral model for studying physiology and modulation of trigeminal nociception. **J Pain** 7: 908 - 914.
<https://doi.org/10.1016/j.jpain.2006.04.010>
- Ma F, Zhang L, Westlund KN. 2009. Reactive oxygen species mediate TNFR1 increase after TRPV1 activation in mouse DRG neurons. **Mol Pain** 5: 1744 - 8069. <https://doi.org/10.1186/1744-8069-5-31>
- Mansouri MT, Naghizadeh B, Ghorbanzadeh B. 2014. Involvement of opioid receptors in the systemic and peripheral antinociceptive actions of ellagic acid in the rat formalin test. **Pharmacol Biochem Behav** 120: 43 - 49. <https://doi.org/10.1016/j.pbb.2014.02.009>
- Marreto RN, Almeida EE, Alves PB, Niculau ES, Nunes RS, Matos CR, Araújo AA. 2008. Thermal analysis and gas chromatography coupled mass spectrometry analyses of hydroxypropyl- β -cyclodextrin inclusion complex containing *Lippia gracilis* essential oil. **Thermochim Acta** 475: 53 - 58.
<https://doi.org/10.1016/j.tca.2008.06.015>
- McNamara CR, Mandel-Brehm J, Bautista DM, Siemens J, Deranian KL, Zhao M, Hayward NJ, Chong JA, Julius D, Moran MM, Fanger CM. 2007. TRPA1 mediates formalin-induced pain. **Proc Nat Acad Sci** 104: 13525 - 13530. <https://doi.org/10.1073/pnas.0705924104>
- Melo LT, Duailibe MAB, Pessoa LM, da Costa FN, Vieira-Neto AE, de Vasconcellos Abdon AP, Campos AR. 2017. (-)- α -Bisabolol reduces orofacial nociceptive behavior in rodents. **Naunyn-Schmiedeberg's Arch Pharmacol** 390: 187 - 195. <https://doi.org/10.1007/s00210-016-1319-2>
- Mendes SS, Bomfim RR, Jesus HCR, Alves PB, Blank AF, Estevam CS, Antonioli AR, Thomazzi SM. 2010. Evaluation of the analgesic and anti-inflammatory effects of the essential oil of *Lippia gracilis* leaves. **J Ethnopharmacol** 129: 391 - 397. <https://doi.org/10.1016/j.jep.2010.04.005>
- Menezes LRA, Santos NN, Meira CS, dos Santos JAF, Guimarães ET, Soares MBP, Nepel A, Barisone A, Costa EV. 2014. A new source of (R)-limonene and rotundifolone from leaves of *Lippia pedunculosa* (Verbenaceae) and their trypanocidal properties. **Nat Prod Commun** 9: 737 - 739.
<https://doi.org/10.1177/1934578x1400900601>
- Menezes PP, Araujo AA, Doria GA, Quintans-Junior LJ, de Oliveira MG, dos Santos MR, de Oliveira JF, Matos JR, Carvalho FM, Alves PB, de Matos IL, dos Santos DA, Marreto RN, da Silva GF, Serafini MR. 2015. Physicochemical characterization and analgesic effect of inclusion complexes of essential oil from *Hyptis pectinata* L. Poit leaves with β -cyclodextrin. **Curr Pharmaceut Biotechnol** 16: 440 - 450.
<https://doi.org/10.2174/1389201015666141202101909>
- Menezes PP, Araujo FO, Andrade TA, Trindade IAS, Filho HGA, Quintans JSS, Quintans-Junior LJ, Menezes LRA, de Almeida RN, Braga RM, Serafini MR, Costa EV, Araujo AAS. 2018. Physicochemical characterization and antinociceptive effect of β -cyclodextrin/*Lippia pedunculosa* essential oil in mice. **Curr Topics Med Chem** 18: 797 - 807. <https://doi.org/10.2174/1568026618666180607081742>
- Millan MJ. 1999. The induction of pain: an integrative review. **Progress Neurobiol** 57: 1 - 164.
- Mohamad AS, Akhtar MN, Zakaria ZA, Perimal EK, Khalid S, Mohd PA, Khalid MH, Israf DA, Lajis NH, Sulaimana MR. 2010. Antinociceptive activity of a synthetic chalcone, flavokawin B on chemical and thermal models of nociception in mice. **Eur J Pharmacol** 647: 103 - 109.
<https://doi.org/10.1016/j.ejphar.2010.08.030>
- Nascimento AMD, Maia TDS, Soares TES, Menezes IRA, Scher R, Costa EV, Cavalcanti SC, La Corte R. 2017. Repellency and larvicidal activity of essential oils from *Xylopi laevigata*, *Xylopi frutescens*, *Lippia pedunculosa*, and their individual compounds against *Aedes aegypti* Linnaeus. **Neotropical Entomol** 46:

- 223 - 230. <https://doi.org/10.1007/s13744-016-0457-z>
- Nascimento SS, Araújo AA, Brito RG, Serafini MR, Menezes PP, DeSantana JM, Lucca W, Alves PB, Blank AF, Oliveira RC, Oliveira AP, Albuquerque RL, Almeida JR, Quintans LJ. 2015. Cyclodextrin-complexed *Ocimum basilicum* leaves essential oil increases Fos protein expression in the central nervous system and produce an antihyperalgesic effect in animal models for fibromyalgia. **Int J Mol Sci** 16: 547 - 563. <https://doi.org/10.3390/ijms16010547>
- Pang MH, Kim Y, Jung KW, Cho S, Lee DH. 2012. A series of case studies: practical methodology for identifying antinociceptive multi-target drugs. **Drug Discov Today** 17: 425 - 434. <https://doi.org/10.1016/j.drudis.2012.01.003>
- Pascual ME, Slowing K, Carretero E, Mata DS, Villar A. 2001. *Lippia*: traditional uses, chemistry and pharmacology: a review. **J Ethnopharmacol** 76: 201 - 214. [https://doi.org/10.1016/s0378-8741\(01\)00234-3](https://doi.org/10.1016/s0378-8741(01)00234-3)
- Pelissier T, Pajot J, Dallel R. 2002. The orofacial capsaicin test in rats: effects of different capsaicin concentrations and morphine. **Pain** 96: 81 - 87. [https://doi.org/10.1016/s0304-3959\(01\)00432-8](https://doi.org/10.1016/s0304-3959(01)00432-8)
- Pereira SS, Lopes LS, Marques RB, Figueiredo KA, Costa DA, Chaves MH, Almeida FRC. 2010. Antinociceptive effect of *Zanthoxylum rhoifolium* Lam. (Rutaceae) in models of acute pain in rodents. **J Ethnopharmacol** 129: 227 - 231. <https://doi.org/10.1016/j.jep.2010.03.009>
- Pinho E, Grootveld M, Soares G, Henriques M. 2014. Cyclodextrins as encapsulation agents for plant bioactive compounds. **Carbohydrate Polym** 101: 121 - 135. <https://doi.org/10.1016/j.carbpol.2013.08.078>
- Quintans-Júnior LJ, Melo MS, de Sousa DP, Araújo AAS, Onofre AC, Gelain DP, Gonçalves JC, Araújo DA, Almeida JR, Bonjardim LR. 2010. Antinociceptive effects of citronellal in formalin-, capsaicin-, and glutamate-induced orofacial nociception in rodents and its action on nerve excitability. **J Orofac Pain** 24: 305 - 312.
- Quintans-Júnior LJ, Barreto RS, Menezes PP, Almeida JR, Viana AFS, Oliveira RC, Oliveira AP, Gelain DP, de Lucca Júnior W, Araújo AA. 2013. β -Cyclodextrin-complexed (-)-linalool produces antinociceptive effect superior to that of (-)-linalool in experimental pain protocols. **Basic Clin Pharmacol Toxicol** 113: 167 - 172. <https://doi.org/10.1111/bcpt.12087>
- Ren K, Torres R. 2009. Role of interleukin-1 β during pain and inflammation. **Brain Res Rev** 60: 57 - 64. <https://doi.org/10.1016/j.brainresrev.2008.12.020>
- Ribas CM, Meotti FC, Nascimento FP, Jacques AV, Dafre AL, Rodrigues ALS, Farina M, Soldi C, Mendes BG, Pizzolatti MG, Santos ARS. 2008. Antinociceptive effect of the *Polygala sabulosa* hydroalcoholic extract in mice: evidence for the involvement of glutamatergic receptors and cytokine pathways. **Basic Clin Pharmacol Toxicol** 103: 43 - 47. <https://doi.org/10.1111/j.1742-7843.2008.00245.x>
- Ro JY, Chung MK, Lee JS, Saloman JL, Joseph J. 2014. Functional interactions between glutamate receptors and TRPV1 in trigeminal sensory neurons. **Mol Pain** 10: O13. <https://doi.org/10.1186/1744-8069-10-S1-O13>
- Rodrigues LB, Martins AOBPB, Ribeiro-Filho J, Cesário FRAS, Castro FF, de Albuquerque TR, Fernandes MNM, da Silva BAF, Quintans Júnior LJ, Araújo AAS, Menezes PP, Nunes PS, Matos IG, Coutinho HDM, Wanderley AG, Menezes IRA. 2017. Anti-inflammatory activity of the essential oil obtained from *Ocimum basilicum* complexed with β -cyclodextrin (β -CD) in mice. **Food Chem Toxicol** 109: 836 - 846. <https://doi.org/10.1016/j.fct.2017.02.027>
- Sakurada T, Matsumura T, Moriyama T, Sakurada C, Ueno S, Sakurada S. 2003. Differential effects of intraplantar capsazepine and ruthenium red on capsaicin-induced desensitization in mice. **Pharmacol Biochem Behav** 75: 115 - 121. [https://doi.org/10.1016/s0091-3057\(03\)00066-2](https://doi.org/10.1016/s0091-3057(03)00066-2)
- Santos JSD, Melo JIMD, Abreu MCD, Sales MFD. 2009. Verbenaceae sensu stricto na região de Xingó: Alagoas e Sergipe, Brasil. **Rodriguésia** 60: 985 - 998. <https://doi.org/10.1590/2175-7860200960412>
- Santos NN, Menezes LR, Meira CS, Guimarães ET, Soares MB, Nepel A, Barisone A, Costa EV. 2014. A new source of (R)-limonene and rotundifolone from leaves of *Lippia pedunculosa* (Verbenaceae) and their trypanocidal properties. **Nat Prod Commun** 9: 737 - 739. <https://doi.org/10.1177/1934578x1400900601>
- Santos PL, Araújo AA, Quintans JS, Oliveira MG, Brito RG, Serafini MR, Menezes PP, Santos MR, Alves PB, de Lucca Júnior W, Blank AF, La Rocca V, Almeida RN, Quintans-Júnior LJ. 2015. Preparation, characterization, and pharmacological activity of *Cymbopogon winterianus* Jowitt ex Bor (Poaceae) leaf essential oil of β -cyclodextrin inclusion complexes. **Evid-Based Compl Alt Med** 2015: 502454.

- <https://doi.org/10.1155/2015/502454>
- Santos PL, Brito RG, Quintans JSS, Araujo AAS, Menezes IRA, Brogden NK, Quintans-Junior LJ. 2017. Cyclodextrins as complexation agents to improve the anti-inflammatory drugs profile: a systematic review and meta-analysis. **Current Pharmaceut Design** 23: 2096 - 2107. <https://doi.org/10.2174/1381612823666170126121926>
- Silva ACDC, Prata APDN, Mello AAD. 2013. Flowering plants of the Grota do Angico Natural Monument, Caatinga of Sergipe, Brazil. **Check List** 9: 733 - 739. <https://doi.org/10.15560/9.4.733>
- Silva JC, Almeida JR, Quintans JS, Gopalsamy RG, Shanmugam S, Serafini MR, Oliveira MRC, Silva BAF, Martins AOBPB, Castro FF, Menezes IRA, Coutinho HDM, Oliveira RCM, Thangaraj P, Araújo AAS, Quintans-Júnior LJ. 2016. Enhancement of orofacial antinociceptive effect of carvacrol, a monoterpene present in oregano and thyme oils, by β -cyclodextrin inclusion complex in mice. **Biomed Pharmacother** 84: 454 - 461. <https://doi.org/10.1016/j.biopha.2016.09.065>
- Siqueira-Lima PS, Araújo AA, Lucchese AM, Quintans JS, Menezes PP, Alves PB, de Lucca W, Santos MR, Bonjardim LR, Quintans-Júnior LJ. 2014. β -cyclodextrin complex containing *Lippia grata* leaf essential oil reduces orofacial nociception in mice—evidence of possible involvement of descending inhibitory pain modulation pathway. **Basic Clin Pharmacol Toxicol** 114: 188 - 196. <https://doi.org/10.1111/bcpt.12145>
- Siqueira-Lima PS, Silva JC, Quintans JS, Antonioli AR, Shanmugam S, Barreto RS, Santos MRV, Almeida JRGS, Bonjardim LR, Menezes IRA, Quintans-Júnior LJ. 2017. Natural products assessed in animal models for orofacial pain, a systematic review. **Rev Bras Farmacogn** 27: 124 - 134. <https://doi.org/10.1016/j.bjp.2016.06.005>
- Sousa PJDC, Linard CFBM, Azevedo-Batista D, Oliveira AC, Coelho-de-Souza AN, Leal-Cardoso JH. 2009. Antinociceptive effects of the essential oil of *Mentha x villosa* leaf and its major constituent piperitenone oxide in mice. **Braz J Med Biol Res** 42: 655 - 659. <https://doi.org/10.1590/s0100-879x2009000700010>
- Terblanché FC, Kornelius G. 1996. Essential oil constituents of the genus *Lippia* (Verbenaceae), a literature review. **J Essent Oil Res** 8: 471 - 485. <https://doi.org/10.1080/10412905.1996.9700673>
- Tjølsen A, Berge OG, Hunskaar S, Rosland JH, Hole K. 1992. The formalin test: an evaluation of the method. **Pain** 51: 5 - 17. [https://doi.org/10.1016/0304-3959\(92\)90003-t](https://doi.org/10.1016/0304-3959(92)90003-t)
- Tomaz-Morais JF, Braga RM, de Sousa FB, de Sousa DP, Pordeus LD, de Almeida RN, de Castro RD. 2017. Orofacial antinociceptive activity of (S)-(-)-perillyl alcohol in mice: a randomized, controlled and triple-blind study. **Int J Oral Maxillofacial Surg** 46: 662 - 667. <https://doi.org/10.1016/j.ijom.2017.01.024>
- Viana GS, do Vale TG, Rao VSN, Matos FJA. 1998. Analgesic and antiinflammatory effects of two chemotypes of *Lippia alba*: a comparative study. **Pharmaceut Biol** 36: 347 - 351. <https://doi.org/10.1076/phbi.36.5.347.4646>
- Waning J, Vriens J, Owsianik G, Stüwe L, Mally S, Fabian A, Frippiat C, Nilius B, Schwab A. 2007. A novel function of capsaicin-sensitive TRPV1 channels: involvement in cell migration. **Cell Calcium** 42: 17 - 25. <https://doi.org/10.1016/j.ceca.2006.11.005>
- Yang KY, Mun JH, Park KD, Kim MJ, Ju JS, Kim ST, Bae YC, Ahn DK. 2015. Blockade of spinal glutamate recycling produces paradoxical antinociception in rats with orofacial inflammatory pain. **Progress Neuro-Psychopharmacol Biol Psychiatry** 57: 100 - 109. <https://doi.org/10.1016/j.pnpbp.2014.10.011>
- Yeung JC, Yaksh TL, Rudy TA. 1977. Concurrent mapping of brain sites for sensitivity to the direct application of morphine and focal electrical stimulation in the production of antinociception in the rat. **Pain** 4: 23 - 40. [https://doi.org/10.1016/0304-3959\(77\)90084-7](https://doi.org/10.1016/0304-3959(77)90084-7)
- Yi J, Zheng JY, Zhang W, Wang S, Yang ZF, Dou KF. 2014. Decreased pain threshold and enhanced synaptic transmission in the anterior cingulate cortex of experimental hypothyroidism mice. **Mol Pain** 10: 1744 - 8069. <https://doi.org/10.1186/1744-8069-10-38>
- Zhang JM, An J. 2007. Cytokines, inflammation and pain. **Int Anesthesiol Clin** 45: 27.