

Revisión / Review

Plants and intestinal parasitosis: a review on ethnopharmacological use by the Kantaruré-Batida indigenous community of Brazil

[Plantas y parasitosis intestinal: una revisión sobre el uso etnofarmacológico de la comunidad indígena Kantaruré-Batida de Brasil]

Deyvison Rhuan Vasco-dos-Santos¹, Jéssica Vieira dos Santos², Maria José Gomes de Andrade², Lidiane Nunes Lima³, Wbaneide Martins de Andrade³, Erika dos Santos Nunes⁴, Artur Gomes Dias-Lima³ & Marcos André Vannier-Santos¹

¹Programa de Pós-Graduação em Biologia Parasitária, Laboratório de Inovações em Terapias, Ensino e Bioprodutos, Instituto Oswaldo Cruz/FIOCRUZ, Rio de Janeiro, Brazil

²Programa de Pós-Graduação em Biodiversidade Vegetal, Departamento de Educação, Universidade do Estado da Bahia, *Campus* VIII, Paulo Afonso, Bahia, Brazil

³Programa de Pós-Graduação em Ecologia Humana e Gestão Socioambiental, Departamento de Tecnologias e Ciências Sociais, Universidade do Estado da Bahia, *Campus* III, Juazeiro, Bahia, Brazil

⁴Laboratório de Estudos em Parasitologia Humana, Departamento de Educação, Universidade do Estado da Bahia, *Campus* VIII, Paulo Afonso, Bahia, Brazil

Reviewed by:

Rosa Degen
Universidad Nacional de Asunción
Paraguay

Elsa Rengifo

Instituto de Investigaciones de la Amazonía Peruana
Peru

Correspondence:

Deyvison Rhuan Vasco-dos-SANTOS:
deyvisonrvs@gmail.com

Section Review

Received: 8 May 2021

Accepted: 1 August 2021

Accepted corrected: 21 October 2021

Published: 30 May 2022

Citation:

Vasco-dos-Santos DR, Santos JV, Andrade MJG, Lima LN, Andrade WM, Nunes ES, Dias-Lima AG, Vannier-Santos MAV.

Plants and intestinal parasitosis: a review on ethnopharmacological use by the Kantaruré-Batida indigenous community of Brazil

Bol Latinoam Caribe Plant Med Aromat

21 (3): 268 - 308 (2022).

<https://doi.org/10.37360/blacpma.22.21.3.17>

Abstract: Indigenous knowledge is one of the most notable traditional sources about plants used to treat diseases. Thus, the aim of this study was to describe the botanical features and to investigate the pharmacological properties of plants used by the Kantaruré-Batida community to treat intestinal parasitosis. Botanical characterization was performed based on integrative review and on access to 'Flora do Brasil' platform, whereas plants' pharmacological properties were investigated through systematic review. Among the 21 ethnospecies used, 48% were described as having activity against intestinal parasites. Although 52% had no such activity described, other properties can account for their use, that indicates that further studies are required. Therefore, this review reinforces the importance of valuing indigenous knowledge as tool to guide antiparasitic agent trials.

Keywords: Ethnobotany; Medicinal plants of the Caatinga; Indigenous health; Ethnoparasitology; Bibliometric analysis.

Resumen: El conocimiento indígena es una de las fuentes tradicionales más notables sobre las plantas utilizadas para tratar enfermedades. Por lo tanto, el objetivo de este estudio fue describir las características botánicas e investigar las propiedades farmacológicas de las plantas utilizadas por la comunidad Kantaruré-Batida para tratar la parasitosis intestinal. La caracterización botánica se realizó con base en una revisión integradora y en el acceso a la plataforma "Flora do Brasil", mientras que las propiedades farmacológicas de las plantas se investigaron mediante una revisión sistemática. Entre las 21 etnoespecies utilizadas, se describió que el 48% tenía actividad contra los parásitos intestinales. Aunque el 52% no tenía tal actividad descrita, otras propiedades pueden explicar su uso, lo que indica que se requieren más estudios. Por lo tanto, esta revisión refuerza la importancia de valorar el conocimiento indígena como herramienta para guiar los ensayos de agentes antiparasitarios.

Palabras clave: Etnobotánica; Plantas medicinales de la Caatinga; Salud indígena; Etnoparasitología; Análisis bibliométrico.

INTRODUCTION

Quinine and its many derivatives, such as hydroxychloroquine, are among the drugs that saved more lives in human history. These drugs derived from the ethnobotanical knowledge of indigenous communities living in the Amazon region (Achan *et al.*, 2011; Renslo, 2013; Luciano, 2015; Jomsky & Kerna, 2020). This knowledge was passed on to the church and generated intense commercial activity through more than 100 joint ventures focused on trading this product. The trading process was coordinated by the Dutch pharmaceutical consortium of Cinchona producers, Cinchona Bureau, between the 1920s and 1930s (Van Der Hoogte & Pieters, 2016), and it generated considerable economic power.

Also, the relevance of ethnopharmacology for human health can be unequivocally verified by the discovery of the antimalarial activity of the *Artemisia annua* L. (Sadiq *et al.*, 2014), for which the Chinese researcher Youyou Tu was awarded the Nobel Prize in 2015 (Chen, 2016). Nowadays, the repositioning of antimalarial drugs has revealed the anti-cancer activity of hydroxychloroquine (Amaravadi *et al.*, 2019; Liu *et al.*, 2020) and *Artemisia* L. compounds (Taleghani *et al.*, 2020). Therefore, plants used by native communities can guide the discovery of new compounds potentially employed for treating different disorders (Diehl, 2013; Araújo *et al.*, 2016).

Informations about which and how medicinal species are used by indigenous communities has aroused the interest of the world's scientific community, as observed in studies conducted in China (Chang *et al.*, 2017), Pakistan (Aziz *et al.*, 2018), Nigeria (Taiwo *et al.*, 2018), Madagascar (Riondato *et al.*, 2019), Saudi Arabia (Tounekti *et al.*, 2019), Ethiopia (Abebe & Garedew, 2019), Argentina (Carabajal *et al.*, 2020), Uganda (Anywar *et al.*, 2020), India (Rana *et al.*, 2021), Italy (Mattalia *et al.*, 2021) and Spain (Tardío *et al.*, 2021).

According to the World Health Organization (2020), traditional indigenous medicine results from knowledge, attitudes and practices exercised in therapeutic procedures to cure different diseases, based on individuals' beliefs and experiences. Brazil has the largest number of publications on the subject among Latin American countries. It happens not only due to the number of researchers, but also to the cultural and plant diversity linked to wide edaphic, climatic and geomorphological variations (Rapini *et*

al., 2009; Oliveira *et al.*, 2009; Albuquerque *et al.*, 2013a; Silva *et al.*, 2015a). The relevance of herbal medicine use in the country has led to the development of the National List of Medicinal Plants of Interest to the Unified Health System^a (Ministry of Health, 2009), which included 71 plant species and estimated that 2,000 species are currently used. The high use rate is mainly attributed to the indigenous culture (Sousa *et al.*, 2008).

Investigations about plants used by native communities in Brazil have revealed promising properties. Oliveira *et al.* (2007), observed mycobactericidal activity in plants deriving from Vanuïre pharmacopoeia (São Paulo State); Moura-Costa *et al.* (2012), reported antifungal, antiviral, leishmanicidal activity and cytotoxic effect of species used by indigenous communities living in Rio das Cobras reserve (Paraná State); Brandelli *et al.* (2013), have explained the action of plants used by Mbyá-Guarani communities (Rio Grande do Sul State) against *Trichomonas vaginalis* Donn  1836; Aquino *et al.* (2017), have partly confirmed the diuretic activity of *Alibertia edulis* (Rich.) A. Rich. ex DC.; and Tirloni *et al.* (2018) reported antioxidant activity and high bioactive compounds extracted from *A. edulis*.

Despite housing 25.5% of the national indigenous population (IBGE, 2010), Northeastern Brazil lacks studies on the knowledge of these communities, mainly about the treatment of infections caused by worms, which comprise major causes of morbidity and mortality associated with socio-sanitary vulnerability (Albuquerque *et al.*, 2007; FUNAI, 2010; Basta *et al.*, 2012; Vasco-dos-Santos *et al.*, 2020). According to Hossain (2018), it is necessary to encourage research about the flora used in traditional systems in order to find new anthelmintics, consequently increasing the scientific interest in ethnopharmacology studies. However, studies conducted in this field in recent years have focused on finding specific properties and on synthesizing new drugs, there has been no record of studies about the flora used in traditional medicine (Yeung *et al.*, 2020).

This study took into consideration the relevance of indigenous knowledge, its demand for research, dissemination and valorization, as well as

^a Relação Nacional de Plantas Medicinais de Interesse ao SUS – RENISUS.

health issues caused by parasitic diseases. Thus, the aim of the present research was to review the botanical characterization and pharmacological properties of plant species used by the Kantaruré-Batida indigenous community to treat intestinal parasites.

MATERIALS AND METHODS

The Kantaruré-Batida territory is located in Glória County, Bahia State (Figure No. 1). It presents semi-arid climate, low rainfall rates and typical caatinga vegetation. Fishing, hunting and plant extraction, mainly of cassava, corn and beans, are the basis of their subsistence (Santos, 2008; ISA, 2018). Community's use of natural resources has low environmental impact, and it is not restricted to food; it includes the use of over 100 fauna and flora species to treat different diseases (Lima, 2014; Lima *et al.*,

2016).

Plants indicated by the Kantaruré community to treat intestinal parasitosis were previously reported (Vasco-dos-Santos *et al.*, 2018). Botanical characterization was carried out based on integrative review (Whittemore & Knafl, 2005) and on Flora do Brasil platform (BFG, 2020), in order to access taxonomic, anatomical and geographical distribution information. Systematic review (Khan *et al.*, 2003) was performed to investigate plants' pharmacological properties by using the scientific name of each species as descriptors in the Science Direct, PubMed and Google Scholar databases. The first 500 titles of studies published in these databases until 2016, without language limitations, were checked, whenever available. Thus, making a search plan of 1,500 studies per species and 31,500, in total.

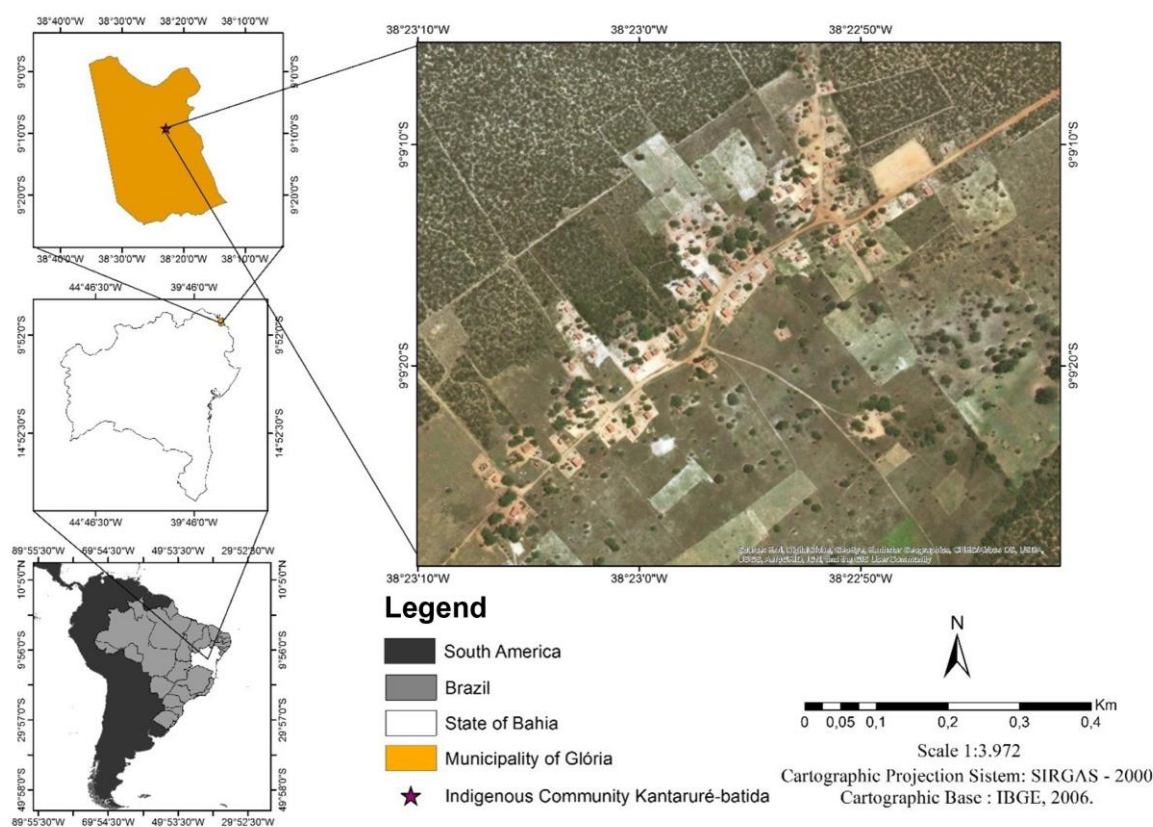


Figure No. 1
Location of the Kantaruré-Batida indigenous community, Bahia State, Brazil

The time frame of systematic review is justified by the period when the ethnobotanical survey was carried out, by taking into consideration

that, likewise biotechnological and pharmaceutical advances, the group of plants composing the pharmacopeia of a given culture is always dynamic,

as reported in the theory of cultural evolution (Mesoudi, 2007; Araújo *et al.*, 2016). Therefore, the time of access to indigenous and pharmaceutical knowledge was standardized. Selective criterion only comprised studies with significant pharmacological properties. Consequently, other research types, such as physiological and ecological studies were disregarded. The most recent research was taken into consideration whenever similar properties were observed for the same species in different studies, except for the ones about intestinal parasites.

The analysis of the collected information was organized into three categories, namely: I) Botanical characterization; II) Diverse pharmacological properties and III) Activities on intestinal parasites. Data deriving from the first category were compiled in order to optimize and detail plants' description, whereas the second category only took into consideration plants' demonstrated medicinal action.

The last category included data extraction process adapted from Sousa *et al.* (2013), it took into consideration the following information: a) plant species/reference; b) used part; c) extract type; d) concentration/dose; e) assay type; f) infected animal/fecal collection and g) tested parasites.

RESULTS

The systematic review strategy adopted in the current study resulted in 334 studies reporting plants pharmacological activity. Among these, 140 were excluded from the sample because they repeated properties for some plant species, except for studies about intestinal parasites. Thus, 194 studies were included in the final analysis (Figure No. 2). The botanical characterization and pharmacological properties observed for each plant are presented below.

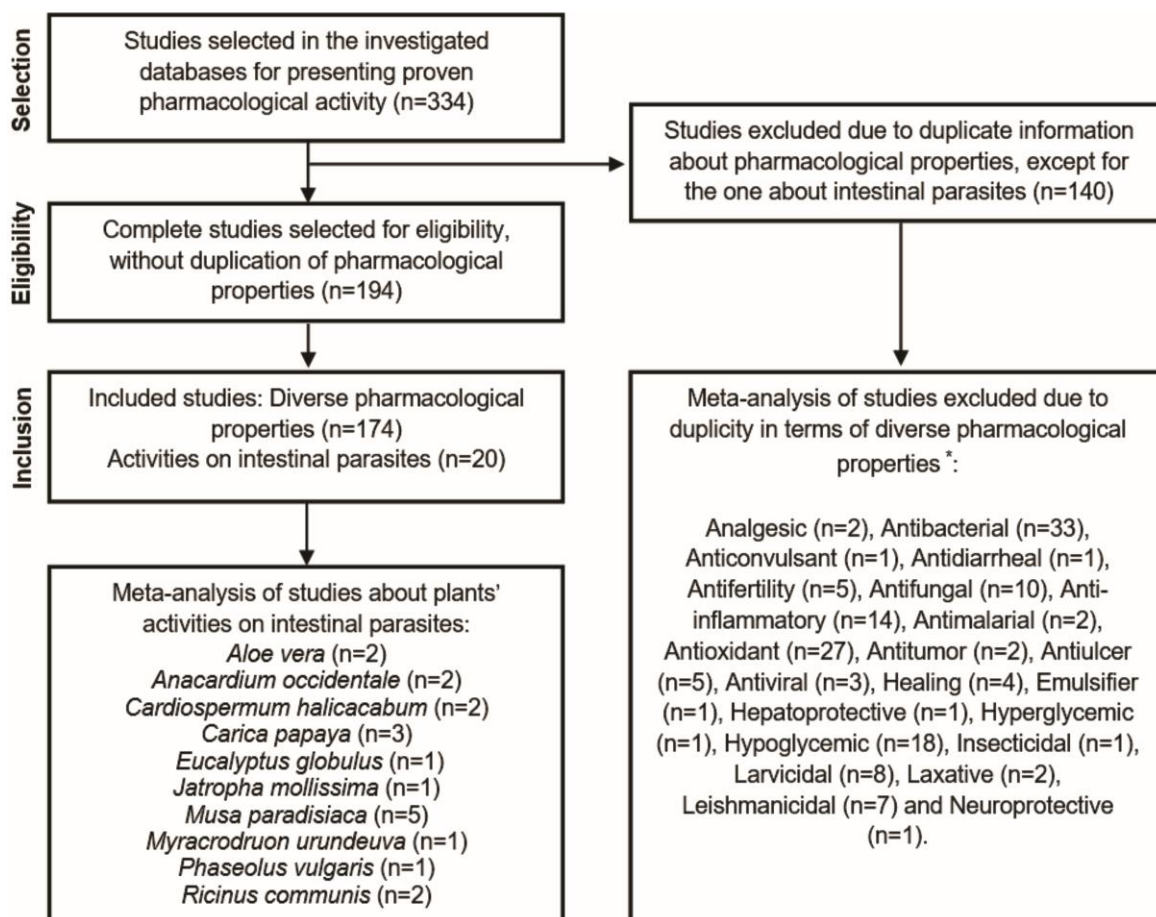


Figure No. 2

Flowchart of systematic review about plants' pharmacological properties used by Kantaruré-Batida indigenous, Bahia State, Brazil. *Quantification based on properties

ALECRIM**Botanical characterization**

Lippia thymoides Mart. & Schauer (Verbenaceae) is a shrub species that reaches up to 1.5 m in height. It presents simple, verticillate, membranaceous leaves, oblanceolate-to-spatulate blade with hyphenated venation; helical axillary summit inflorescence, flowers with white-to-orange-yellow corolla and schizocarp fruit. The species is endemic to Brazil; it naturally grows in the Northeastern (Bahia, Pernambuco, Rio Grande do Norte and Sergipe) and Southeastern regions (Minas Gerais), in phytogeographic domains such as Caatinga and Cerrado (Melo *et al.*, 2010a; BFG, 2020).

Diverse pharmacological properties

Antiseptic (Pinto *et al.*, 2013).

Activities on intestinal parasites

Not reported.

ALGODÃO (Figure No. 3B)**Botanical characterization**

Gossypium hirsutum L. (Malvaceae) it is a subshrub or shrub that reaches ca. 1.5 m in height. It presents simple, alternate, cartaceous leaves; 3-lobed or partly 5-lobed blade and reticulated venation; overall solitary flowers, yellow corolla and vinaceous base, androphore, as well as capsule fruit with white kapok and blackened seeds. Reported to be originated in both Jamaica and in warm South American regions. The taxon was introduced in Brazil and it can be seen in all five regions of the country in phytogeographic domains such as the Amazon, Caatinga, Cerrado and Atlantic Forest (Watt, 1907; Bayer & Kubitzki, 2003; BFG, 2020).

Diverse pharmacological properties

Larvicidal (Patil *et al.*, 2014).

Activities on intestinal parasites

Not reported.

AMEIXA DO MATO (Figure No. 3C)**Botanical characterization**

Ximenia americana L. (Olacaceae) is a shrub or small tree species that reaches 3-4 m in height. It presents zigzag thorny branches; simple, alternate, cartaceous leaves; yellow-white aromatic flowers with curved petals arranged in short, axillary or terminal racemes;

and drupe fruits. The species is often found in Africa, India, New Zealand, Central and South America. It can be found in dry and open forests, in rocky and riverside slopes, as well as in coastal forests. The species is native to Brazil; it grows in all five regions in phytogeographic domains such as the Amazon, Caatinga, Cerrado and Atlantic Forest (Sacande & Voutier, 2006; Brasileiro *et al.*, 2008; BFG, 2020).

Diverse pharmacological properties

Antibacterial (Adamu *et al.*, 2005), anticonvulsant (Quintans-Júnior *et al.*, 2002), antifungal (Traoré *et al.*, 2015), anti-inflammatory (Shettar *et al.*, 2015), antineoplastic (Voss *et al.*, 2006), antioxidant (Almeida *et al.*, 2016), antipyretic (Soro *et al.*, 2009) and hepatoprotective (Jayaveera *et al.*, 2009).

Activities on intestinal parasites

Not reported.

AROEIRA**Botanical characterization**

Myracrodruon urundeuva Allemão (Anacardiaceae) is a tree species that reaches 5-30 m in height. It presents aromatic, composed, alternate, imparipinnate leaves with 6 to 14 opposite leaflets; light yellow or green female flowers arranged in terminal or axillary panicles and male purple flowers arranged in hanging panicles, persistent sepals; dark-brown drupe fruits with rough surface. The species is native to Brazil and widely distributed in the Northern, Northeastern, Midwestern, Southeastern and Southern regions in phytogeographic domains such as Caatinga, Cerrado, Atlantic Forest, Pampa and Pantanal (Siqueira Filho *et al.*, 2009; BFG, 2020).

Diverse pharmacological properties

Analgesic (Viana *et al.*, 2003), antiadherent (Alves *et al.*, 2009), antibacterial (Jandú *et al.*, 2013), antidiarrheal (Chaves *et al.*, 1998), anti-inflammatory (Machado *et al.*, 2012), antioxidant (Vieira *et al.*, 2015), antiulcer (Carlini *et al.*, 2010), antiviral (Cecílio *et al.*, 2016), insecticidal (Napoleão *et al.*, 2013), larvicidal (Souza *et al.*, 2015), neuroprotective (Calou *et al.*, 2014) and wound healing (Mello *et al.*, 2013).

Activities on intestinal parasites

Anthelmintic (Oliveira *et al.*, 2011).

BABOSA (Figure No. 3D)**Botanical characterization**

Aloe vera (L.) Burm.f. (Asparagaceae) is an herbal species that reaches 30-60 cm in height. It presents short and stoloniferous stem; rosette, erect, ensiform and fleshy leaves; central racemose upright inflorescence that reaches 1 to 1.50 m in height; tubular yellowish-white flowers and capsule-type fruits. The species is mainly grown in tropical and subtropical regions worldwide and it likely has its origin in the African Mediterranean region (Dimitri, 1978; Teske & Trentini, 1994). In addition, it grows in anthropic areas of all five regions in Brazil (BFG, 2020).

Diverse pharmacological properties

Anti-acne (Orafidiya *et al.*, 2004), antiasthmatic (Rivas *et al.*, 2004), antibacterial (Dinesh *et al.*, 2015), anticarcinogenic (Saini *et al.*, 2010), anticonvulsant (Rathor *et al.*, 2014), antifungal (Das *et al.*, 2011), anti-inflammatory (El Sayed *et al.*, 2016), antineoplastic (Tabolacci *et al.*, 2015), antioxidant (Haritha *et al.*, 2014), antiseptic (Yun *et al.*, 2009), effective in reducing gastroesophageal reflux symptoms (Panahi *et al.*, 2015), efficient in treating psoriasis (Syed *et al.*, 1996) and oral lichen planu (Choonhakarn *et al.*, 2008), postoperative pain decrease (Eshghi *et al.*, 2010), hepatotoxic (Gbadegesin *et al.*, 2009), immune system improvement (Toliopoulos *et al.*, 2012), larvicidal (Subramaniam *et al.*, 2012), leishmanicidal (Dutta *et al.*, 2007), mosquitocidal (Dinesh *et al.*, 2015), neuroprotective agent in ischemia treatment (Guven *et al.*, 2016), potential to treat diabetes (Haritha *et al.*, 2014), reepithelizer (Atiba *et al.*, 2015) and wound healing (Anjum *et al.*, 2016).

Activities on intestinal parasites

Anticoccidial (El-banna *et al.*, 2013) and anthelmintic (Sprenger, 2015).

BANANA (Figure No. 3E)**Botanical characterization**

Musa paradisiaca L. (Musaceae) is a robust herbal species that reaches up to 9 m in height. It presents underground stem; simple, spiral, cartaceous leaves with secondary ribs parallel to the main one and overlapping sheaths forming a pseudostem; cymose inflorescence with proximal female tops and distal male tops, as well as berry-type oblong fruit. It is

mainly grown in tropical and subtropical countries worldwide and presents high dietary potential. The species is cultivated and widely distributed in all five Brazilian regions (Imam & Akter, 2011; BFG, 2020).

Diverse pharmacological properties

Antibacterial (Karuppiyah & Mustaffa, 2013), antidiarrheal (Yakubu *et al.*, 2015), antifungal (Jawla *et al.*, 2012), antihypertensive (Jaiprakash *et al.*, 2006), antioxidant (Karuppiyah & Mustaffa, 2013), antiulcer potential (Goel *et al.*, 2001), antivenom (Borges *et al.*, 2005), hemostatic (Weremfo *et al.*, 2011), hepatoprotective (Nirmala *et al.*, 2012), hypocholesterolemic (Saraswathi & Gnanam, 1997), hypoglycemic (Iroaganachi *et al.*, 2015), it improves breast milk production (Mahmood *et al.*, 2012), and leishmanicidal (Silva *et al.*, 2014).

Activities on intestinal parasites

Anthelmintic (Oliveira *et al.*, 2010; Hussain *et al.*, 2010; Marie-Magdeleine *et al.*, 2010; Hussain *et al.*, 2011; Marie-Magdeleine *et al.*, 2014).

BATATA DE PURGA**Botanical characterization**

Operculina macrocarpa (L.) Urb. (Convolvulaceae) is a liana species presenting milky latex; compound, digitized and 5-foliolate leaves; cyme inflorescence; 1-2-flora, white flowers with infundibuliform corolla, pubescent on the mesopetal veins; operculated and globose capsule fruits. The species is native to Brazil and it is widely distributed in the country's Northern, Northeastern, Midwestern and Southeastern regions, in phytogeographic domains such as the Caatinga, Cerrado and Atlantic Forest (Nepomuceno *et al.*, 2016; BFG, 2020).

Diverse pharmacological properties

Antiplatelet (Pierdoná *et al.*, 2014), larvicidal (Paiva *et al.*, 2011) and laxative (Paganotte *et al.*, 2016). It is noteworthy that laxatives may be used in antihelmintic therapy (Futija *et al.*, 2008) and the use of laxatives in the fight against intestinal parasites may date over 5300 years old (Capasso, 1998).

Activities on intestinal parasites

Not reported.

BETERRABA**Botanical characterization**

Beta vulgaris L. (Amaranthaceae) is a cultivated herbal species that reaches up to 1.2 m in height (it rarely reaches 2 m in height). It presents basal leaves forming a rosette; small hermaphroditic flowers gathered in dense inflorescences and arranged in glomeruli in groups of one to three (rarely eight) flowers in the armpits of bracts or in the upper half of bractless inflorescences; utricle fruit. The species is native to temperate regions in Europe and Northern Africa; it is one of the main agricultural crops and it accounts for approximately 30% of the world sugar supply. In Brazil, it is grown in the Southeastern and Southern regions of the country and presents likely incidence in the Northern, Northeastern, Midwestern, Southeastern and Southern regions (Lange *et al.*, 1999; Biondo *et al.*, 2014; Kew Science, 2020; BFG, 2020).

Diverse pharmacological properties

Antiacetylcholinesterase (Sacan & Yanardag, 2010), antibacterial (Koochak *et al.*, 2010), anticancer (Kazimierczak *et al.*, 2014), antifungal (Kragh *et al.*, 1995), anti-inflammatory (Jain *et al.*, 2011), antioxidant (Sacan & Yanardag, 2010), anti-radical (Escribano *et al.*, 1998), antiviral (Iglesias *et al.*, 2015), hepatoprotective (Agarwal *et al.*, 2006), hypoglycemic (Yoshikawa *et al.*, 1996) and neuroprotective (Nade *et al.*, 2015).

Activities on intestinal parasites

Not reported.

CAÇATINGA (Figure No. 3F)

Botanical characterization

Croton argyrophyllus Kunth (Euphorbiaceae) is a shrub species that reaches 1-4 m in height. It presents silver-green branches; stellate, lepidote-to-dentate-lepidote trichomes; translucent latex; simple, alternate, cartaceous leaves; monoic terminal inflorescence showing silver-yellow flowers staminated at the apex and pistilized at the base, as well as capsule fruit. It has disjunct distribution in South America (Bolivia, Paraguay, Venezuela and Brazil). The species is native to Brazil and naturally grows in the Northern and Northeastern regions of the country, in phytogeographic domains such as Caatinga and Amazon, mainly in semi-arid environments, in sandy or stony soils with semi-deciduous-to-deciduous vegetation (Carneiro-Torres, 2009; Silva *et al.*, 2010; BFG, 2020).

Diverse pharmacological properties

Anti-inflammatory (Ramos *et al.*, 2013).

Activities on intestinal parasites

Not reported.

CAJUEIRO (Figure No. 3G)

Botanical characterization

Anacardium occidentale L. (Anacardiaceae) is a tree species that reaches 2-15 m in height. It presents simple, whole, alternate, cartaceous-to-subcoriate leaves; obovate, oblong or widely oblong blade; terminal tyrsoid inflorescence; flowers with white or light-green petals presenting red or pink lines in anthesis and dark-red lines after fertilization; piriform drupe fruit; yellow, orange or red fleshy hipocarp. The species is native to Brazil and is widely distributed in virtually all states in the Northern, Northeastern, Midwestern and Southeastern regions of the country, in phytogeographic domains such as the Amazon, Caatinga, Atlantic Forest, Pampa and Pantanal (Silva-Luz, 2011; BFG, 2020).

Diverse pharmacological properties

Aphrodisiac (Mbatchou & Kosoono, 2012), antibacterial (Ajileye *et al.*, 2015), antidiarrheal (Araújo *et al.*, 2015), antifungal (Mahata *et al.*, 2014), anti-inflammatory (Olajide *et al.*, 2013), anti-mutagenic (Barcelos *et al.*, 2007), antioxidant (Encarnação *et al.*, 2016), anti-tumor (Mothé *et al.*, 2008), antiulcerogenic (Konan & Bacchi, 2007), emulsifying (Porto & Cristianini, 2014), gastroprotective (Morais *et al.*, 2010), hypoglycemic (Dionísio *et al.*, 2015), larvicidal (Torres *et al.*, 2015), leishmanicidal (Franca *et al.*, 1993), molluscicidal (Souza *et al.*, 1992), renal function improvement (Tedong *et al.*, 2006) and thrombolytic (Khan *et al.*, 2011).

Activities on intestinal parasites

Anthelmintic (Ademola & Eloff, 2011) and antiprotozoal (Neiva *et al.*, 2014).

CATINGUEIRA PREPREM (Figure No. 3H)

Botanical characterization

Poincianella microphylla (Mart. ex G. Don) L. P. Queiroz (Fabaceae) it is a shrub or small tree species that reaches up to 1.5 m in height. It presents compound, bipinnate, alternate and cartaceous leaves; inflorescence in terminal or axillary raceme up to 6-

cm long; flowers with golden-yellow petals, vexillum petal with reddish spots; and strongly compressed and apiculate oblong-to-oblongate fruit. The species is endemic to Brazil; it is typical of and widely distributed in Caatinga environments found in Bahia, Pernambuco and Piauí States (Northeastern region) (Queiroz, 2009; BFG, 2020).

Diverse pharmacological properties

Antibacterial (Silva *et al.*, 2006) and antibiofilm (Silva *et al.*, 2015b).

Activities on intestinal parasites

Not reported.

CHUCAINHO

Botanical characterization

Cardiospermum halicacabum L. (Sapindaceae) it is an herbaceous vine species presenting compound, biternate, 9-foliolate and cartaceous leaves; axillary, spiciform tyrsoid inflorescences with three branches, flowers with white petals presenting yellowish crest at the apex; globose, 3-winged and inflated septifragal capsule fruit. Cosmopolitan species that is widely distributed in the Americas, Africa, South Asia and Oceania. It naturally grows in the Brazilian Northern, Northeastern, Midwestern, Southeastern and Southern regions, in phytogeographic domains such as the Amazon, Caatinga, Cerrado, Atlantic Forest, Pampa and Pantanal (Ferrucci, 2000; Pereira *et al.*, 2016; BFG, 2020).

Diverse pharmacological properties

Analgesic (Muthumani *et al.*, 2010), antibacterial (Jeyadevi *et al.*, 2013), anticonvulsant (Dhayabaran *et al.*, 2012), antidiarrheal (Rao *et al.*, 2006), antifungal (Jeyadevi *et al.*, 2013), anti-inflammatory (Huang *et al.*, 2011), antioxidant (Rupeshkumar *et al.*, 2012), antiparasmodial (Waako *et al.*, 2005), antipyretic (Asha & Pushpangadan, 1999), antispasmodic (Pillai & Vijayamma, 1985), antiulcer (Muthumani *et al.*, 2010), anxiolytic (Kumar *et al.*, 2011), collagenase inhibitor (Ganesan *et al.*, 2011), diuretic (Chandra *et al.*, 2008), hepatoprotective (Rupeshkumar *et al.*, 2012), hypoglycemic (Veeramani *et al.*, 2012), hypolipidemic (Veeramani *et al.*, 2010), hypotensive (Somanadhan *et al.*, 1999), male fertility enhancer (Peiris *et al.*, 2015), potential to treat chronic skin diseases (Jong *et al.*, 2013), repellent (Govindarajan & Sivakumar, 2012) and

sedative (Pillai & Vijayamma, 1985).

Activities on intestinal parasites

Anthelmintic (Khunkitti *et al.*, 2000; Boonmars *et al.*, 2005).

EUCALIPTO

Botanical characterization

Eucalyptus globulus Labill. (Myrtaceae) is a forest tree species that reaches 30-55 m in height; it exceptionally exceeds 90 m in height. It presents erect and slender trunk, with branches only in the terminal part, where they form a sparse and irregular canopy at great height. Its leaves are considerably polymorphic. Young individuals present opposite, sessile, oblanceolate 6-to-15 cm long leaves covered by a waxy bluish tegument; they emerge in alternating pairs and give juvenile plants alternifolia features in quadrangular section stems. On the other hand, adult trees present narrow, rectilinear to falciform-shaped, petiolate, 15-to-35 cm long leaves, with grayish green tegument emerging alternately along cylindrical stems; white or cream flowers, either solitary or grouped in up to three individuals; and capsule fruit. The species is native to Australia and Tasmania and it is currently one of the most widely grown trees in production forests. It was introduced in several countries in Europe, South America, Africa and Asia, mainly to produce timber, paper, cellulose and essential oils. It also grows in anthropic areas in all five Brazilian regions (Kirkpatrick, 1974; Jordan *et al.*, 1993; Naithani, 2014; Salehi *et al.*, 2019; BFG, 2020).

Diverse pharmacological properties

Anti-acetylcholinesterase (Aazza *et al.*, 2011), anti-acne (Athikomkulchai *et al.*, 2008), antibacterial (Dezsi *et al.*, 2015), anti-inflammatory (Medeiros *et al.*, 2007), antioxidant (Dezsi *et al.*, 2015), lipid-lowering (Eidi *et al.*, 2009) and potential antiasthma (Ikawati *et al.*, 2001).

Activities on intestinal parasites

Anthelmintic (Macedo *et al.*, 2009).

FEIJÃO DE ARRANCA

Botanical characterization

Phaseolus vulgaris L. (Fabaceae) is a liana species of determined or indeterminate growth. It presents compound, 3-foliolate, alternate and cartaceous

leaves; racemous, axillary and terminal inflorescences; papilionaceous flowers; white, pink or violet petals uniformly distributed throughout the corolla, or corolla with vexillum and wings with different colors or shades; as well as legume fruit presenting two valves joined by two sutures, one dorsal and one ventral. The species is cultivated in Brazil, where it is widely distributed in anthropic areas in all its five regions (Silva & Costa, 2003; BFG, 2020).

Diverse pharmacological properties

Antibacterial (Amarowicz *et al.*, 2008), antidiabetic (Mojica & Mejía, 2016), anti-inflammatory (Monk *et al.*, 2015), antimutagenic, antioxidant (Rocha-Guzmán *et al.*, 2007) and insecticidal (Karbache *et al.*, 2011).

Activities on intestinal parasites

Anthelmintic (Ríos-de Álvarez *et al.*, 2012).

JATOBÁ MANSO

Botanical characterization

Hymenaea courbaril L. (Fabaceae) is a tree species that reaches 6-15 m in height. It presents compound, bifoliolate, alternate, coriaceous, glabrous, predominantly lanceolate-to-oblong leaves with acute and obtuse apex; inflorescence in terminal panicle; flowers with campanulate hypanthium and white petals; legume fruits with dark-brown lenticelous surface. It is widely distributed from Southern Mexico and the Antilles to Southwestern Brazil; the species is mostly represented in forest environments. It is native to Brazil and is widely distributed in the Northern, Northeastern, Midwestern, Southeastern and Southern regions of the country, in phytogeographic domains such as the Amazon, Caatinga, Cerrado, Atlantic Forest and Pantanal (Queiroz, 2009; BFG, 2020).

Diverse pharmacological properties

Antifungal (Costa *et al.*, 2014), anti-inflammatory, antioxidant (Bezerra *et al.*, 2013), antiviral, (Cecílio *et al.*, 2012) and myorelaxant (Bezerra *et al.*, 2013).

Activities on intestinal parasites

Not reported.

MAMÃO (Figure No. 3I)

Botanical characterization

Carica papaya L. (Caricaceae) is a tree species that can reach 6-9 m in height. It presents palm-lobed leaves that emerge from the upper parts of the stem in spiral array, in hollow, succulent petioles; axillary inflorescence with pistillate, staminate or bisexual flowers; fleshy and lightly scented petals; and berry fruit. The species is typically tropical and has likely origin in Central America. It is widely distributed in all states of all five Brazilian regions and in phytogeographic domains such as the Amazon, Caatinga, Cerrado and Atlantic Forest (Paull & Duarte, 2011; Wijaya & Chen, 2013; BFG, 2020).

Diverse pharmacological properties

Abortive (Anuar *et al.*, 2008), action against fish ectoparasite (Ekanem *et al.*, 2004), analgesic (Hasimun *et al.*, 2014), antibacterial (Kokila *et al.*, 2016), anticancer (Li *et al.*, 2012), anti-fertility (Nwaehujor *et al.*, 2014), anti-fungal (Chávez-Quintal *et al.*, 2011), anti-inflammatory (Owoyele *et al.*, 2008), anti-malarial (Tarkang *et al.*, 2014), antioxidant (Galang *et al.*, 2016), antisickling (Oduola *et al.*, 2006), antithrombotic (Zunjar *et al.*, 2016), antiulcerogenic (Oloyede *et al.*, 2015), antiviral (Chinnappan *et al.*, 2016), anxiolytic (Kebebew & Shibeshi, 2013), astringent action (Oloyede, 2005), contraceptive (Lohiya *et al.*, 2005), hepatoprotective (Awodele *et al.*, 2016), hypoglycemic (Juárez-Rojop *et al.*, 2012), molluscicidal (Jaiswal & Singh, 2008), potential to treat allergic diseases (Otsuki *et al.*, 2010), proteolytic (Thomás *et al.*, 2009) and wound healing (Gurung & Škalko-Basnet, 2009).

Activities on intestinal parasites

Anthelmintic (Satrija *et al.*, 1995; Kermanshai *et al.*, 2001; Okeniyi *et al.*, 2007). It is worth highlighting that *C. papaya* compounds obtained from both latex (Moraes *et al.*, 2017) and seeds (Cabral *et al.*, 2019) have anthelmintic activity upon *Strongyloides venezuelensis* Brumpt, 1934 eggs and larvae. The *C. papaya* action on intestinal nematodes is largely due to the worm cuticle disruption by cysteine proteinases (Levecke *et al.*, 2014; Luoga *et al.*, 2015).

MAMONA (Figure No. 3J)

Botanical characterization

Ricinus communis L. (Euphorbiaceae) is a shrub species that reaches ca. 1.5 m in height. It presents simple, alternate and cartaceous leaves; racemose

inflorescence; unisexual flowers: staminate flowers at the base of branches and pistillate flowers at the apex of branches; as well as capsule fruits. The species naturally grows in India, Africa, Europe and Brazil, where it is cultivated and widely distributed in all states of all its five regions, in phytogeographic domains such as the Amazon, Caatinga, Cerrado, Atlantic Forest, Pampa and Pantanal (Linnaeus, 1753; BFG, 2020).

Diverse pharmacological properties

Acaricidal (Ghosh *et al.*, 2013), antiasthmatic (Taur & Patil, 2011), antibacterial (Rampadarath & Puchooa, 2016), antifungal (Naz & Bano, 2012), anti-inflammatory (Ilavarasan *et al.*, 2006), antinociceptive (Taur *et al.*, 2011), antioxidant (Wafa *et al.*, 2014), antitumor (Lin & Liu, 1986), binder (Etzler & Branstrator, 1974), contraceptive (Nath *et al.*, 2013), hypoglycemic (Shokeen *et al.*, 2008), immunomodulatory (Mannoji *et al.*, 1986), insecticidal (Rampadarath *et al.*, 2014), potential to be used in osteoarthritis treatment (Ziaei *et al.*, 2016), as well as in bone and reconstruction processes (Leite & Ramalho, 2008), larvicidal (Rampadarath & Puchooa, 2016), leishmanicidal (Jumba *et al.*, 2015), neuroleptic (Ferraz *et al.*, 1999), neuroprotective (Lee *et al.*, 2012), toxic (Coopman *et al.*, 2009).

Activities on intestinal parasites

Anthelmintic (Zahir *et al.*, 2012; Salles *et al.*, 2014).

MANJERICÃO

Botanical characterization

Ocimum campechianum Mill. (Lamiaceae) is an herbal species that reaches up to 60 cm in height. It presents aromatic, opposite, simple leaves; lax, branched inflorescence with lateral branches smaller than, or equal to, the main branch; whorls with 6 flowers; fruitful calyx with upper lip flowing along the tube and internally-purple, white corolla included in the calyx; uniformly brown nut-type fruit. The species is native to Brazil, where it is widely distributed in all its five regions, in phytogeographic domains such as the Amazon, Caatinga, Cerrado and Atlantic Forest (Albuquerque & Andrade, 1998; BFG, 2020).

Diverse pharmacological properties

Antibacterial (Carović-Stanko *et al.*, 2010), repellent (Benitez *et al.*, 2014) and antioxidant (Dzib-Guerra *et*

al., 2016).

Activities on intestinal parasites

Not reported.

PINHÃO BRABO (Figure No. 3L)

Botanical characterization

Jatropha mollissima (Pohl) Baill. (Euphorbiaceae) is a subshrub or shrub species that reaches up to 3 m in height. It presents simple, 5-lobed, alternate and membranaceous leaves; inflorescences in terminal dichasium with unisexual flowers; staminate and pistillate flowers with red petals; as well as tricoca capsule fruit. The species is native to Brazil and is widely distributed in the Northern, Northeastern, Midwestern and Southeastern regions, in phytogeographic domains such as the Amazon, Caatinga and Cerrado (Sátiro & Roque, 2008; BFG, 2020).

Diverse pharmacological properties

Antioxidant (Melo *et al.*, 2010b).

Activities on intestinal parasites

Anthelmintic (Ribeiro *et al.*, 2014).

PITÓ

Botanical characterization

Croton heliotropiifolius Kunth (Euphorbiaceae) is a subshrub or shrub species that reaches 0,7–2,5 m in height. It has simple, alternate and membranaceous leaves presenting the same color at both sides; racemose inflorescence with unisexual flowers, greenish-white staminate and greenish-yellow pistillate flowers; as well as tricoca capsule fruit. The species is widely distributed in the neotropical region. It is native to Brazil and more often observed in the Northeastern region of the country, although it also grows in the Midwestern and Southeastern regions of it, in phytogeographic domains such as the Amazon, Caatinga, Cerrado and Atlantic Forest (Sátiro & Roque, 2008; BFG, 2020).

Diverse pharmacological properties

Antiacetylcholinesterase, antifungal (Queiroz *et al.*, 2014) and insecticidal (Silva *et al.*, 2012).

Activities on intestinal parasites

Not reported.

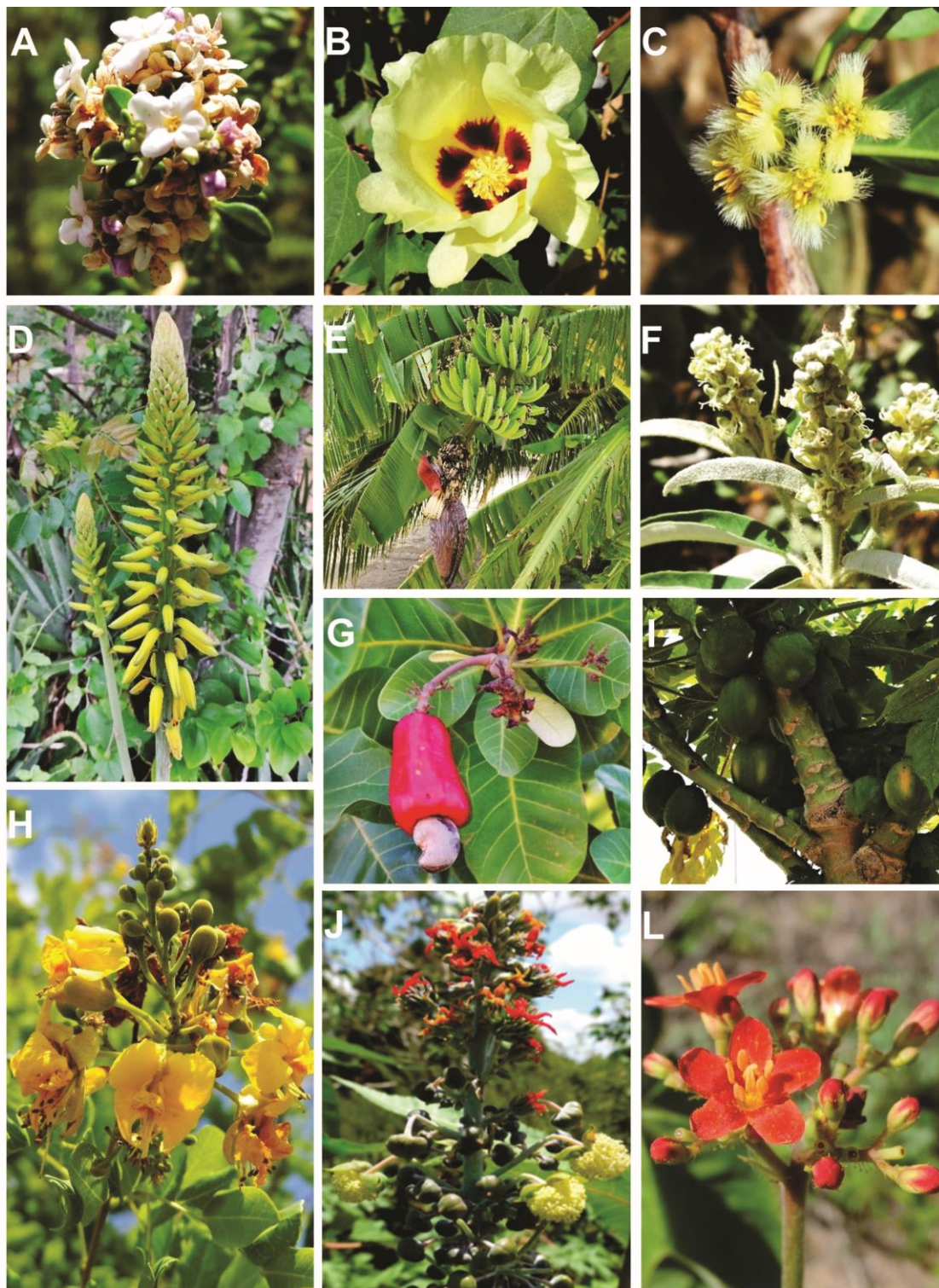


Figure No. 3

Ethnospecies indicated as antiparasitic agents by Kantaruré-Batida indigenous, Bahia State, Brazil. A: *Lippia thymoides*; B: *Gossypium hirsutum*; C: *Ximenia americana*; D: *Aloe vera*; E: *Musa paradisiaca*; F: *Croton argyrophyllus*; G: *Anacardium occidentale*; H: *Poincianella microphylla*; I: *Carica papaya*; J: *Ricinus communis*; L: *Jatropha molissima* (Source: Vasco-dos-Santos DRV, Santos JV)

QUIXABEIRA

Botanical characterization

Sideroxylon obtusifolium (Roem. & Schult.) T.D.Penn. (Sapotaceae) it is a tree species that can reach up to 18 m in height. The apex of its branches are hanging and thorny; it presents simple, coriaceous leaves, either opposite to each other or arranged in a spiral; inflorescence with white or yellow-green or cream, aromatic flower clusters comprising 2-20 flowers; as well as drupe fruit. It is found in some countries in North and South America. The species is native to Brazil, it is widely distributed in the Northern region of the country (Tocantins State), as well as in all states in the Northeastern, Midwestern, Southeastern and Southern regions, in phytogeographic domains such as Caatinga, Cerrado, Atlantic Forest and Pantanal (Silva & Dantas, 2017; BFG, 2020).

Diverse pharmacological properties

Antiandrogenic (Bobach *et al.*, 2014), antibacterial (Aquino *et al.*, 2016), antinociceptive (Araujo-Neto *et al.*, 2010), anti-inflammatory (Leite *et al.*, 2015) and antioxidant (Figueiredo & Lima, 2015).

Activities on intestinal parasites

Not reported.

DISCUSSION

Based on data accessed through the present systematic review, 48% of plants used by Kantaruré indigenous have effect on intestinal parasites, namely: Aroeira (*M. urundeuva*), Babosa (*A. vera*), Banana (*M. paradisiaca*), Cajueiro (*A. occidentale*), Chucaíno (*C. halicacabum*), Eucalipto (*E. globulus*), Feijão de arranca (*P. vulgaris*), Mamão (*C. papaya*), Mamona (*R. communis*) and Pinhão brabo (*J. molissima*) (Table No. 1). Studies conducted with Tamang indigenous people living in Rasuwa district, Nepal (Upriety *et al.*, 2010), as well as with residents in Dominica, West Indies (Quinlan *et al.*, 2002), have shown agreement between local knowledge about and pharmacological potential of plants.

Concerning the assay types, 80% were performed *in vitro*, 5% were conducted *in vivo* and 15% of studies performed both types. According to Wink (2012), despite the promising results of tests performed *in vitro*, clinical progress is neglected because parasites mainly affect economically vulnerable populations who cannot afford drugs.

Therefore, they are not priority for the pharmaceutical industry. Most articles (85%) presented data about parasites of veterinary interest, mainly about *Haemonchus contortus* (Rudolphi, 1803), which is a gastrointestinal worm accounting for considerable economic damage to global carpinoculture (Silva, 2014). Thus, it is essential emphasizing that some of the human anthelmintic drugs available in the market nowadays derive from veterinary medicine.

Only three of the analyzed studies performed experiments with human parasites, two of them, Boonmars *et al.* (2005) and Neiva *et al.* (2014) were carried out *in vitro*. Boonmars *et al.* (2005) observed *C. halicacabum* extract activity against third-stage *Strongyloides stercoralis* (Bavay, 1876) larvae whereas Neiva *et al.* (2014) recorded giardicidal activity of hydroalcoholic extract of *A. occidentale* leaves. Okeniyi *et al.* (2007) was the only study conducted *in vivo*, whose sample, which comprised 60 parasitized children, was treated with elixir added with *C. papaya* seeds, which was effective against 7 parasites (Table No. 1). Recently, Azadbakht *et al.* (2020) have shown maximum activity of essential oils of *E. globulus* against *Giardia lamblia* Ancy, 1906 and *Entamoeba histolytica* Schaudinn, 1903. However, the aforementioned studies did not explain the active compounds and their respective action mechanisms, which could have been investigated by using electron microscopy to determine cellular and subcellular targets of antiparasitic drugs (Vannier-Santos & De Castro, 2009; Dos Anjos *et al.*, 2016).

Secondary metabolism products such as alkaloids, terpenoids, polysaccharides and peptides, which can promote lesser costly and more effective therapy, are the main plant molecule constituents used to treat worm infections (Bahmani *et al.*, 2014; Singh *et al.*, 2020). *Artemisia annua* L., which is historically used as dewormer, anti-hemorrhoid and antipyretic drug in China, is a successful example of it (Oriakpono *et al.*, 2012). Artemisinin is one of the active agents of this species; this sesquiterpene lactone presenting endoperoxide bond, is isolated from plant leaves (Rehder *et al.*, 2002; Taranto *et al.*, 2006). Its compounds may also display antiviral activity against the severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2), recently reviewed (Khan *et al.*, 2021; Remali & Aizat, 2021). Despite remarkable controversy, other antimalarials such as chloroquine and its derivative

hydroxychloroquine are in test for COVID 19 (Million *et al.*, 2020; Güner *et al.*, 2021) and such activity may be explained at least in part for the

autophagy downmodulation (Tang *et al.*, 2020; Klionsky *et al.*, 2021).

Table No. 1
Antiparasitic activities of species used by Kantaruré-Batida community, Bahia State, Brazil, accessed through systematic review. Note: DNF=Data not found

Plant species/ reference	Used part	Extract type(s)	Concentration/ Dose	Assay type(s)	Infected animal/ Fecal collection	Tested parasite(s)
<i>Aloe vera</i> (L.) Berm.f.						
El-Banna <i>et al.</i> , 2013	Leaves	Aqueous	100 ppm	<i>In vivo</i>	Chickens	<i>Eimeria acervulina</i> <i>E. maxima</i> <i>E. necatrix</i> <i>E. tenella</i> <i>E. brunette</i>
Sprenger, 2015	Leaves	Aqueous and ethanolic	0.78 to 100 mg/mL	<i>In vitro</i>	Eggs from goat feces	<i>Haemonchus</i> <i>contortus</i>
<i>Anacardium occidentale</i> L.						
Ademola & Ellof, 2011	Leaves	Liquid (acetone)	0.311 and 1.72 mg/mL	<i>In vitro</i>	Eggs from sheep feces	<i>Haemonchus</i> <i>contortus</i>
Neiva <i>et al.</i> , 2014	Leaves	Hydroalcoholic	5 mg/mL	<i>In vitro</i>	Axenic strains	<i>Giardia lamblia</i>
<i>Cardiospermum halicacabum</i> L.						
Khunkitti <i>et al.</i> , 2000	Shoot	Aqueous and ethanolic	0, 100, 500,1000 and 2000 µg/mL ⁻¹	<i>In vitro</i>	Peritoneal cavity of birds	<i>Brugia pahangi</i>
Boonmars <i>et al.</i> , 2005	DNF	Hydroalcoholic	2000 µg/mL	<i>In vitro</i>	Larvae from dog feces	<i>Strongyloides</i> <i>stercoralis</i>
<i>Carica papaya</i> L.						
Satrija <i>et al.</i> , 1995	Fruits	Aqueous	2, 4, 6 and 8 g/kg of body weight	<i>In vivo</i>	Rats	<i>Heligmosomoides</i> <i>polygyrus</i>
Kermanshai <i>et al.</i> , 2001	Seed	Aqueous and oil	10 to 20 µL 0.04 and 0.1 µL	<i>In vitro</i>	DNF	<i>Caenorhabditis elegans</i>
Okeniyi <i>et al.</i> , 2007	Seed	Seeds and honey (Placebo)	20 mL	<i>In vivo</i>	Humans (Children)	<i>Ascaris lumbricoides</i> <i>Entamoeba histolytica</i> <i>Giardia lamblia</i> <i>Necator americanus</i> <i>Strongyloides</i> <i>stercoralis</i> <i>Taenia saginata</i> <i>Trichuris trichiura</i>

<i>Eucalyptus globulus</i> Labill.						
Macedo <i>et al.</i> , 2009	DNF	Oil	Different concentrations	<i>In vitro</i>	Eggs from sheep feces	<i>Haemonchus contortus</i>
<i>Jatropha mollissima</i> (Pohl) Baill.						
Ribeiro <i>et al.</i> , 2014	Stem	I Aqueous	I 1% at concentrations 100, 500 and 1000 µg/mL	I <i>In vivo</i>	I Sheep	<i>Haemonchus contortus</i>
		II Ethanolic	II 2.5 ml of ethanolic extract 660.80 µg/mL	II <i>In vitro</i>	II Eggs from sheep feces	
<i>Musa paradisiaca</i> L.						
Oliveira <i>et al.</i> , 2010	Leaves and pseudo stems	Aqueous	25, 75 and 150 mg/mL ⁻¹	<i>In vitro</i>	Eggs from sheep feces	<i>Haemonchus</i> spp.
Hussain <i>et al.</i> , 2010	Leaves	Aqueous and ethanolic	0.0207 and 0.4813 mg/mL ⁻¹	<i>In vitro</i>	Eggs from sheep feces	Nematoides
Hussain <i>et al.</i> , 2011	Folhas	Aqueous methanolic	I DNF II 100, 50, 25, 12.5, 6.25, 3.12, 1.56, 0.78, 0.39 and 0.19 mg/mL ⁻¹	I <i>In vivo</i> II <i>In vitro</i>	I Sheep II Eggs and adult individuals from sheep feces	<i>Haemonchus contortus</i>
Marie-Magdeleine <i>et al.</i> , 2010	Stems and leaves	Mixed with food	7000 g/day	<i>In vivo</i>	Lamb	<i>Haemonchus contortus</i>
Marie-Magdeleine <i>et al.</i> , 2014	Stems and leaves	Aqueous, methanolic and/or dichloromethane	2400, 1200, 600 and 300 µg/mL	<i>In vitro</i>	Lamb	<i>Haemonchus contortus</i>
<i>Myracrodruon urundeuva</i> Allemão						
Oliveira <i>et al.</i> , 2011	Leaves and stems	Aqueous and ethanolic	0.31 to 5 mg/mL	<i>In vitro</i>	Eggs and larvae from sheep feces	<i>Haemonchus contortus</i>
<i>Phaseolus vulgaris</i> L.						
Ríos-de Álvarez <i>et al.</i> , 2012	Seeds	Aqueous/ flour	I 2.3 mg of semi-purified PHA lectin per kg of body weight II DNF	I <i>In vivo</i> II <i>In vitro</i>	I Sheep II Larvae removed from tissue	<i>Trichostrongylus colubriformis</i> <i>Teladorsagia circumcincta</i>
<i>Ricinus communis</i> L.						
Zahir <i>et al.</i> , 2012	Leaves	Alcoholic	1.0 mL	<i>In vitro</i>	Adult individuals collected from the rumen of infected	<i>Paramphistomum cervi</i>

sheep						
Salles <i>et al.</i> (2014)	Seeds	Aqueous, NaCl and Tris-HCl	DNF	<i>In vitro</i>	Eggs from goat feces	<i>Haemonchus contortus</i>

Mengarda *et al.* (2020) have tested the antischistosomal activity of piperlongumine *in vivo*. This alkaloid, which is found in *Piper tuberculatum* Jacq. roots, has significantly reduced worm burden and egg production. Furthermore, the study has also shown damage to parasites' integument, based on electron microscopy. Another plant product, curcumin, derived from *Curcuma* sp., was shown to be cytotoxic and antiparasitic, largely due to redox properties, inducing oxidative stress in cancer, protozoal and helminthic models, triggering both autophagy and apoptosis (Haddad *et al.*, 2011; De Paula Aguiar *et al.*, 2016; Araveti & Srivastava, 2019; Gupta *et al.*, 2020). Its action on microtubules of the cytoskeleton in *G. lamblia*, *Plasmodium falciparum* (Wech, 1897), *Trypanosoma cruzi* (Chagas, 1909) and cancer cells, apparently for direct binding to tubulin molecules, was demonstrated by fluorescence spectroscopy and docking *in silico* (Banerjee *et al.*, 2010; Chakrabarti *et al.*, 2013; Sueth-Santiago *et al.*, 2016; Gutiérrez-Gutiérrez *et al.*, 2017). Thus, the development of new tools can lead to the elucidation of the various mechanisms of action of antiparasitic agents.

Other investigations have also reported the effectiveness of active plant compounds against intestinal parasites. Among them, one finds allicin, aspidin, ascaridol, toosendanin and pipartine, which derive from *Allium sativum* L. (Lima *et al.*, 2011), *Dryopteris* spp. (Magalhães *et al.*, 2010), *Chenopodium ambrosoides* L. (Kliks, 1985), *Melia toosendan* Siebold & Zucc. (Zhou, 2014) and *P. tuberculatum* (Moraes *et al.*, 2012), respectively.

Regarding species habit, 23.8% of the investigated species were herbs; 9.5% were lianas; 14.3% subshrubs or shrubs; 14.3% shrubs; 9.5% shrubs or small tree; and 28.7% were average-sized trees. Most plants with antiparasitic potential (Table 1) were trees (40%); they were followed by herbs (30%), as well as by lianas, subshrubs and shrubs (10%, each). This outcome can substantiate the discussion about the ecological appearance hypothesis, since the most apparent plants (trees)

have their potential best known and are mostly used likely due to their greater availability (Albuquerque *et al.*, 2013b).

Data collected in the current study do not substantiate the prediction that herbaceous plants are associated with the greater medicinal or cultural importance of a given species since, based on the free list, bush and tree plants such as *P. microphylla* (23%), *C. papaya* (13%) and *M. urundeuva* (10%) were the most cited by Kantaruré (Vasco-dos-Santos *et al.*, 2018). Therefore, seasonality appears to be a better predictor that points towards woody species as the most used plants in the traditional therapeutic arsenal available in caatinga phytogeographic domains, as observed by Almeida *et al.* (2011).

Silva *et al.* (2020) have investigated whether individuals' perception about the taste of plants could influence the selection of medicinal resources; however, they did not find data to substantiate their hypothesis. Thus, apparently, isolated aspects such as plant habit and taste are not good predictors to help better understanding selection mechanisms applied to therapeutic resources. They are likely to be better understood in light of the socio-ecological theory of maximization, according to which, selection processes are linked to cognitive and behavioral strategies adopted in different environmental contexts in order to expand the benefits to, and survival of, human groups (Albuquerque *et al.*, 2019).

Although 52% of the indicated plants do not have reported action in intestinal parasites, it is not possible inferring their ineffectiveness due to lack of standardized and validated antiparasitic tests. In addition, the investigated species have other properties that can help understanding their use, based on their likelihood to enable users' improvement, as reported by Vasco-dos-Santos *et al.* (2019). The need of conducting further research to better understand the properties of indigenous flora was also reported by Baranwal *et al.* (2012) and Raj and Dave (2019), who addressed antiarthritic and antidiabetic medicinal plants, respectively.

Based on the study conducted by Elisabetsky

and Posey (1989), who investigated plants used as remedies associated with sexuality and fertility by the Kayapó community, 48% of plant species were also used by other cultures for the same purpose, which indicated their potential effectiveness. Similarly, this aspect can also be applied to Babosa (*A. vera*) and Mamão (*C. papaya*) plants used by the Kantaruré community, which is also used by other indigenous communities in Northeastern Brazil to treat worm infections. *Aloe vera* is used by the Guajajara community (Coutinho *et al.*, 2002), whereas *C. papaya* plant is used by the Funil-ô (Silva, 2003), Tapeba (Morais *et al.*, 2005) and Pataxó communities (Cunha Lima *et al.*, 2012).

Based on data collected by Vasco-dos-Santos *et al.* (2018), the way plants are prepared by indigenous communities likely has implications on traditional therapy. For example, the combined use of banana (*M. paradisiaca*) and garlic (*Alium sativum* L.) can be a strategy adopted to enhance their effect, since both species have antiparasitic properties (Santos *et al.*, 2012; Marie-Magdeleine *et al.*, 2014). Other combinations reported in the literature comprised Batata de purga (*O. macrocarpa*) and Mamão (*C. papaya*); as well as Ameixa (*X. americana*) and Aroeira (*M. urundeuva*). Among them, only *M. urundeuva* and *C. papaya* have information available about their antiparasitic properties (Okeniyi *et al.*, 2007; Oliveira *et al.*, 2011). Therefore, further studies should be conducted about these species, as well as about the risks and benefits generated by their interaction, to test the potential-use hypothesis. After all, some medicinal plants that have potentially toxic substances should be used based on toxicological risks (Veiga Junior *et al.*, 2005).

REFERENCES

- Aazza S, Lyoussi B, Miguel MG. 2011. Antioxidant and antiacetylcholinesterase activities of some commercial essential oils and their major compounds. **Molecules** 16: 7672 - 7690.
<https://doi.org/10.3390/molecules16097672>
- Abebe D, Garedew B. 2019. Ethnobotanical survey of plants traditionally used for malaria prevention and treatment in indigenous villages of Tepi Town South West Ethiopia. **J Pharmacogn Phytother** 11: 9 - 16.
<https://doi.org/10.5897/jpp2018.0532>
- Achan J, Talisuna AO, Erhart A, Adoke Y, Tibenderana JK, Baliraine FN, Rosenthal PJ, D'Alessandro U. 2011. Quinine, an old anti-malarial drug in a modern world: Role in the treatment of malaria. **Malaria J** 10: 1 - 12. <https://doi.org/10.1186/1475-2875-10-144>
- Adamu HM, Abayeh OJ, Agho MO, Abdullahi AL, Uba A, Dukku HU, Wufem BM. 2005. An ethnobotanical survey of Bauchi State herbal plants and their antimicrobial activity. **J Ethnopharmacol** 99: 1 - 4.
<https://doi.org/10.1016/j.jep.2004.12.025>

CONCLUSIONS

Taken together, the current data have confirmed that indigenous ethno-knowledge about medicinal plants plays key role in species selection for pharmacological and phytochemical studies aimed at finding new drugs. They also highlight the importance of conducting research about parasites in traditional communities, mainly in indigenous communities living in Northeastern Brazil. Based on the present literature review, it is possible to assume that some plants have the potential to be investigated as deworming agents, if one takes into consideration the lack of studies performed for this purpose. Among them, one finds Catingueira prepem (*P. microphylla*), Caçatinga (*C. argyrophyllus*), Pitó (*C. heliotropiifolius*) and Alecrim (*L. thymoides*), which stand out as the main species used by Kantaruré-Batida indigenous community.

ACKNOWLEDGEMENTS

We acknowledge to the financial support for the translation of the Programa Interno de Auxílio Financeiro à Publicação em Periódicos Nacionais e Internacionais Qualificados para a Pós-Graduação *Stricto sensu* (PROPUBLIC), notice 113/2017 of Universidade do Estado da Bahia (UNEB), under granting term No. 14/2017; Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) for PhD grant to DRVS and for research productivity grant awarded to MAVS, as well as the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) – Finance Code 001 to JVS and LNL; and to Fundação de Apoio à Pesquisa do Estado da Bahia (FAPESB) for financial support (PIE0023/2016) to MJGA.

- Ademola IO, Eloff JN. 2011. Anthelmintic efficacy of cashew (*Anacardium occidentale* L.) on *in vitro* susceptibility of the ova and larvae of *Haemonchus contortus*. **Afr J Biotechnol** 10: 9700 - 9705. <https://doi.org/10.5897/ajb09.1706>
- Agarwal M, Srivastava VK, Saxena KK, Kumar A. 2006. Hepatoprotective activity of *Beta vulgaris* against CCl₄-induced hepatic injury in rats. **Fitoterapia** 77: 91 - 93. <https://doi.org/10.1016/j.fitote.2005.11.004>
- Ajileye OO, Obuotor EM, Akinkunmi EO, Aderogba MA. 2015. Isolation and characterization of antioxidant and antimicrobial compounds from *Anacardium occidentale* L. (Anacardiaceae) leaf extract. **J King Saud Univ Sci** 27: 244 - 252. <https://doi.org/10.1016/j.jksus.2014.12.004>
- Albuquerque UP, Andrade L. 1998. El género *Ocimum* L.(Lamiaceae) en el nordeste del Brasil. **An Jard Bot Madrid** 56: 43 - 64.
- Albuquerque UP, Medeiros PM, Almeida ALS, Monteiro JM, Lins Neto EMF, Melo JG, Santos, JP 2007. Medicinal plants of the caatinga (semi-arid) vegetation of NE Brazil: A quantitative approach. **J Ethnopharmacol** 114: 325 - 354. <https://doi.org/10.1016/j.jep.2007.08.017>
- Albuquerque UP, Silva JS, Loureiro J, Campos A, Sousa RS, Silva TC, Alves, RRN. 2013a. The current status of ethnobiological research in Latin America: gaps and perspectives. **J Ethnobiol Ethnomed** 9: 2 - 9.
- Albuquerque UP, Soldati GT, Ramos MA, Melo JG, Medeiros PM, Nascimento ALB, Ferreira Júnior WS. 2013b. **Pode o ambiente influenciar as nossas escolhas sobre o uso de recursos naturais? Evidências da aparência.** In: Etnobiologia bases ecológicas e evolutivas, NUPEEA, Recife, Brasil.
- Albuquerque UP, Medeiros PM, Ferreira Júnior WS, Silva TC, Silva RRV, Gonçalves-Souza T. 2019. Social-ecological theory of maximization: basic concepts and two initial models. **Biol Theory** 14: 73 - 85. <https://doi.org/10.1007/s13752-019-00316-8>
- Almeida CFCBR, Amorim ELC, Albuquerque UP. 2011. Insights into the search for new drugs from traditional knowledge: An ethnobotanical and chemical-ecological perspective. **Pharm Biol** 49: 1 - 10. <https://doi.org/10.3109/13880209.2010.551777>
- Almeida MLB, Freitas WES, Morais PLD, Sarmiento JDA, Alves RE. 2016. Bioactive compounds and antioxidant potential fruit of *Ximenia americana* L. **Food Chem** 192: 1078 - 1082. <https://doi.org/10.1016/j.foodchem.2015.07.129>
- Alves PM, Queiroz LMG, Pereira JV, Pereira MDSV. 2009. Atividade antimicrobiana, antiaderente e antifúngica *in vitro* de plantas medicinais brasileiras sobre microrganismos do biofilme dental e cepas do gênero *Candida*. **Rev Soc Bras Med Trop** 42: 222 - 224. <https://doi.org/10.1590/S0037-86822009000200028>
- Amaravadi RK, Kimmelman AC, Debnath J. 2019. Targeting autophagy in cancer: Recent advances and future directions. **Cancer Discov** 9: 1167 - 1181. <https://doi.org/10.1158/2159-8290.CD-19-0292>
- Amarowicz R, Dykes GA, Pegg RB. 2008. Antibacterial activity of tannin constituents from *Phaseolus vulgaris*, *Fagopyrum esculentum*, *Corylus avellana* and *Juglans nigra*. **Fitoterapia** 79: 217 - 219. <https://doi.org/10.1016/j.fitote.2007.11.019>
- Anjum S, Gupta A, Sharma D, Gautam D, Bhan S, Sharma A, Kapil A, Gupta B. 2016. Development of novel wound care systems based on nanosilver nanohydrogels of polymethacrylic acid with *Aloe vera* and curcumin. **Mater Sci Eng C** 64: 157 - 166. <https://doi.org/10.1016/j.msec.2016.03.069>
- Anuar NS, Zahari SS, Taib IA, Rahman MT. 2008. Effect of green and ripe *Carica papaya* epicarp extracts on wound healing and during pregnancy. **Food Chem Toxicol** 46: 2384 - 2389. <https://doi.org/10.1016/j.fct.2008.03.025>
- Anywar G, Kakudidi E, Byamukama R, Mukonzo J, Schubert A, Oryem-Origa H. 2020. Indigenous traditional knowledge of medicinal plants used by herbalists in treating opportunistic infections among people living with HIV/AIDS in Uganda. **J Ethnopharmacol** 246: 112205. <https://doi.org/10.1016/j.jep.2019.112205>
- Aquino DFS, Tirloni CAS, Menegati SELT, Cardoso CAL, Vieira SCH, Vieira MC, Simonet AM, Macías FA, Gasparotto Júnior AA. 2017. *Alibertia edulis* (L.C. Rich.) A.C. Rich - A potent diuretic arising from Brazilian indigenous species. **J Ethnopharmacol** 196: 193 - 200. <https://doi.org/10.1016/j.jep.2016.12.024>
- Aquino P, Gomes Figueredo F, Pereira N, Nascimento E, Martin A, Veras H, Oliveira C, Ferreira S, Leandro L, Silva M, Menezes I. 2016. Avaliação da atividade anti-inflamatória tópica e antibacteriana do extrato

- metanólico das folhas de *Sideroxylon obtusifolium*. *Acta Biol Colomb* 21: 131 - 140.
<https://doi.org/10.15446/abc.v21n1.48170>
- Araujo-Neto V, Bomfim RR, Oliveira VOB, Passos AMPR, Oliveira JPR, Lima CA, Mendes SS, Estevam CS, Thomazzi SM. 2010. Therapeutic benefits of *Sideroxylon obtusifolium* (Humb. ex Roem. & Schult.) T.D. Penn., Sapotaceae, in experimental models of pain and inflammation. *Rev Bras Farmacogn* 20: 933 - 938.
<https://doi.org/10.1590/S0102-695X2010005000043>
- Araújo TAS, Melo JG, Ferreira Júnior WS, Albuquerque UP. 2016. **Medicinal plants**. pp. 143–150. In Albuquerque UP, Alves RNN: **Introduction to ethnobiology**. Springer Cham Heidelberg, Germany.
https://doi.org/DOI.10.1007/978-3-319-28155-1_22
- Araújo TSL, Costa DS, Sousa NA, Souza LKM, Araújo S, Oliveira AP, Sousa FBM, Silva DA, Barbosa ALR, Leite JRSA, Medeiros JVR. 2015. Antidiarrheal activity of cashew GUM, a complex heteropolysaccharide extracted from exudate of *Anacardium occidentale* L. in rodents. *J Ethnopharmacol* 174: 299 - 307.
<https://doi.org/10.1016/j.jep.2015.08.020>
- Araveti PB, Srivastava A. 2019. Curcumin induced oxidative stress causes autophagy and apoptosis in bovine leucocytes transformed by *Theileria annulata*. *Cell Death Discov* 5: 1 - 14.
<https://doi.org/10.1038/s41420-019-0180-8>
- Asha VV, Pushpangadan P. 1999. Antipyretic activity of *Cardiospermum halicacabum*. *Indian J Exp Biol* 37: 411 - 414.
- Athikomkulchai S, Watthanachaiyingcharoen R, Tunvichien S, Vayumhasuwan P, Karnsomkiet P, Sae-Jong P, Ruangrunsi N. 2008. The development of anti-acne products from *Eucalyptus globulus* and *Psidium Guajava* oil. *J Health Res* 22: 109 - 113.
- Atiba A, Wasfy T, Abdo W, Ghoneim A, Kamal T, Shukry M. 2015. *Aloe vera* gel facilitates re-epithelialization of corneal alkali burn in normal and diabetic rats. *Clin Ophthalmol* 9: 2019 - 2026.
<https://doi.org/10.2147/OPTH.S90778>
- Awodele O, Yemitan O, Ise PU, Ikumawoyi VO. 2016. Modulatory potentials of aqueous leaf and unripe fruit extracts of *Carica papaya* Linn. (Caricaceae) against carbon tetrachloride and acetaminophen-induced hepatotoxicity in rats. *J Intercult Ethnopharmacol* 5: 27 - 35.
<https://doi.org/10.5455/jice.20160124113528>
- Azadbakht M, Chabra A, Akbarabadi AS, Hossein M, Monadi T, Akbari F. 2020. Anti-parasitic activity of some medicinal plants essential oils on *Giardia lamblia* and *Entamoeba histolytica*, *in vitro*. *Res J Pharmacogn* 7: 41 - 47. <https://doi.org/10.22127/rjp.2019.168142.1462>
- Aziz MA, Adnan M, Khan AH, Shahat AA, Al-Said MS, Ullah R. 2018. Traditional uses of medicinal plants practiced by the indigenous communities at Mohmand Agency, FATA, Pakistan. *J Ethnobiol Ethnomed* 14: 1 - 16. <https://doi.org/10.1186/s13002-017-0204-5>
- Bahmani M, Saki K, Rafieian-Kopaei M, Karamati SA, Eftekhari Z, Jelodari M. 2014. The most common herbal medicines affecting *Sarcomastigophora* branches: A review study. *Asian Pac J Trop Med* 7: S14 - S21.
[https://doi.org/10.1016/S1995-7645\(14\)60198-X](https://doi.org/10.1016/S1995-7645(14)60198-X)
- Banerjee M, Singh P, Panda D. 2010. Curcumin suppresses the dynamic instability of microtubules, activates the mitotic checkpoint and induces apoptosis in MCF-7 cells. *FEBS J* 277: 3437 - 3448.
<https://doi.org/10.1111/j.1742-4658.2010.07750.x>
- Baranwal V, Irchhaiya R, Alok S. 2012. Anti-arthritis activity of some indigenous plants: a review. *Res J Phytochem* 4: 981 - 986.
- Barcelos GRM, Shimabukuro F, Mori MP, Maciel MAM, Cólus IMS. 2007. Evaluation of mutagenicity and antimutagenicity of cashew stem bark methanolic extract *in vitro*. *J Ethnopharmacol* 114: 268 - 273.
<https://doi.org/10.1016/j.jep.2007.08.006>
- Basta PC, Orellana JDY, Arantes R. 2012. **Perfil epidemiológico dos povos indígenas no Brasil: Notas sobre agravos selecionados**. In Garnelo L, Pontes AL. Saúde indígena: Uma introdução ao tema. Ministério da Educação, Brasília, Brasil.
- Bayer C, Kubitzki K. 2003. **Malvaceae**. In Kubitzki K, Bayer C: **The families and Genera of vascular plants. Flowering plants Dicotyledons. Malvales, Capparales and non-betalain Caryophyllales**. Springer

- Verlag, Berlin, Germany. <https://doi.org/10.1007/978-3-662-07255-4>
- Benitez NP, Gallardo KC, Valencia CM. 2014. Actividad repelente de especies Vegetales que Crecen en la Región del Chocó. **Rev Ciênc Farm Básica Apl** 10: 196 - 203. <https://doi.org/10.18359/rfcb.330>
- Bezerra GP, Góis RWDS, Brito TS, Lima FJB, Bandeira MAM, Romero NR, Magalhães PJC, Santiago GMP. 2013. Phytochemical study guided by the myorelaxant activity of the crude extract, fractions and constituent from stem bark of *Hymenaea courbaril* L. **J Ethnopharmacol** 149: 62 - 69. <https://doi.org/10.1016/j.jep.2013.05.052>
- BFG - Flora do Brasil. 2020. **Flora do Brasil**. <http://floradobrasil.jbrj.gov.br>
- Biondo PBF, Boeing JS, Barizão ÉO, Souza NE, Matsushita M, Oliveira CC, Borosk M, Visentainer JV. 2014. Evaluation of beetroot (*Beta vulgaris* L.) leaves during its developmental stages: A chemical composition study. **Food Sci Technol** 34: 94 - 101. <https://doi.org/10.1590/S0101-20612014005000007>
- Bobach C, Schurwanz J, Franke K, Denkert A, Sung TV, Kuster R, Mutiso PC, Seliger B, Wessjohann LA. 2014. Multiple readout assay for hormonal (androgenic and antiandrogenic) and cytotoxic activity of plant and fungal extracts based on differential prostate cancer cell line behavior. **J Ethnopharmacol** 155: 721 - 730. <https://doi.org/10.1016/j.jep.2014.06.008>
- Boonmars T, Khunkitti W, Sithithaworn P, Fujimaki Y. 2005. *In vitro* antiparasitic activity of extracts of *Cardiospermum halicacabum* against third-stage larvae of *Strongyloides stercoralis*. **Parasitol Res** 97: 417 - 419. <https://doi.org/10.1007/s00436-005-1470-z>
- Borges MH, Alves DLF, Raslan DS, Piló-Veloso D, Rodrigues VM, Homs-Brandeburgo MI, Lima ME. 2005. Neutralizing properties of *Musa paradisiaca* L. (Musaceae) juice on phospholipase A2, myotoxic, hemorrhagic and lethal activities of crotalidae venoms. **J Ethnopharmacol** 98: 21 - 29. <https://doi.org/10.1016/j.jep.2004.12.014>
- Brandelli CLC, Vieira PDB, Macedo AJ, Tasca T. 2013. Remarkable anti-trichomonas vaginalis activity of plants traditionally used by the Mbyá-guarani indigenous group in Brazil. **Biomed Res Int** 2013: 1 - 7. <https://doi.org/10.1155/2013/826370>
- Brasileiro M, Egito A, Lima JR, Randau K, Pereira G, Rolim Neto P. 2008. *Ximenia americana* L.: botânica, química e farmacologia no interesse da tecnologia farmacêutica. **Fitoterapia** 89: 164 - 167.
- Cabral ERM, Moraes D, Levenhagen MA, Matos RAF, Costa-Cruz JM, Rodrigues RM. 2019. *In vitro* ovicidal and larvicidal activity of *Carica papaya* seed hexane extract against *strongyloides venezuelensis*. **Rev Inst Med Trop Sao Paulo** 61: 1 - 7. <https://doi.org/10.1590/s1678-9946201961059>
- Calou I, Bandeira MA, Aguiar-Galvão W, Cerqueira G, Siqueira R, Neves KR, Brito GA, Viana G. 2014. Neuroprotective properties of a standardized extract from *Myracrodruon urundeuva* Fr. All. (Aroeira-Do-Sertão), as evaluated by a Parkinson's disease model in rats. **Parkinson Dis** 2014: 1 - 11. <https://doi.org/10.1155/2014/519615>
- Capasso L. 1998. 5300 years ago, the Ice Man used natural laxatives and antibiotics. **Lancet** 352: 1864. [https://doi.org/10.1016/S0140-6736\(05\)79939-6](https://doi.org/10.1016/S0140-6736(05)79939-6)
- Carabajal MPA, Perea MC, Isla MI, Zampini IC. 2020. The use of jarilla native plants in a Diaguita-Calchaquí indigenous community from northwestern Argentina: An ethnobotanical, phytochemical and biological approach. **J Ethnopharmacol** 247: 112258. <https://doi.org/10.1016/j.jep.2019.112258>
- Carlini EA, Duarte-Almeida JM, Rodrigues E, Tabach R. 2010. Assessment of the toxicity of the Brazilian pepper trees *Schinus terebinthifolius* Raddi (Aroeira-da-praia) and *Myracrodruon urundeuva* Allemão (Aroeira-do-sertão). **Phytother Res** 20: 140 - 146. <https://doi.org/10.1002/ptr.4767>
- Carneiro-Torres DS. 2009. **Diversidade de Croton L. (Euphorbiaceae) no bioma Caatinga**. Thesis, Universidade Estadual de Feira de Santana, Brasil.
- Carović-Stanko K, Orlić S, Politeo O, Strikić F, Kolak I, Milos M, Satovic Z. 2010. Composition and antibacterial activities of essential oils of seven *Ocimum taxa*. **Food Chem** 119: 196 - 201. <https://doi.org/10.1016/j.foodchem.2009.06.010>
- Cecílio AB, Oliveira PC, Caldas S, Campana PRV, Francisco FL, Duarte MGR, Mendonça LAM, Almeida VL. 2016. Antiviral activity of *Myracrodruon urundeuva* against rotavirus. **J Ethnopharmacol** 26: 197 - 202. <https://doi.org/10.1016/j.jep.2015.10.005>

- Cecílio AB, Faria DB, Oliveira PDC, Caldas S, Oliveira DA, Sobral MEG, Duarte MGR, Moreira CPDS, Silva CG, Almeida VL. 2012. Screening of Brazilian medicinal plants for antiviral activity against rotavirus. **J Ethnopharmacol** 141: 975 - 981. <https://doi.org/10.1016/j.jep.2012.03.031>
- Chakrabarti R, Rawat PS, Cooke BM, Coppel RL, Patankar S. 2013. Cellular effects of curcumin on *Plasmodium falciparum* include disruption of microtubules. **Plos One** 8: 1 - 14. <https://doi.org/10.1371/journal.pone.0057302>
- Chandra PK, Kuppast IJ, Manjunath C, Jawahar N, Jubie S, Swapna B. 2008. Diuretic activity of whole plant extracts of *Cardiospermum halicacabum* (linn). **Pharmacogn Mag** 4: 80 - 82.
- Chang N, Luo Z, Li D, Song H. 2017. Indigenous uses and pharmacological activity of traditional medicinal plants in Mount Taibai, China. **Evid Based Complement Alternat Med** 1 - 11. <https://doi.org/10.1155/2017/8329817>
- Chaves MC, Santos FA, Menezes AMS, Rao VSN. 1998. Experimental evaluation of *Myracrodruon urundeuva* bark extract for antidiarrhoeal activity. **Phytother Res** 12: 549 - 552. [https://doi.org/10.1002/\(SICI\)1099-1573\(199812\)12:8<549::AIDPTR357>3.0.CO;2-L](https://doi.org/10.1002/(SICI)1099-1573(199812)12:8<549::AIDPTR357>3.0.CO;2-L)
- Chávez-Quintal P, González-Flores T, Rodríguez-Buenfil I, Gallegos-Tintoré S. 2011. Antifungal activity in ethanolic extracts of *Carica papaya* L. cv. Maradol leaves and seeds. **Indian J Microbiol** 51: 54 - 60. <https://doi.org/10.1007/s12088-011-0086-5>
- Chen WJ. 2016. Honoring antiparasitics: The 2015 Nobel Prize in physiology or medicine. **Biomed J** 39: 93 - 97. <https://doi.org/10.1016/j.bj.2016.04.002>
- Chinnappan S, Ramachandrapa VS, Tamilarasu K, Krishnan UM, Pillai AKB, Rajendiran S. 2016. Inhibition of platelet aggregation by the leaf extract of *Carica papaya* during dengue infection: An *in vitro* study. **Viral Immunol** 29: 164 - 168. <https://doi.org/10.1089/vim.2015.0083>
- Choonhakarn C, Busaracome P, Sripanidkulchai B, Sarakarn P. 2008. The efficacy of *Aloe vera* gel in the treatment of oral *Lichen planus*: A randomized controlled trial. **Br J Dermatol** 158: 573 - 577. <https://doi.org/10.1111/j.1365-2133.2007.08370.x>
- Coopman V, De Leeuw M, Cordonnier J, Jacobs W. 2009. Suicidal death after injection of a castor bean extract (*Ricinus communis* L.). **Forensic Sci Int** 189: 13 - 20. <https://doi.org/10.1016/j.forsciint.2009.04.019>
- Costa MP, Bozinis MCV, Andrade WM, Costa CR, Silva AL, Oliveira CMA, Kato L, Fernandes OFL, Souza LKH, Silva MRR. 2014. Antifungal and cytotoxicity activities of the fresh xylem sap of *Hymenaea courbaril* L. and its major constituent fisetin. **BMC Complement Altern Med** 14: 1 - 7. <https://doi.org/10.1186/1472-6882-14-245>
- Coutinho DF, Travassos LMA, Amaral FMM. 2002. Estudo etnobotânico de plantas medicinais utilizadas em comunidades indígenas no estado do Maranhão - Brasil. **Visão Acadêmica** 3: 7 - 12. <https://doi.org/10.5380/acd.v3i1.493>
- Cunha Lima ST, Rodrigues ED, Alves C, Merrigan TL, Melo T, Guedes MLS, Nascimento AF, Toralles M. 2012. The use of medicinal plants by an indigenous Pataxó community in NE Brazil. **Rev Bras Plant Med** 14: 84 - 91. <https://doi.org/10.1590/s1516-05722012000100012>
- Das S, Mishra B, Gill K, Ashraf MS, Singh AK, Sinha M, Sharma S, Xess I, Dalal K, Singh TP, Dey S. 2011. Isolation and characterization of novel protein with anti-fungal and anti-inflammatory properties from *Aloe vera* leaf gel. **Int J Biol Macromol** 48: 38 - 43. <https://doi.org/10.1016/j.ijbiomac.2010.09.010>
- De Paula Aguiar D, Moscardini MBM, Morais ER, De Paula RG, Ferreira PM, Afonso A, Belo S, Ouchida AT, Curti C, Cunha WR, Rodrigues V, Magalhães LG. 2016. *Curcumin* generates oxidative stress and induces apoptosis in adult *Schistosoma mansoni* worms. **Plos One** 11: 1 - 25. <https://doi.org/10.1371/journal.pone.0167135>
- Dezsi S, Badarau AS, Bischin C, Vodnar DC, Silaghi-Dumitrescu R, Gheldiu AM, Mocan A, Vlase L. 2015. Antimicrobial and antioxidant activities and phenolic profile of *Eucalyptus globulus* Labill. and *Corymbia ficifolia* (F. Muell.) K.D. Hill & L.A.S. Johnson leaves. **Molecules** 20: 4720 - 4734. <https://doi.org/10.3390/molecules20034720>
- Dhayabaran D, Florance J, Krsihnadas N, Indumathi, Muralidhar. 2012. Anticonvulsant activity of alcoholic root extract of *Cardiospermum halicacabum*. **Rev Bras Farmacogn** 22: 623 - 629.

<https://doi.org/10.1590/s0102-695x2012005000017>

- Diehl EE. 2013. **Etnobiologia e saúde dos povos indígenas**. In M. Haverroth M. : **Etnobiologia e saúde dos povos indígenas**. NUPEAA, Recife, Brasil.
- Dimitri MJ. 1978. **Enciclopedia argentina de agricultura y jardineria**. Editorial ACME S.A.C.I., Buenos Aires, Argentina.
- Dinesh D, Murugan K, Madhiyazhagan P, Panneerselvam C, Kumar PM, Nicoletti M, Jiang W, Benelli G, Chandramohan B, Suresh U. 2015. Mosquitocidal and antibacterial activity of green-synthesized silver nanoparticles from *Aloe vera* extracts: towards an effective tool against the malaria vector *Anopheles stephensi*? **Parasitol Res** 114: 1519 - 1529. <https://doi.org/10.1007/s00436-015-4336-z>
- Dionísio AP, Carvalho-Silva LB, Vieira NM, Goes TS, Wurlitzer NJ, Borges MF, Brito ES, Ionta M, Figueiredo RW. 2015. Cashew-apple (*Anacardium occidentale* L.) and yacon (*Smallanthus sonchifolius*) functional beverage improve the diabetic state in rats. **Food Res Int** 77: 171 - 176. <https://doi.org/10.1016/j.foodres.2015.07.020>
- Dos Anjos DO, Sobral Alves ES, Gonçalves VT, Fontes SS, Nogueira ML, Suarez-Fontes AM, Neves da Costa JB, Rios-Santos F, Vannier-Santos MA. 2016. Effects of a novel β -lapachone derivative on *Trypanosoma cruzi*: Parasite death involving apoptosis, autophagy and necrosis. **Int J Parasitol Drugs Drug Resist** 6: 207 - 219. <https://doi.org/10.1016/j.ijpddr.2016.10.003>
- Dutta A, Bandyopadhyay S, Mandal C, Chatterjee M. 2007. *Aloe vera* leaf exudate induces a caspase-independent cell death in *Leishmania donovani* promastigotes. **J Med Microbiol** 56: 629 - 636. <https://doi.org/10.1099/jmm.0.47039-0>
- Dzib-Guerra WC, Escalante-Erosa F, García-Sosa K, Derbré S, Blanchard P, Richomme P, Peña-Rodríguez LM. 2016. Anti-advanced glycation endproduct and free radical scavenging activity of plants from the Yucatecan flora. **Pharmacogn Res** 8: 276 - 279. <https://doi.org/10.4103/0974-8490.188883>
- Eidi A, Eidi M, Givianrad MH, Abaspour N. 2009. Hypolipidemic effects of alcoholic extract of eucalyptus (*Eucalyptus globulus* Labill.) leaves on diabetic and non-diabetic rats. **J Diabetes Metab Disord** 8: 105 - 112.
- Ekanem AP, Obiekezie A, Kloas W, Knopf K. 2004. Effects of crude extracts of *Mucuna pruriens* (Fabaceae) and *Carica papaya* (Caricaceae) against the protozoan fish parasite *Ichthyophthirius multifiliis*. **Parasitol Res** 92: 361 - 366. <https://doi.org/10.1007/s00436-003-1038-8>
- El-banna H, Latif A, Soliman M. 2013. Anticoccidial activity of *Allium sativum* and *Aloe Vera* in broiler chickens. **Int J Agro Vet Med Sci** 7: 117 - 125.
- El Sayed AM, Ezzat SM, El Naggar MM, El Hawary SS. 2016. In vivo diabetic wound healing effect and HPLC-DAD-ESI-MS/MS profiling of the methanol extracts of eight *Aloe* species. **Rev Bras Farmacogn** 26: 352 - 362. <https://doi.org/10.1016/j.bjp.2016.01.009>
- Elisabetsky E, Posey DA. 1989. Use of contraceptive and related plants by the Kayapo Indians (Brazil). **J Ethnopharmacol** 26: 299 - 316. [https://doi.org/10.1016/0378-8741\(89\)90103-7](https://doi.org/10.1016/0378-8741(89)90103-7)
- Encarnação S, Mello-Sampayo C, Graça NAG, Catarino L, Silva IBM, Lima BS, Silva OMD. 2016. Total phenolic content, antioxidant activity and pre-clinical safety evaluation of an *Anacardium occidentale* stem bark Portuguese hypoglycemic traditional herbal preparation. **Ind Crops Prod** 82: 171 - 178. <https://doi.org/10.1016/j.indcrop.2015.11.001>
- Escribano J, Pedreño MA, García-Carmona F, Muñoz R. 1998. Characterization of the antiradical activity of betalains from *Beta vulgaris* L. roots. **Phytochem Anal** 9: 124 - 127. [https://doi.org/10.1002/\(SICI\)1099-1565\(199805/06\)9:3<124::AID-PCA401>3.0.CO;2-0](https://doi.org/10.1002/(SICI)1099-1565(199805/06)9:3<124::AID-PCA401>3.0.CO;2-0)
- Eshghi F, Hosseini-mehr SJ, Rahmani N, Khademloo M, Norozi MS, Hojati O. 2010. Effects of *Aloe vera* cream on posthemorrhoidectomy pain and wound healing: Results of a randomized, blind, placebo-control study. **J Altern Complement Med** 16: 647 - 650. <https://doi.org/10.1089/acm.2009.0428>
- Etzler ME, Branstrator ML. 1974. Differential localization of cell surface and secretory components in rat intestinal epithelium by use of lectins. **J Cell Biol** 62: 329 - 343. <https://doi.org/10.1083/jcb.62.2.329>
- Ferraz AC, Angelucci MEM, Costa ML, Batista IR, Oliveira BH, Cunha C. 1999. Pharmacological evaluation of ricinine, a central nervous system stimulant isolated from *Ricinus communis*. **Pharmacol Biochem Behav**

- 63: 367 - 375. [https://doi.org/10.1016/S0091-3057\(99\)00007-6](https://doi.org/10.1016/S0091-3057(99)00007-6)
- Ferrucci MS. 2000. **Revision taxonómica de los géneros *Cardiospermum* y *Urvillea* para el neotropico (Sapindaceae)**. Tesis, Universidad Nacional de Córdoba, Córdoba, Argentina.
- Figueiredo FJ, LimaVLG. 2015. Antioxidant activity of anthocyanins from quixabeira (*Sideroxylon obtusifolium*) fruits. **Rev Bras Plant Med** 17: 473 - 479. https://doi.org/10.1590/1983-084x/14_005
- Franca F, Cuba CAC, Moreira EA, Miguel O, Almeida M, Das Virgens LM, Marsden PD. 1993. Avaliacao do efeito do extratao de casca de Cajueiro-Branco (*Anacardium Occidentale* L.) sobre a infeccao por *Leishmania* (*Viannia*) *brasiliensis*. **Rev Soc Bras Med Trop** 26: 151 - 155. <https://doi.org/10.1590/S0037-86821993000300004>
- FUNAI. 2010. Fundação Nacional do Índio. **Quem são os índios no Brasil**. <http://www.funai.gov.br/index.php/indios-no-brasil/quem-sao>
- Futija A, Migita M, Ueda T, Fukunaga Y. 2008. A 2-year-old Girl with *Diphyllobothrium nihonkaiense* infection treated with pral praziquantel and a laxative. **J Nippon Med School** 75: 225 - 227. <https://doi.org/10.1272/jnms.75.225>
- Galang MGM, Macabeo APG, Chang WC, Isobe M, Aguinaldo MAM. 2016. Glucosides from the unripe fruit juice of *Carica papaya* Linn. (Caricaceae) cultivar “Red Lady” with antioxidant activity. **J Funct Foods** 22: 358 - 362. <https://doi.org/10.1016/j.jff.2016.01.024>
- Ganesan K, Sehgal PK, Mandal AB, Sayeed S. 2011. Protective effect of withania somnifera and *Cardiospermum halicacabum* extracts against collagenolytic degradation of collagen. **Appl Biochem Biotechnol** 165: 1075 - 1091. <https://doi.org/10.1007/s12010-011-9326-8>
- Gbadegesin MA, Odunola OA, Akinwumi KA, Osifeso OO. 2009. Comparative hepatotoxicity and clastogenicity of sodium arsenite and three petroleum products in experimental swiss Albino Mice: The modulatory effects of *Aloe vera* gel. **Food Chem Toxicol** 47: 2454 - 2457. <https://doi.org/10.1016/j.fct.2009.07.002>
- Ghosh S, Tiwari SS, Srivastava S, Sharma AK, Kumar S, Ray DD, Rawat, AKS. 2013. Acaricidal properties of *Ricinus communis* leaf extracts against organophosphate and pyrethroids resistant *Rhipicephalus (Boophilus) microplus*. **Vet Parasitol** 192: 259 - 267. <https://doi.org/10.1016/j.vetpar.2012.09.031>
- Goel RK, Sairam K, Rao CV, Raman A. 2001. Role of gastric antioxidant and anti-Helicobacter pylori activities in antiulcerogenic activity of plantain banana (*Musa sapientum* var. paradisiaca). **Indian J Exp Biol** 39: 719 - 722.
- Govindarajan M, Sivakumar, R. 2012. Repellent properties of *Cardiospermum halicacabum* Linn. (Family: Sapindaceae) plant leaf extracts against three important vector mosquitoes. **Asian Pac J Trop Biomed** 2: 602 - 607. [https://doi.org/10.1016/S2221-1691\(12\)60105-1](https://doi.org/10.1016/S2221-1691(12)60105-1)
- Güner R, Hasanoglu I, Kayaaslan B, Aypak A, Akinci E, Bodur H, Eser F, Kalem AK, Küçükşahin O, Ateş I, Bastug A, Tekçe TY, Bilgiç Z, Gürsoy FM, Akçal HN, İzdes S, Erdem D, Asfuroğlu E, Hezer H, Kilic H, Cıvak, M, Aydoğan S, Buzgan T. 2021. Comparing ICU admission rates of mild/moderate COVID-19 patients treated with hydroxychloroquine, favipiravir, and hydroxychloroquine plus favipiravir. **J Infect Public Health** 14: 365 - 370. <https://doi.org/10.1016/j.jiph.2020.12.017>
- Gupta N, Verma K, Nalla S, Kulshreshtha A, Lall R, Prasad S. 2020. Free radicals as a double-edged sword: The cancer preventive and therapeutic roles of curcumin. **Molecules** 25: 1 - 20. <https://doi.org/10.3390/molecules25225390>
- Gurung S, Škalco-Basnet N. 2009. Wound healing properties of *Carica papaya* latex: *In vivo* evaluation in mice burn model. **J Ethnopharmacol** 121: 338 - 341. <https://doi.org/10.1016/j.jep.2008.10.030>
- Gutiérrez-Gutiérrez F, Palomo-Ligas L, Hernández-Hernández JM, Pérez-Rangel A, Aguayo-Ortiz R, Hernández-Campos A, Castillo R, González-Pozos S, Cortés-Zárate R, Ramírez-Herrera MA, Mendoza-Magaña ML, Castillo-Romero A. 2017. Curcumin alters the cytoskeleton and microtubule organization on trophozoites of *Giardia lamblia*. **Acta Trop** 172: 113 - 121. <https://doi.org/10.1016/j.actatropica.2017.04.027>
- Güven M, Gölge UH, Aslan E, Sehitoglu MH, Aras AB, Akman T, Cosar M. 2016. The effect of *Aloe vera* on ischemia-reperfusion injury of sciatic nerve in rats. **Biomed Pharmacother** 79: 201 - 207. <https://doi.org/10.1016/j.biopha.2016.02.023>
- Haddad M, Sauvain M, Deharo E. 2011. Curcuma as a parasiticidal agent: A review. **Planta Med** 77: 672 - 678.

- <https://doi.org/10.1055/s-0030-1250549>
- Haritha K, Ramesh B, Saralakumari D. 2014. Effect of *Aloe vera* gel on antioxidant enzymes in streptozotocin-induced cataractogenesis in male and female Wistar rats. **J Acute Med** 4: 38 - 44.
<https://doi.org/10.1016/j.jacme.2014.01.005>
- Hasimun P, Suwendar, Ernasari GI. 2014. Analgetic activity of papaya (*Carica papaya* L.) leaves extract. **Procedia Chem** 13: 147 - 149. <https://doi.org/10.1016/j.proche.2014.12.019>
- Hossain E. 2018. Ethno-Helminthology - Newer perspective for discovery of newer anthelmintic drugs. **Madridge J Nov Drug Res** 2: 35 - 36. <https://doi.org/10.18689/mjndr-1000106>
- Huang MH, Huang SS, Wang BS, Wu CH, Sheu MJ, Hou WC, Lin SS, Huang GJ. 2011. Antioxidant and anti-inflammatory properties of *Cardiospermum halicacabum* and its reference compounds *ex vivo* and *in vivo*. **J Ethnopharmacol** 133: 743 - 750. <https://doi.org/10.1016/j.jep.2010.11.005>
- Hussain A, Khan MN, Sajid MS, Iqbal Z, Khan MK, Abbas RZ, Raza MA, Needham GR. 2010. *In vitro* screening of the leaves of *Musa paradisiaca* for anthelmintic activity. **J Anim Plant Sci** 20: 5 - 8.
- Hussain A, Khan MN, Iqbal Z, Sajid MS, Khan MK. 2011. Anthelmintic activity of *Trianthema portulacastrum* L. and *Musa paradisiaca* L. against gastrointestinal nematodes of sheep. **Vet Parasitol** 179: 92 - 99.
<https://doi.org/10.1016/j.vetpar.2011.02.022>
- IBGE. 2010. (Instituto Brasileiro de Geografia e Estatística). **Indígenas**. <https://indigenas.ibge.gov.br>
- Iglesias R, Citores L, Di Maro A, Ferreras JM. 2015. Biological activities of the antiviral protein BE27 from sugar beet (*Beta vulgaris* L.). **Planta** 241: 421 - 433. <https://doi.org/10.1007/s00425-014-2191-2>
- Ikawati Z, Wahyuono S, Maeyama K. 2001. Screening of several Indonesian medicinal plants for their inhibitory effect on histamine release from RBL-2H3 cells. **J Ethnopharmacol** 75: 249 - 256.
[https://doi.org/10.1016/S0378-8741\(01\)00201-Xv](https://doi.org/10.1016/S0378-8741(01)00201-Xv)
- Ilavarasan R, Mallika M, Venkataraman S. 2006. Anti-inflammatory and free radical scavenging activity of *Ricinus communis* root extract. **J Ethnopharmacol** 103: 478 - 480. <https://doi.org/10.1016/j.jep.2005.07.029>
- Imam MZ, Akter S. 2011. *Musa paradisiaca* L. and *Musa sapientum* L.: A phytochemical and pharmacological review. **J Appl Pharm Sci** 1: 14 - 20.
- Iroaganachi M, Eleazu CO, Okafor PN, Nwaohu N. 2015. Effect of unripe plantain (*Musa paradisiaca*) and ginger (*Zingiber officinale*) on renal dysfunction in streptozotocin-induced diabetic rats. **JOP** 9: 1 - 6.
<https://doi.org/10.6092/1590-8577/2954>
- ISA. 2018. (Instituto Socioambiental). **Povos Indígenas no Brasil: Kantaruré**.
<https://pib.socioambiental.org/pt/Povo:Kantarur%C3%A9>
- Jain S, Garg VK, Sharma PK. 2011. Anti-Infl ammatory activity of aqueous extract of *Beta vulgaris* L. **J Basic Clin Pharm** 2: 83 - 86.
- Jaiprakash B, Habbu PV, Karadi RV, Lavalhe MS, Savadi, RV, Sudha R, Joshi SD. 2006. Antihypertensive activity of Amuri from *Musa paradisiaca* L. **Indian J Trad Knowledge** 5: 197 - 200.
- Jaiswal P, Singh DK. 2008. Molluscicidal activity of *Carica papaya* and *Areca catechu* against the freshwater snail *Lymnaea acuminata*. **Vet Parasitol** 152: 264 - 270. <https://doi.org/10.1016/j.vetpar.2007.12.033>
- Jandú JJB, Silva LCN, Pereira APC, Souza RM, Silva Júnior CA, Figueiredo RCBQ, Araújo JM, Correia MTS, Silva MV. 2013. *Myracrodruon urundeuva* bark: an antimicrobial, antioxidant and non-cytotoxic agent. **J Med Plant Res** 7: 413 - 418. <https://doi.org/10.5897/JMPR12.681>
- Jawla S, Kumar Y, Khan MSY. 2012. Antimicrobial and antihyperglycemic activities of *Musa paradisiaca* flowers. **Asian Pac J Trop Biomed** 2: 914 - 918. [https://doi.org/10.1016/S2221-1691\(12\)60336-0](https://doi.org/10.1016/S2221-1691(12)60336-0)
- Jayaveera KN, Siddaiah M, Swamy BMV. 2009. Hepatoprotective activity of *Ximenia americana* against carbon tetrachloride induced hepatic damage in rats. **Int J Infect Dis** 13: S58.
[https://doi.org/10.1016/s1201-9712\(09\)60422-7](https://doi.org/10.1016/s1201-9712(09)60422-7)
- Jeyadevi R, Sivasudha T, Ilavarasi A, Thajuddin N. 2013. Chemical constituents and antimicrobial activity of indian green leafy vegetable *Cardiospermum halicacabum*. **Indian J Microbiol** 53: 208 - 213.
<https://doi.org/10.1007/s12088-012-0333-4>
- Jomsky MG, Kerna NA. 2020. Could low-dose quinine prevent or treat coronavirus infection? **EC Pharmacol Toxicol** 8: 62 - 64.

- Jong MC, Ermuth U, Augustin M. 2013. Plant-based ointments versus usual care in the management of chronic skin diseases: A comparative analysis on outcome and safety. **Complement Ther Med** 21: 453 - 459. <https://doi.org/10.1016/j.ctim.2013.07.002>
- Jordan GJ, Potts BM, Kirkpatrick JB, Gardiner C. 1993. Variation in the *Eucalyptus globulus* complex revisited. **Aust J Bot** 41: 763 - 785. <https://doi.org/10.1071/BT9930763>
- Juárez-Rojop IE, Díaz-Zagoya JC, Ble-Castillo JL, Miranda-Orsorio PH, Castell-Rodríguez AE, Tovilla-Zárate CA, Rodríguez-Hernández A, Aguilar-Mariscal H, Ramón-Frías T, Bermúdez-Ocaña, DY. 2012. Hypoglycemic effect of *Carica papaya* leaves in streptozotocin-induced diabetic rats. **BMC Complement Altern Med** 12: 1 - 11. <https://doi.org/10.1186/1472-6882-12-236>
- Jumba BN, Anjili CO, Makwali J, Ingonga J, Nyamano R, Marango S, Choge JK, Khayeka-Wandabwa C. 2015. Evaluation of leishmanicidal activity and cytotoxicity of *Ricinus communis* and *Azadirachta indica* extracts from western Kenya: *In vitro* and *in vivo* assays. **BMC Res Notes** 8: 1 - 11. <https://doi.org/10.1186/s13104-015-1605-y>
- Karbach F, Mouhouche F, Fleurat-Lessard F. 2011. Deterrent and insecticidal properties of bean seed (*Phaseolus vulgaris* L.) whole meal or protein extract incorporated into the diet of *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). **J Stored Prod Res** 47: 197 - 203. <https://doi.org/10.1016/j.jspr.2011.02.002>
- Karuppiiah P, Mustaffa M. 2013. Antibacterial and antioxidant activities of *Musa* sp. leaf extracts against multidrug resistant clinical pathogens causing nosocomial infection. **Asian Pac J Trop Biomed** 3: 737 - 742. [https://doi.org/10.1016/S2221-1691\(13\)60148-3](https://doi.org/10.1016/S2221-1691(13)60148-3)
- Kazmierczak R, Hallmann E, Lipowski J, Drela N, Kowalik A, Püssa T, Matt D, Luik A, Gozdowski D, Rembiałkowska E. 2014. Beetroot (*Beta vulgaris* L.) and naturally fermented beetroot juices from organic and conventional production: Metabolomics, antioxidant levels and anticancer activity. **J Sci Food Agric** 94: 2618 - 2629. <https://doi.org/10.1002/jsfa.6722>
- Kebebew Z, Shibeshi W. 2013. Evaluation of anxiolytic and sedative effects of 80% ethanolic *Carica papaya* L. (Caricaceae) pulp extract in mice. **J Ethnopharmacol** 150: 665 - 671. <https://doi.org/10.1016/j.jep.2013.09.023>
- Kermanshah R, McCarry, BE, Rosenfeld J, Summers PS, Weretilnyk EA, Sorger GJ. 2001. Benzyl isothiocyanate is the chief or sole anthelmintic in papaya seed extracts. **Phytochemistry** 57: 427 - 435. [https://doi.org/10.1016/S0031-9422\(01\)00077-2](https://doi.org/10.1016/S0031-9422(01)00077-2)
- Kew Science. 2020. *Beta vulgaris* L. <http://powo.science.kew.org/taxon/164505-1>
- Khan IN, Habib MR, Rahman MM, Mannan A, Sarker MMI, Hawlader S. 2011. Thrombolytic potential of *Ocimum sanctum* L., *Curcuma longa* L., *Azadirachta indica* L. and *Anacardium occidentale* L. **J Basic Clin Pharm** 5: 125 - 127. <https://doi.org/10.4103/0976-0105.141942>
- Khan KS, Kunz R, Kleijnen J, Antes G. 2003. Five steps to conducting a systematic review. **J Royal Soc Med** 96: 118 - 121. <https://doi.org/10.1258/jrsm.96.3.118>
- Khan T, Khan MA, Mashwani Z, Ullah N, Nadhman A. 2021. Therapeutic potential of medicinal plants against COVID-19 : The role of antiviral medicinal metabolites. **Biocatal Agric Biotechnol**. 31.
- Khunkitti W, Fujimaki Y, Aoki Y. 2000. *In vitro* antifilarial activity of extracts of the medicinal plant *Cardiospermum halicacabum* against *Brugia pahangi*. **J Helminthol** 74: 241 - 246. <https://doi.org/10.1017/S0022149X00000342>
- Kirkpatrick JB. 1974. The numerical intraspecific taxonomy of *Eucalyptus globulus* Labill. (Myrtaceae). **Bot J Linn Soc** 69: 89 - 104. <https://doi.org/10.1111/j.1095-8339.1974.tb01618.x>
- Kliks MM. 1985. Studies on the traditional herbal anthelmintic *Chenopodium ambrosioides* L.: Ethnopharmacological evaluation and clinical field trials. **Soc Sci Med** 21: 879 - 886. [https://doi.org/10.1016/0277-9536\(85\)90144-3](https://doi.org/10.1016/0277-9536(85)90144-3)
- Klionsky DJ, Abdel-Aziz AK, Abdelfatah S, Abdellatif M, Abdoli A, Abel S, Abeliovich H, Abildgaard MH, Abudu YP, Acevedo-Arozena A, Adamopoulos IE, Adeli K, Adolph TE, Adornetto A, Aflaki E, Agam G, Agarwal A, Aggarwal BB, Agnello M, Agostinis P, Agrewala JN, Agrotis A, Aguilar PV, Ahmad ST, Ahmed ZM, Ahumada-Castro U, Aits S, Aizawa S, Akkoc Y, Akoumianaki T, Akpinar HA, Al-Abd AM, Al-Akra L, Al-Gharaibeh A, Alaoui-Jamali MA, Alberti S, Alcocer-Gómez E, Alessandri C, Ali M, Al-Bari

MAA, Aliwaini S, Alizadeh J, Almacellas E, Almasan A, Alonso A, Alonso GD, Altan-Bonnet N, Altieri DC, Álvarez EMC, Alves S, Costa CA, Alzaharna MM, Amadio M, Amantini C, Amaral C, Ambrosio S, Amer AO, Ammanathan V, An Z, Andersen SU, Andrabi SA, Andrade-Silva M, Andres MM, Angelini S, Ann D, Anozie UC, Ansari MY, Antas P, Antebi A, Antón Z, Anwar T, Apetoh L, Apostolova N, Araki Y, Araki Y, Arasaki K, Araújo WL, Araya J, Arden C, Arévalo MA, Arguelles S, Arias E, Arikath J, Arimoto H, Arosia AR, Armstrong-James D, Arnauné-Pelloquin D, Aroca A, Arroyo DS, Arsov I, Artero R, Asaro DML, Aschner M, Ashrafizadeh M, Ashur-Fabian O, Atanasov AG, Au AK, Auberger P, Auner HW, Aurelian L, Autelli R, Avagliano L, Ávalos Y, Aveic S, Aveleira CA, Avin-Wittenberg T, Aydin Y, Ayton S, Ayyadevara S, Azzopardi M, Baba M, Backer JM, Backues SK, Bae DH, Bae ON, Bae SH, Baehrecke EH, Baek A, Baek SH, Baek SH, Bagetta G, Bagniewska-Zadworna A, Bai H, Bai J, Bai X, Bai Y, Bairagi N, Bakshi S, Balbi T, Baldari CT, Balduini W, Ballabio A, Ballester M, Balazadeh S, Balzan R, Bandopadhyay R, Banerjee S, Banerjee S, Bánréti A, Bao Y, Baptista MS, Baracca A, Barbat C, Bargiela A, Barilà D, Barlow PG, Barmada SJ, Barreiro E, Barreto GE, Bartek J, Bartel B, Bartolome A, Barve GR, Basagoudanavar SH, Bassham DC, Bast RC, Basu A, Batoko H, Batten I, Baulieu EE, Baumgarner BL, Bayry J, Beale R, Beau I, Beaumatin F, Bechara LRG, Beck GR, Beers MF, Begun J, Behrends C, Behrens GMN, Bei R, Bejarano E, Bel S, Behl C, Belaid A, Touzé NB, Bellarosa C, Belleudi F, Pérez MB, Bello-Morales R, Beltran JSO, Beltran S, Benbrook DM, Bendorius M, Benitez BA, Cuesta IB, Bensalem J, Berchtold MB, Berezowska S, Bergamaschi D, Bergami M, Bergmann A, Berliocchi L, Berlioz-Torrent C, Bernard A, Berthouix L, Besirli CG, Besteiro S, Betin VM, Beyaert R, Bezbradica JS, Bhaskar K, Bhatia-Kissova I, Bhattacharya R, Bhattacharya S, Bhattacharyya S, Bhuiyan MS, Bhutia SK, Bi L, Bi X, Biden TJ, Bijian K, Billes VA, Binart N, Bincoletto C, Birgisdottir AB, Bjorkoy G, Blanco G, Blas-Garcia A, Blasiak J, Blomgran R, Blomgren K, Blum JS, Boada-Romero E, Boban M, Boesze-Battaglia K, Boeuf P, Boland B, Bomont P, Bonaldo P, Bonam SR, Bonfili L, Bonifacino JS, Boone BA, Bootman MD, Bordi M, Borner C, Bornhauser BC, Borthakur G, Bosch J, Bose S, Botana LM, Botas J, Boulanger CM, Boulton ME, Bourdenx M, Bourgeois B, Bourke NM, Bousquet G, Boya P, Bozhkov PV, Bozi LHM, Bozkurt TO, Brackney DE, Brandts CH, Braun RJ, Braus GH, Bravo-Sagua R, Bravo-San Pedro JM, Brest P, Bringer MA, BrionesHerrera A, Broaddus VC, Brodersen P, Brodsky JL, Brody SL, Bronson PG, Bronstein JM, Brown CN, Brown RE, Brum PC, Brumell JH, Brunetti-Pierri N, Bruno D, Bryson-Richardson RJ, Bucci C, Buchrieser C, Bueno M, Buitrago-Molina LE, Buraschi S, Buch S, Buchan JR, Buckingham EM, Budak H, Budini M, Bultynck G, Burada F, Burgoyne JR, Burón MI, Bustos V, Büttner S, Butturini E, Byrd A, Cabas I, CabreraBenitez S, Cadwell K, Cai J, Cai L, Cai Q, Cairó M, Calbet JA, Caldwell GA, Caldwell KA, Call JA, Calvani R, Calvo AC, Calvo-Rubio Barrera M, Camara NOS, Camonis JH, Camougrand N, Campanella M, Campbell EM, Campbell-Valois FX, Campello S, Campesi I, Campos JC, Camuzard O, Cancino J, Almeida DC, Canesi L, Caniggia I, Canonico B, Cantí C, Cao B, Caraglia M, Caramés B, Carchman EH, Cardenal-Muñoz E, Cardenas C, Cardenas L, Cardoso SM, Carew JS, Carle GF, Carleton G, Carloni S, Carmona-Gutierrez D, Carneiro LA, Carnevali O, Carosi JM, Carra S, Carrier A, Carrier L, Carroll B, Carter AB, Carvalho AN, Casanova M, Casas C, Casas J, Cassioli C, Castillo EF, Castillo K, Castillo-Lluva S, Castoldi F, Castori M, Castro AF, Castro-Caldas M, Castro-Hernandez J, Castro-Obregon S, Catz SD, Cavadas C, Cavaliere F, Cavallini G, Cavinato M, Cayuela ML, Rica PC, Cecarini V, Cecconi F, Cechowska-Pasko M, Cenci S, Ceperuelo-Mallafré V, Cerqueira JJ, Cerutti JM, Cervia D, Bozok Cetintas V, Cetrullo S, Chae HJ, Chagin AS, Chai CY, Chakrabarti G, Chakrabarti O, Chakraborty T, Chakraborty T, Chami M, Chamilos G, Chan DW, Chan EYW, Chan ED, Chan HYE, Chan HH, Chan H, Chan MTV, Chan YS, Chandra PK, Chang CP, Chang C, Chang HC, Chang K, Chao J, Chapman T, Charlet-Berguerand N, Chatterjee S, Chaube SK, Chaudhary A, Chauhan S, Chaum E, Checler F, Cheetham ME, Chen CS, Chen GC, Chen JF, Chen LL, Chen L, Chen L, Chen M, Chen MK, Chen N, Chen Q, Chen RH, Chen S, Chen W, Chen W, Chen XM, Chen XW, Chen X, Chen Y, Chen YG, Chen Y, Chen Y, Chen YJ, Chen YQ, Chen ZS, Chen Z, Chen ZH, Chen ZJ, Chen Z, Cheng H, Cheng J, Cheng SY, Cheng W, Cheng X, Cheng XT, Cheng Y, Cheng Z, Chen Z, Cheong H, Cheong JK, Chernyak BV, Cherry S, Cheung CFR, Cheung CHA, Cheung KH, Chevet E, Chi RJ, Chiang AKS, Chiaradonna F, Chiarelli R, Chiariello M, Chica N, Chiocca S, Chiong M, Chiou SH, Chiramel AI, Chiurchiù V, Cho DH, Choe SK,

Choi AMK, Choi ME, Choudhury KR, Chow NS, Chu CT, Chua JP, Chua JJE, Chung H, Chung KP, Chung S, Chung SH, Chung YL, Cianfanelli V, Ciechomska IA, Cifuentes M, Cinque L, Cirak S, Cirone M, Clague MJ, Clarke R, Clementi E, Coccia EM, Codogno P, Cohen E, Cohen MM, Colasanti T, Colasuonno F, Colbert RA, Colell A, Čolić M, Coll NS, Collins MO, Colombo MI, Colón-Ramos DA, Combaret L, Comincini S, Cominetti MR, Consiglio A, Conte A, Conti F, Contu VR, Cookson MR, Coombs KM, Coppens I, Corasaniti MT, Corkery DP, Cordes N, Cortese K, Costa MC, Costantino S, Costelli P, Coto-Montes A, Crack PJ, Crespo JL, Criollo A, Crippa V, Cristofani R, Csizmadia T, Cuadrado A, Cui B, Cui J, Cui Y, Cui Y, Culetto E, Cumino AC, Cybulsky AV, Czaja MJ, Czuczwar SJ, D'Adamo S, D'Amelio M, D'Arcangelo D, D'Lugos AC, D'Orazi G, Silva JA, Dafsari HS, Dagda RK, Dagdas Y, Daglia M, Dai X, Dai Y, Dai Y, Col JD, Dalhaimer P, Valle LD, Dallenga T, Dalmasso G, Damme M, Dando I, Dantuma NP, Darling AL, Das H, Dasarathy S, Dasari SK, Dash S, Daumke O, Dauphinee AN, Davies JS, Dávila VA, Davis RJ, Davis T, Naidu SD, de Amicis F, De Bosscher K, De Felice F, De Franceschi L, De Leonibus C, Barbosa MGM, de Meyer GRG, De Milito A, De Nunzio C, De Palma C, De Santi M, De Virgilio C, De Zio D, Debnath J, De Bosch BJ, Decuypere JP, Deehan MA, Deflorian G, De Gregori J, Dehay B, del Rio G, Delaney JR, Delbridge LMD, Delorme-Axford E, Delpino MV, Demarchi F, Dembitz V, Demers ND, Deng H, Deng Z, Dengjel J, Dent P, Denton D, De Pamphilis ML, Der CJ, Deretic V, Descoteaux A, Devis L, Devkota S, Devuyst O, Dewson G, Dharmasivam M, Dhiman R, di Bernardo D, Di Cristina M, Di Domenico F, Di Fazio P, Di Fonzo A, Di Guardo G, Di Guglielmo GM, Di Leo L, Di Malta C, Di Nardo A, Di Rienzo M, Di Sano F, Diallinas G, Diao J, Diaz-Araya G, Díaz-Laviada I, Dickinson JM, Diederich M, Dieudé M, Dikic I, Ding S, Ding WX, Dini L, Dinić J, Dinic M, Dinkova-Kostova AT, Dionne MC, Distler JHW, Diwan A, Dixon IMC, Djavaheri-Mergny M, Dobrinski I, Dobrovinskaya O, Dobrowolski R, Dobson RCJ, Đokić J, Emre SD, Donadelli M, Dong B, Dong X, Dong Z, Dorn GW, Dotsch V, Dou H, Dou J, Dowaidar M, Dridi S, Drucker L, Du A, Du C, Du G, Du HN, Du LL, du Toit A, Duan SB, Duan X, Duarte SP, Dubrovskaya A, Dunlop EA, Dupont N, Durán RV, Dwarakanath BS, Dyshlovoy SA, Fakhari DE, Eckhart L, Edelstein CL, Efferth T, Eftekharpour E, Eichinger L, Eid N, Eisenberg T, Eissa NT, Eissa S, Ejarque M, El Andaloussi A, El-Hage N, El-Naggar S, Eleuteri AM, El-Shafey ES, Elgendy M, Eliopoulos AG, Elizalde MM, Elks PM, Elsasser HP, Elsherbiny ES, Emerling BM, Emre NCT, Eng CH, Engedal N, Engelbrecht AM, Engelsen AST, Enserink JM, Escalante R, Escatine A, Escobar-Henriques M, Eskelinen EL, Espert L, Eusebio MO, Fabrias G, Fabrizi C, Facchiano A, Facchiano F, Fadeel B, Fader C, Faesen AC, Fairlie WD, Falcó A, Falkenburger BH, Fan D, Fan J, Fan Y, Fang EF, Fang Y, Fang Y, Fanto M, Farfel-Becker T, Faure M, Fazeli G, Fedele AO, Feldman AM, Feng D, Feng J, Feng L, Feng Y, Feng Y, Feng W, Araujo TF, Ferguson TA, Fernández AF, Fernandez-Checa JC, Fernández-Veledo S, Fernie AR, Ferrante AW, Ferraresi A, Ferrari MF, Ferreira JCB, Ferro-Novick S, Figueras A, Filadi R, Filigheddu N, Filippi-Chiela E, Filomeni G, Fimia GM, Fineschi V, Finetti F, Finkbeiner S, Fisher EA, Fisher PB, Flamigni F, Fliesler SJ, Flo TH, Florance I, Florey O, Florio T, Fodor E, Follo C, Fon EA, Forlino A, Fornai F, Fortini P, Fracassi A, Fraldi A, Franco B, Franco R, Franconi F, Frankel LB, Friedman SL, Fröhlich LF, Frühbeck G, Fuentes JM, Fujiki Y, Fujita N, Fujiwara Y, Fukuda M, Fulda S, Furic L, Furuya N, Fusco C, Gack MU, Gaffke L, Galadari S, Galasso A, Galindo MF, Kankanamalage SG, Galluzzi L, Galy V, Gammoh N, Gan B, Ganley IG, Gao F, Gao H, Gao M, Gao P, Gao SJ, Gao W, Gao X, Garcera A, Garcia MN, Garcia VE, García-Del Portillo F, Garcia-Escudero V, Garcia-Garcia A, Garcia-Macia M, García-Moreno D, Garcia-Ruiz C, García-Sanz P, Garg AD, Gargini R, Garofalo T, Garry RF, Gassen NC, Gatica D, Ge L, Ge W, Geiss-Friedlander R, Gelfi C, Genschik P, Gentle IE, Gerbino V, Gerhardt C, Germain K, Germain M, Gewirtz DA, Afshar EG, Ghavami S, Ghigo A, Ghosh M, Giamas G, Giampietri C, Giatromanolaki A, Gibson GE, Gibson SB, Ginet V, Giniger E, Giorgi C, Girao H, Girardin SE, Giridharan M, Giuliano S, Giulivi C, Giuriato S, Giustiniani J, Gluschnko A, Goder V, Goginashvili A, Golab J, Goldstone DC, Golebiewska A, Gomes LR, Gomez R, Gómez-Sánchez R, Gomez-Puerto MC, Gomez-Sintes R, Gong Q, Goni FM, González-Gallego J, Gonzalez-Hernandez T, Gonzalez-Polo RA, Gonzalez-Reyes JA, González-Rodríguez P, Goping IS, Gorbatyuk MS, Gorbunov NV, Görgülü K, Gorjod RM, Gorski SM, Goruppi S, Gotor C, Gottlieb RA, Gozes I, Gozuacik D, Graef M, Gräler MH, Granatiero V, Grasso D, Gray JP, Green DR, Greenhough A,

Gregory SL, Griffin EF, Grinstaff MW, Gros F, Grose C, Gross AS, Gruber F, Grumati P, Grune T, Gu X, Guan JL, Guardia CM, Guda K, Guerra F, Guerri C, Guha P, Guillén C, Gujar S, Gukovskaya A, Gukovsky I, Gunst J, Günther A, Guntur AR, Guo C, Guo C, Guo H, Guo LW, Guo M, Gupta P, Gupta SK, Gupta S, Gupta VB, Gupta V, Gustafsson AB, Gutterman DD, Ranjitha HB, Haapasalo A, Haber JE, Hać A, Hadano S, Hafrén AJ, Haidar M, Hall BS, Halldén G, Hamacher-Brady A, Hamann A, Hamasaki M, Han W, Hansen M, Hanson PI, Hao Z, Harada M, Harhaji-Trajkovic L, Hariharan N, Haroon N, Harris J, Hasegawa T, Nagoor NH, Haspel JA, Haucke V, Hawkins WD, Hay BA, Haynes CM, Hayrabydyan SB, Hays TS, He C, He Q, He RR, He YW, He YY, Heakal Y, Heberle AM, Hejtmancik JF, Helgason GV, Henkel V, Herb M, Hergovich A, Herman-Antosiewicz A, Hernández A, Hernandez C, Hernandez-Diaz S, Hernandez-Gea V, Herpin A, Herreros J, Hervás JH, Hesselson D, Hetz C, Heussler VT, Higuchi Y, Hilfiker S, Hill JA, Hlavacek WS, Ho EA, Ho YHT, Ho PWL, Ho SL, Ho WY, Hobbs GA, Hochstrasser M, Hoet PHM, Hofius D, Hofman P, Höhn A, Holmberg CI, Hombrebueno JR, Hong CW, Hong YR, Hooper LV, Hoppe T, Horos R, Hoshida Y, Hsin IL, Hsu HY, Hu B, Hu D, Hu LF, Hu MC, Hu R, Hu W, Hu YC, Hu ZW, Hua F, Hua J, Hua Y, Huan C, Huang C, Huang C, Huang C, Huang C, Huang H, Huang K, Huang MLH, Huang R, Huang S, Huang T, Huang X, Huang YJ, Huber TB, Hubert V, Hubner CA, Hughes SM, Hughes WE, Humbert M, Hummer G, Hurley JH, Hussain S, Hussain S, Hussey PJ, Hutabarat M, Hwang HY, Hwang S, Ieni A, Ikeda F, Imagawa Y, Imai Y, Imbriano C, Imoto M, Inman DM, Inoki K, Iovanna J, Iozzo RV, Ippolito G, Irazoqui JE, Iribarren P, Ishaq M, Ishikawa M, Ishimwe N, Isidoro C, Ismail N, Navikas SI, Itakura E, Ito D, Ivankovic D, Ivanova S, Iyer AKV, Izquierdo JM, Izumi M, Jäättelä M, Jabir MS, Jackson WT, Jacobo-Herrera N, Jacomin AC, Jacquin E, Jadiya P, Jaeschke H, Jagannath C, Jakobi AJ, Jakobsson J, Janji B, Jansen-Dürr P, Jansson PJ, Jantsch J, Januszewski S, Jassey A, Jean S, Jeltsch-David H, Jendelova P, Jenny A, Jensen TE, Jessen N, Jewell JL, Ji J, Jia L, Jia R, Jiang L, Jiang Q, Jiang R, Jiang T, Jiang X, Jiang Y, Jimenez-Sanchez M, Jin EJ, Jin F, Jin H, Jin L, Jin L, Jin M, Jin S, Jo EK, Joffre C, Johansen T, Johnson GVV, Johnston SA, Jokitalo E, Jolly MK, Joosten LAB, Jordan J, Joseph B, Ju D, Ju JS, Ju J, Juárez E, Judith D, Juhász G, Jun Y, Jung CH, Jung SC, Jung YK, Jungbluth H, Jungverdorben J, Just S, Kaarniranta K, Kaasik A, Kabuta T, Kaganovich D, Kahana A, Kajimura S, Kalamvoki M, Kalia M, Kalinowski DS, Kaludercic N, Kalvari I, Kaminska J, Kaminsky VO, Kanamori H, Kanasaki K, Kang C, Kang R, Kang SS, Kaniyappan S, Kanki T, Kanneganti TD, Kanthasamy AG, Kanthasamy A, Kantorow M, Kapuy O, Karamouzis MV, Karim MR, Karmakar P, Katore RG, Kato M, Kaufmann SHE, Kauppinen A, Kaushal GP, Kaushik S, Kawasaki K, Kazan K, Ke PY, Keating DJ, Keber U, Kehrl JH, Keller KE, Keller CW, Kemper JK, Kenific CM, Kepp O, Kermorgant S, Kern A, Ketteler R, Keulers TG, Khalfin B, Khalil H, Khambu B, Khan SY, Khandelwal VKM, Khandia R, Kho W, Khobreakar NV, Khuansuwan S, Khundadze M, Killackey SA, Kim D, Kim DR, Kim DH, Kim DE, Kim EY, Kim EK, Kim HR, Kim HS, Kim HR, Kim JH, Kim JK, Kim JH, Kim J, Kim JH, Kim KI, Kim PK, Kim SJ, Kimball SR, Kimchi A, Kimmelman AC, Kimura T, King MA, Kinghorn KJ, Kinsey CG, Kirkin V, Kirshenbaum LA, Kiselev SL, Kishi S, Kitamoto K, Kitaoka Y, Kitazato K, Kitsis RN, Kittler JT, Kjaerulff O, Klein PS, Klopstock T, Klucken J, Knævelsrud H, Knorr RL, Ko BCB, Ko F, Ko JL, Kobayashi H, Kobayashi S, Koch I, Koch JC, Koenig U, Kögel D, Koh YH, Koike M, Kohlwein SD, Kocaturk NM, Komatsu M, König J, Kono T, Kopp BT, Korcsmaros T, Korkmaz G, Korolchuk VI, Suárez-Korsnes M, Koskela A, Kota J, Kotake Y, Kotler ML, Kou Y, Koukourakis MI, Koustas E, Kovacs AL, Kovács T, Koya D, Kozako T, Kraft C, Krainc D, Krämer H, Krasnodembskaya AD, Kretz-Remy C, Kroemer G, Ktistakis NT, Kuchitsu K, Kuenen S, Kuerschner L, Kukar T, Kumar A, Kumar A, Kumar D, Kumar D, Kumar S, Kume S, Kumsta C, Kundu CN, Kundu M, Kunnumakkara AB, Kurgan L, Kutateladze TG, Kutlu O, Kwak SA, Kwon HJ, Kwon TK, Kwon YT, Kyrnizi I, La Spada A, Labonté P, Ladoire S, Laface I, Lafont F, Lagace DC, Lahiri V, Lai Z, Laird AS, Lakkaraju A, Lamark T, Lan SH, Landajuela A, Lane DJR, Lane JD, Lang CH, Lange C, Langel U, Langer R, Lapaquette P, Laporte J, LaRusso NF, Lastres-Becker I, Lau WCY, Laurie GW, Lavandero S, Law BYK, Law HKW, Layfield R, Le W, Le Stunff H, Leary AY, Lebrun JJ, Leck LYW, Leduc-Gaudet JP, Lee C, Lee CP, Lee DH, Lee EB, Lee EF, Lee GM, Lee HJ, Lee HK, Lee JM, Lee JS, Lee JA, Lee JY, Lee JH, Lee M, Lee MG, Lee MJ, Lee MS, Lee SY, Lee SJ, Lee SY, Lee SB, Lee WH, Lee YR, Lee Y, Lee Y, Lefebvre C, Legouis R, Lei YL, Lei Y, Leikin S,

Leitinger G, Lemus L, Leng S, Lenoir O, Lenz G, Lenz HJ, Lenzi P, León Y, Leopoldino AM, Leschczyk C, Leskelä S, Letellier E, Leung CT, Leung PS, Leventhal JS, Levine B, Lewis PA, Ley K, Li B, Li DQ, Li J, Li J, Li J, Li K, Li L, Li M, Li M, Li M, Li M, Li M, Li PL, Li MQ, Li Q, Li S, Li T, Li W, Li W, Li X, Li YP, Li Y, Li Z, Li Z, Li Z, Lian J, Liang C, Liang Q, Liang W, Liang Y, Liang YT, Liao G, Liao L, Liao M, Liao YF, Librizzi M, Lie PPY, Lilly MA, Lim HJ, Lima TRR, Limana F, Lin C, Lin CW, Lin DS, Lin FC, Lin JD, Lin KM, Lin KH, Lin LT, Lin PH, Lin Q, Lin S, Lin SJ, Lin W, Lin X, Lin YX, Lin YS, Linden R, Lindner P, Ling SC, Lingor P, Linnemann AK, Liou YC, Lipinski MM, Lipovšek S, Lira VA, Lisiak N, Liton PB, Liu C, Liu CH, Liu CF, Liu CH, Liu F, Liu H, Liu HS, Liu HF, Liu H, Liu J, Liu J, Liu J, Liu L, Liu L, Liu M, Liu Q, Liu W, Liu W, Liu XH, Liu X, Liu X, Liu X, Liu X, Liu Y, Liu Y, Liu Y, Liu Y, Liu Y, Livingston JA, Lizard G, Lizcano JM, Ljubojevic-Holzer S, Lleonart ME, Llobet-Navàs D, Llorente A, Lo CH, Lobato-Márquez D, Long Q, Long YC, Loos B, Loos JA, López MG, López-Doménech G, López-Guerrero JA, López-Jiménez AT, López-Pérez O, López-Valero I, Lorenowicz MJ, Lorente M, Lorincz P, Lossi L, Lotersztajn S, Lovat PE, Lovell JF, Lovy A, Löw P, Lu G, Lu H, Lu JH, Lu JJ, Lu M, Lu S, Luciani A, Lucocq JM, Ludovico P, Luftig MA, Luhr M, Luis-Ravelo D, Lum JJ, Luna-Dulcey L, Lund AH, Lund VK, Lünemann JD, Lüningschrör P, Luo H, Luo R, Luo S, Luo Z, Luparello C, Lüscher B, Luu L, Lyakhovich A, Lyamzaev KG, Lystad AH, Lytvynchuk L, Ma AC, Ma C, Ma M, Ma NF, Ma QH, Ma X, Ma Y, Ma Z, MacDougald OA, Macian F, MacIntosh GC, MacKeigan JP, Macleod KF, Maday S, Madeo F, Madesh M, Madl T, Madrigal-Matute J, Maeda A, Maejima Y, Magarinos M, Mahavadi P, Maiani E, Maiese K, Maiti P, Maiuri MC, Majello B, Major MB, Makareeva E, Malik F, Mallilankaraman K, Malorni W, Maloyan A, Mammadova N, Man GCW, Manai F, Mancias JD, Mandelkow EM, Mandell MA, Manfredi AA, Manjili MH, Manjithaya R, Manque P, Manshian BB, Manzano R, Manzoni C, Mao K, Marchese C, Marchetti S, Marconi AM, Marcucci F, Mardente S, Mareninova OA, Margeta M, Mari M, Marinelli S, Marinelli O, Mariño G, Mariotto S, Marshall RS, Marten MR, Martens S, Martin APJ, Martin KR, Martin S, Martin S, Martín-Segura A, Martín-Acebes MA, Martin-Burriel I, Martin-Rincon M, Martin-Sanz P, Martina JA, Martinet W, Martinez A, Martinez A, Martinez J, Martinez-Velazquez M, Martinez-Lopez N, Martinez-Vicente M, Martins DO, Martins JO, Martins WK, Martins-Marques T, Marzetti E, Masaldan S, Masclaux-Daubresse C, Mashek DG, Massa V, Massieu L, Masson GR, Masuelli L, Masyuk AI, Masyuk TV, Matarrese P, Matheu A, Matoba S, Matsuzaki S, Mattar P, Matte A, Mattoscio D, Mauriz JL, Mauthe M, Mauvezin C, Maverakis E, Maycotte P, Mayer J, Mazzocchi G, Mazzoni C, Mazzulli JR, McCarty N, McDonald C, McGill MR, McKenna SL, McLaughlin BA, McLoughlin F, McNiven MA, McWilliams TG, Mechta-Grigoriou F, Medeiros TC, Medina DL, Megeney LA, Megyeri K, Mehrpour M, Mehta JL, Meijer AJ, Meijer AH, Mejlvang J, Meléndez A, Melk A, Memisoglu G, Mendes AF, Meng D, Meng F, Meng T, Menna-Barreto R, Menon MB, Mercer C, Mercier AE, Mergny JL, Merighi A, Merkley SD, Merla G, Meske V, Mestre AC, Metur SP, Meyer C, Meyer H, Mi W, Miolet-Perez J, Miao J, Micalé L, Miki Y, Milan E, Milczarek M, Miller DL, Miller SI, Miller S, Millward SW, Milosevic I, Minina EA, Mirzaei H, Mirzaei HR, Mirzaei M, Mishra A, Mishra N, Mishra PK, Marjanovic MM, Misasi R, Misra A, Misso G, Mitchell C, Mitou G, Miura T, Miyamoto S, Miyazaki M, Miyazaki M, Miyazaki T, Miyazawa K, Mizushima N, Mogensen TH, Mograbi B, Mohammadinejad R, Mohamud Y, Mohanty A, Mohapatra S, Möhlmann T, Mohammed A, Moles A, Moley KH, Molinari M, Mollace V, Møller AB, Mollereau B, Mollinedo F, Montagna C, Monteiro MJ, Montella A, Montes LR, Montico B, Mony VK, Compagnoni GM, Moore MN, Moosavi MA, Mora AL, Mora M, Morales-Alamo D, Moratalla R, Moreira PI, Morelli E, Moreno S, Moreno-Blas D, Moresi V, Morga B, Morgan AH, Morin F, Morishita H, Moritz OL, Moriyama M, Moriyasu Y, Morleo M, Morselli E, Moruno-Manchon JF, Moscat J, Mostowy S, Motori E, Moura AF, Moustaid-Moussa N, Mrakovcic M, Muciño-Hernández G, Mukherjee A, Mukhopadhyay S, Levy JMM, Mulero V, Muller S, Münch C, Munjal A, Munoz-Canoves P, Muñoz-Galdeano T, Münz C, Murakawa T, Muratori C, Murphy BM, Murphy JP, Murthy A, Myöhänen TT, Mysorekar IU, Mytych J, Nabavi SM, Nabissi M, Nagy P, Nah J, Nahimana A, Nakagawa I, Nakamura K, Nakatogawa H, Nandi SS, Nanjundan M, Nanni M, Napolitano G, Nardacci R, Narita M, Nassif, Nathan I, Natsumeda M, Naude RJ, Naumann C, Naveiras O, Navid F, Nawrocki ST, Nazarko TY, Nazio F, Negoita F, Neill T, Neisch AL, Neri LM, Netea MG, Neubert P, Neufeld TP,

Neumann D, Neutzner A, Newton PT, Ney PA, Nezis IP, Ng CCW, Ng TB, Nguyen HTT, Nguyen LT, Ni HM, Cheallaigh CN, Ni Z, Nicolao MC, Nicoli F, Nieto-Diaz M, Nilsson P, Ning S, Niranjan R, Nishimune H, Niso-Santano M, Nixon RA, Nobili A, Nobrega C, Noda T, Nogueira-Recalde U, Nolan TM, Nombela I, Novak I, Novoa B, Nozawa T, Nukina N, Nussbaum-Krammer C, Nylandsted J, O'Donovan TR, O'Leary SM, O'Rourke EJ, O'Sullivan MP, O'Sullivan TE, Oddo S, Oehme I, Ogawa M, Ogier-Denis E, Ogmundsdottir MH, Ogretmen B, Oh GT, Oh SH, Oh YJ, Ohama T, Ohashi Y, Ohmuraya M, Oikonomou V, Ojha R, Okamoto K, Okazawa H, Oku M, Oliván S, Oliveira JMA, Ollmann M, Olzmann JA, Omari S, Omary MB, Önal G, Ondrej M, Ong SB, Ong SG, Onnis A, Orellana JA, Orellana-Muñoz S, Ortega-Villaizán MM, Ortiz-Gonzalez XR, Ortona E, Osiewacz HD, Osman AHK, Osta R, Otegui MS, Otsu K, Ott C, Ottobriini L, Ou JJ, Outeiro TF, Oynebraten I, Ozturk M, Pagès G, Pahari S, Pajares M, Pajvani UB, Pal R, Paladino S, Pallet N, Palmieri M, Palmisano G, Palumbo C, Pampaloni F, Pan L, Pan Q, Pan W, Pan X, Panasyuk G, Pandey R, Pandey UB, Pandya V, Paneni F, Pang SY, Panzarini E, Papademetrio DL, Papaleo E, Papinski D, Papp D, Park EC, Park HT, Park JM, Park JI, Park JT, Park J, Park SC, Park SY, Parola AH, Parys JB, Pasquier A, Pasquier B, Passos JF, Pastore N, Patel HH, Patschan D, Pattingre S, Pedraza-Alva G, Pedraza-Chaverri J, Pedrozo Z, Pei G, Pei J, Peled-Zehavi H, Pellegrini JM, Pelletier J, Peñalva MA, Peng D, Peng Y, Penna F, Pennuto M, Pentimalli F, Pereira CMF, Pereira GJS, Pereira LC, Almeida LP, Perera ND, Pérez-Lara A, Perez-Oliva AB, Pérez-Pérez ME, Periyasamy P, Perl A, Perrotta C, Perrotta I, Pestell RG, Petersen M, Petrache I, Petrovski G, Pfirrmann T, Pfister AS, Philips JA, Pi H, Picca A, Pickrell AM, Picot S, Pierantoni GM, Pierdominici M, Pierre P, Pierrefite-Carle V, Pierzynowska K, Pietrocola F, Pietruczuk M, Pignata C, Pimentel-Muñoz FX, Pinar M, Pinheiro RO, Pinkas-Kramarski R, Pinton P, Piracs K, Piya S, Pizzo P, Plantinga TS, Platta HW, Plaza-Zabala A, Plomann M, Plotnikov EY, Plun-Favreau H, Pluta R, Pocock R, Pöggeler S, Pohl C, Poirot M, Poletti A, Ponpuak M, Popelka H, Popova B, Porta H, Alcon SP, Portilla-Fernandez E, Post M, Potts MB, Poulton J, Powers T, Prahlad V, Prajsnar TK, Praticò D, Prencipe R, Priault M, Proikas-Cezanne T, Promponas VJ, Proud CG, Puertollano R, Puglielli L, Pulinilkunnit T, Puri D, Puri R, Puyal J, Qi X, Qi Y, Qian W, Qiang LI, Qiu I, Quadrilatero J, Quarleri J, Raben N, Rabinowich H, Ragona D, Ragusa MJ, Rahimi N, Rahmati M, Raia V, Raimundo N, Rajasekaran NS, Rao SR, Rami A, Ramírez-Pardo I, Ramsden DB, Randow F, Rangarajan PN, Ranieri D, Rao H, Rao L, Rao R, Rathore S, Ratnayaka JA, Ratovitski EA, Ravanan P, Ravegnini G, Ray SK, Razani B, Rebecca V, Reggiori F, Régnier-Vigouroux A, Reichert AS, Reigada D, Reiling JH, Rein T, Reipert S, Rekha RS, Ren H, Ren J, Ren W, Renault T, Renga G, Reue K, Rewitz K, Ramos BRA, Riazuddin SA, Ribeiro-Rodrigues TM, Ricci JE, Ricci R, Riccio V, Richardson DR, Rikihisa Y, Risbud MV, Risueño RM, Ritis K, Rizza S, Rizzuto R, Roberts HC, Roberts LD, Robinson KJ, Roccheri MC, Rocchi S, Rodney GG, Rodrigues T, Silva VRR, Rodriguez A, Rodriguez-Barrueco R, Rodriguez-Henche N, Rodriguez-Rocha H, Roelofs J, Rogers RS, Rogov VV, Rojo AI, Rolka K, Romanello V, Romani L, Romano A, Romano PS, Romeo-Guitart D, Romero LC, Romero M, Roney JC, Rongo C, Roperto S, Rosenfeldt MT, Rosenstiel P, Rosenwald AG, Roth KA, Roth L, Roth S, Kasper M.A. Rouschop, Roussel BD, Roux S, Rovere-Querini P, Roy A, Rozieres A, Ruano D, Rubinsztein DC, Rubtsova MP, Ruckdeschel, Ruckenstein C, Rudolf E, Rudolf R, Ruggieri A, Ruparelia AA, Rusmini P, Russell RR, Russo GL, Russo M, Russo R, Ryabaya OO, Ryan KM, Ryu KY, Sabater-Arcis M, Sachdev U, Sacher M, Sachse C, Sadhu A, Sadoshima J, Safren N, Saftig P, Sagana AP, Sahay G, Sahebkar A, Sahin M, Sahin O, Sahni S, Saito N, Saito S, Saito T, Sakai R, Sakai Y, Sakamaki JI, Saksela K, Salazar G, Salazar-Degracia A, Salekdeh GH, Saluja AK, Marques BS, Sanchez MC, Sanchez-Alcazar JA, Sanchez-Vera V, Sancho-Shimizu V, Sanderson JT, Sandri M, Santaguida S, Santambrogio L, Santana MM, Santoni G, Sanz A, Sanz P, Saran S, Sardiello M, Sargeant TJ, Sarin A, Sarkar C, Sarkar S, Sarrias MR, Sarkar S, Sarmah DT, Sarparanta J, Sathyanarayan A, Sathyanarayanan R, Scaglione KM, Scatozza F, Schaefer L, Schafer ZT, Schaible UE, Schapira AHV, Scharl M, Schatzl HM, Schein CH, Scheper W, Scheuring D, Schiaffino MV, Schiappacassi M, Schindl R, Schlattner U, Schmidt O, Schmitt R, Schmidt SD, Schmitz I, Schmukler E, Schneider A, Schneider BE, Schober R, Schoijet AC, Schott MB, Schramm M, Schröder B, Schuh K, Schüller C, Schulze RJ, Schürmanns L, Schwamborn JC, Schwarten M, Scialo F, Sciarretta S, Scott MJ, Scotto KW, Scovassi AI, Scrima A, Scrivo A, Sebastian D, Sebti S, Sedej S, Segatori L, Segev

N, Seglen PO, Seiliez I, Seki E, Selleck SB, Sellke FW, Selsby JT, Sendtner M, Senturk S, Seranova E, Sergi C, Serra-Moreno R, Sesaki H, Settembre C, Setty SRG, Sgarbi G, Sha O, Shacka JJ, Shah JA, Shang D, Shao C, Shao F, Sharbati S, Sharkey LM, Sharma D, Sharma G, Sharma K, Sharma P, Sharma S, Shen HM, Shen H, Shen J, Shen M, Shen W, Shen Z, Sheng R, Sheng Z, Sheng ZH, Shi J, Shi X, Shi YH, Shiba-Fukushima K, Shieh JJ, Shimada Y, Shimizu S, Shimozaawa M, Shintani T, Shoemaker CJ, Shojaei S, Shoji I, Shravage BV, Shridhar V, Shu CW, Shu HB, Shui K, Shukla AK, Shutt TE, Sica V, Siddiqui A, Sierra A, Sierra-Torre V, Signorelli S, Sil P, Silva BJA, Silva JD, Silva-Pavez E, Silvente-Poirot S, Simmonds RE, Simon AK, Simon HU, Simons M, Singh A, Singh LP, Singh R, Singh SV, Singh SK, Singh SB, Singh S, Singh SP, Sinha D, Sinha RA, Sinha S, Sirko A, Sirohi K, Sivridis EL, Skendros P, Skirycz A, Slaninová I, Smaili SS, Smertenko A, Smith MD, Soenen SJ, Sohn EJ, Sok SPM, Solaini G, Soldati T, Soleimanpour SA, Soler RM, Solovchenko A, Somarelli JA, Sonawane A, Song F, Song HK, Song JX, Song K, Song Z, Soria LR, Sorice M, Soukas AA, Soukup SF, Sousa D, Sousa N, Spagnuolo PA, Spector SA, Bharath MMS, St. Clair D, Stagni V, Staiano L, Stalneck CA, Stankov MV, Stathopoulos PB, Stefan KS, Stefan SM, Stefanis L, Steffan JS, Steinkasserer A, Stenmark H, Sterneckert J, Stevens C, Stoka V, Storch S, Stork B, Strappazon F, Strohecker AM, Stupack DG, Su H, Su LY, Su L, Suarez-Fontes AM, Carlos S. Subauste, Subbian S, Subirada PV, Sudhandiran G, Sue CM, Sui X, Summers C, Sun G, Sun J, Sun K, Sun MX, Sun Q, Sun Y, Sun Z, Sunahara KKS, Sundberg E, Susztak K, Sutovsky P, Suzuki H, Sweeney G, Symons JD, Sze SCW, Szewczyk NJ, Tabęcka-Lonczynska A, Tabolacci C, Tacke F, Taegtmeier H, Tafani M, Tagaya M, Tai H, Tait SWG, Takahashi Y, Takats S, Talwar P, Tam C, Tam SY, Tampellini D, Tamura A, Tan CT, Tan EK, Tan YQ, Tanaka M, Tanaka M, Tang D, Tang J, Tang TS, Tanida I, Tao Z, Taouis M, Tatenhorst L, Tavernarakis N, Taylor A, Taylor GA, Taylor JM, Tchetina E, Tee AR, Tegeder I, Teis D, Teixeira N, Teixeira-Clerc F, Tekirdag KA, Tencomnao TE, Tenreiro S, Tepikin AV, Testillano PS, Tettamanti G, Tharaux PL, Thedieck K, Thekkinghat AA, Thellung S, Thinwa JW, Thirumalaikumar VP, Thomas SM, Thomes PG, Thorburn A, Thukral L, Thum T, Thumm M, Tian L, Tichy A, Till A, Timmerman V, Titorenko VI, Todi SV, Todorova K, Toivonen JM, Tomaipitina L, Tomar D, TomasZapico C, Tomić S, Tong BCK, Tong C, Tong X, Tooze SA, Torgersen ML, Torii S, Torres-López L, Torriglia A, Towers CG, Towns R, Toyokuni S, Trajkovic V, Tramontano D, Tran QG, Travassos LH, Trelford CB, Tremel S, Trougakos IP, Tsao BP, Tschan MP, Tse HF, Tse TF, Tsugawa H, Tsvetkov AS, Tumbarello DA, Tumtas Y, Tuñón MJ, Turcotte S, Turk B, Turk V, Turner BJ, Tuxworth RI, Tyler JK, Tyutereva EV, Uchiyama Y, Ugun-Klusek A, Uhlig HH, Ułamek-Kozioł M, Ulasov IV, Umekawa M, Ungermann C, Unno R, Urbe S, Uribe-Carretero E, Üstün S, Uversky VN, Vaccari T, Vaccaro MI, Vahsen BF, Vakifahmetoglu-Norberg H, Valdor R, Valente MJ, Valko A, Vallee RB, Valverde AM, Van den Berghe G, van der Veen S, Van Kaer L, van Loosdregt J, van Wijk SJJ, Vandenbergh W, Vanhorebeek I, Vannier-Santos MA, Vannini N, Vanrell MC, Vantaggiato C, Varano G, Varela-Nieto I, Varga M, Vasconcelos MH, Vats S, Vavvas DG, Vega-Naredo I, Vega-Rubin-de-Celis S, Velasco G, Velázquez AP, Vellai T, Vellenga E, Velotti F, Verdier M, Verginis P, Vergne I, Verkade P, Verma M, Verstrecken P, Vervliet T, Vervoorts J, Vessoni AT, Victor VM, Vidal M, Vidoni C, Vieira OV, Vierstra RD, Viganó S, Vihinen H, Vijayan V, Vila M, Vilar M, Villalba JM, Villalobo A, Villarejo-Zori B, Villarroja F, Villarroja J, Vincent O, Vindis C, Viret C, Viscomi MT, Visnjic D, Vitale I, Vocablo DJ, Voitsekhovskaja OV, Volonté C, Volta M, Vomero M, Von Haefen C, Vooijs MA, Voos W, Vucicevic L, Wade-Martins R, Waguri S, Waite KA, Wakatsuki S, Walker DW, Walker MJ, Walker SA, Walter J, Wandosell FG, Wang B, Wang CY, Wang C, Wang C, Wang C, Wang CY, Wang D, Wang F, Wang F, Wang F, Wang G, Wang H, Wang H, Wang H, Wang HG, Wang J, Wang J, Wang J, Wang J, Wang K, Wang L, Wang L, Wang MH, Wang M, Wang N, Wang P, Wang P, Wang P, Wang P, Wang P, Wang QJ, Wang Q, Wang QK, Wang QA, Wang WT, Wang W, Wang X, Wang X, Wang Y, Wang Y, Wang Y, Wang YY, Wang Y, Wang Y, Wang Y, Wang Y, Wang Z, Wang Z, Wang Z, Wang Z, Warnes G, Warnsmann V, Watada H, Watanabe E, Watchon M, Wawrzyńska A, Weaver TE, Wegrzyn G, Wehman AM, Wei H, Wei L, Wei T, Wei Y, Weiergräber OH, Weihl CC, Weindl G, Weiskirchen R, Wells A, Wen RH, Wen X, Werner A, Weykopf B, Wheatley SP, Whitton JL, Whitworth AJ, Wiktorska K, Wildenberg ME, Wileman T, Wilkinson S, Willbold D, Williams B, Williams RSB, Williams RL, Williamson PR, Wilson RA, Winner B, Winsor NJ, Witkin SS, Wodrich

- H, Woehlbier U, Wollert T, Wong E, Wong JH, Wong RW, Wong VKW, Wong WWL, Wu AG, Wu C, Wu J, Wu J, Wu KK, Wu M, Wu SY, Wu S, Wu SY, Wu S, Wu WKK, Wu X, Wu X, Wu YW, Wu Y, Xavier RJ, Xia H, Xia L, Xia Z, Xiang G, Xiang J, Xiang M, Xiang W, Xiao B, Xiao G, Xiao H, Xiao HT, Xiao J, Xiao L, Xiao S, Xiao Y, Xie B, Xie CM, Xie M, Xie Y, Xie Z, Xie Z, Xilouri M, Xu C, Xu E, Xu H, Xu J, Xu JR, Xu L, Xu WW, Xu X, Xue Y, Yakhine-Diop SMS, Yamaguchi M, Yamaguchi O, Yamamoto A, Yamashina S, Yan S, Yan SJ, Yan Z, Yanagi Y, Chuanbin Yang, Dun-Sheng Yang, Huan Yang, Huang-Tian Yang, Hui Yang, Yang JM, Yang J, Yang J, Yang L, Yang L, Yang M, Yang PM, Yang Q, Yang S, Yang S, Yang SF, Yang W, Yang WY, Yang X, Yang X, Yang Y, Yang Y, Yao H, Yao S, Yao X, Yao YG, Yao YM, Yasui T, Yazdankhah M, Yen PM, Yi C, Yin XM, Yin Y, Yin Z, Yin Z, Ying M, Ying Z, Yip CK, Yiu SPT, Yoo YH, Yoshida K, Yoshii SR, Yoshimori T, Yousefi B, Yu B, Yu H, Yu J, Yu J, Yu L, Yu ML, Yu SW, Yu VC, Yu WH, Yu Z, Yu Z, Yuan J, Yuan LQ, Yuan S, Yuan SSF, Yuan Y, Yuan Z, Yue J, Yue Z, Yun J, Yung RL, Zacks DN, Zaffagnini G, Zambelli VO, Zanella I, Zang QS, Zanivan S, Zappavigna S, Zaragoza P, Zarbalis KS, Zarebkohan A, Zarrouk A, Zeitlin SO, Zeng J, Zeng JD, Žerovnik E, Zhan L, Zhang B, Zhang DD, Zhang H, Zhang H, Zhang H, Zhang H, Zhang H, Zhang H, Zhang H, Zhang HL, Zhang J, Zhang J, Zhang JP, Zhang KYB, Zhang LW, Zhang L, Zhang L, Zhang L, Zhang L, Zhang M, Zhang P, Zhang S, Zhang W, Zhang X, Zhang XW, Zhang X, Zhang X, Zhang X, Zhang X, Zhang XD, Zhang Y, Zhang Y, Zhang Y, Zhang YD, Zhang Y, Zhang YY, Zhang Y, Zhang Z, Zhang Z, Zhang Z, Zhang Z, Zhang Z, Zhao H, Zhao L, Zhao S, Zhao T, Zhao XF, Zhao Y, Zhao Y, Zhao Y, Zhao Y, Zheng G, Zheng K, Zheng L, Zheng S, Zheng XL, Zheng Y, Zheng ZG, Zhivotovsky B, Zhong Q, Zhou A, Zhou B, Zhou C, Zhou G, Zhou H, Zhou H, Zhou H, Zhou J, Zhou J, Zhou J, Zhou J, Zhou K, Zhou R, Zhou XJ, Zhou Y, Zhou Y, Zhou Y, Zhou ZY, Zhou Z, Zhu B, Zhu C, Zhu GQ, Zhu H, Zhu H, Zhu H, Zhu WG, Zhu Y, Zhu Y, Zhuang H, Zhuang X, Zientara-Rytter K, Zimmermann CM, Ziviani E, Zoladek T, Zong WX, Zorov DB, Zorzano A, Zou W, Zou Z, Zou ZZ, Zuryn S, Zwierschke W, Brand-Saberi B, Dong XC, Kenchappa CS, Li Z, Lin Y, Oshima S, Rong Y, Sluimer JC, Stallings CL, Tong CK. 2021. Guidelines for the use and interpretation of assays for monitoring autophagy. **Autophagy** 17: 1 - 382. <https://doi.org/10.1080/15548627.2020.1797280>
- Kokila T, Ramesh PS, Geetha D. 2016. Biosynthesis of AgNPs using *Carica papaya* peel extract and evaluation of its antioxidant and antimicrobial activities. **Ecotoxicol Environ Saf** 134: 1 - 7. <https://doi.org/10.1016/j.ecoenv.2016.03.021>
- Konan NA, Bacchi EM. 2007. Antiulcerogenic effect and acute toxicity of a hydroethanolic extract from the cashew (*Anacardium occidentale* L.) leaves. **Ecotoxicol Environ Saf** 112: 237 - 242. <https://doi.org/10.1016/j.jep.2007.03.003>
- Koochak H, Seyyednejad SM, Motamedi H. 2010. Preliminary study on the antibacterial activity of some medicinal plants of Khuzestan (Iran). **Asian Pac J Trop Med** 3: 180 - 184. [https://doi.org/10.1016/S1995-7645\(10\)60004-1](https://doi.org/10.1016/S1995-7645(10)60004-1)
- Kragh KM, Nielsen JE, Nielsen KK, Dreboldt S, Mikkelsen JD. 1995. Characterization and localization of new antifungal cysteine-rich proteins from *Beta vulgaris*. **Mol Plant Microbe Interact** 8: 424 - 434. <https://doi.org/10.1094/MPMI-8-0424>
- Kumar R, Murugananthan G, Nandakumar K, Talwar S. 2011. Isolation of anxiolytic principle from ethanolic root extract of *Cardiospermum halicacabum*. **Phytomedicine** 18: 219 - 223. <https://doi.org/10.1016/j.phymed.2010.07.002>
- Lange W, Brandenburg WA, De Bock TSM. 1999. Taxonomy and cultonony of beet (*Beta vulgaris* L.). **Bot J Linn Soc** 130: 81 - 96. <https://doi.org/10.1006/bojl.1998.0250>
- Lee E, Eom JE, Kim HL, Kang DH, Jun KY, Jung DS, Kwon Y. 2012. Neuroprotective effect of undecylenic acid extracted from *Ricinus communis* L. through inhibition of μ -calpain. **Eur J Pharm Sc** 46: 17 - 25. <https://doi.org/10.1016/j.ejps.2012.01.015>
- Leite FRM, Ramalho LTDO. 2008. Bone regeneration after demineralized bone matrix and castor oil (*Ricinus communis*) polyurethane implantation. **J Appl Oral Sci** 16: 122 - 126. <https://doi.org/10.1590/S1678-77572008000200008>
- Leite NS, Lima AP, Araújo-Neto V, Estevam CS, Pantaleão SM, Camargo EA, Fernandes RPM, Costa SKP,

- Muscará MN, Thomazzi SM. 2015. Avaliação das atividades cicatrizante, anti-inflamatória tópica e antioxidante do extrato etanólico da *Sideroxylon obtusifolium* (quixabeira). **Rev Bras Plant Med** 17: 164 - 170. https://doi.org/10.1590/1983-084X/09_189
- Levecke B, Buttle DJ, Behnke JM, Duce IR, Vercruysse J. 2014. Cysteine proteinases from papaya (*Carica papaya*) in the treatment of experimental *Trichuris suis* infection in pigs: Two randomized controlled trials. **Parasit Vectors** 7: 1 - 8. <https://doi.org/10.1186/1756-3305-7-255>
- Li ZY, Wang Y, Shen WT, Zhou P. 2012. Content determination of benzyl glucosinolate and anti-cancer activity of its hydrolysis product in *Carica papaya* L. **Asian Pac J Trop Med** 5: 231 - 233. [https://doi.org/10.1016/S1995-7645\(12\)60030-3](https://doi.org/10.1016/S1995-7645(12)60030-3)
- Lima CMBL, Freitas FIS, Moraes LCSL, Cavalcanti MGS, Silva LF, Padilha RJR, Barbosa CGS, Santos FAB, Alves LC, Diniz MFFM. 2011. Ultrastructural study on the morphological changes to male worms of *Schistosoma mansoni* after *in vitro* exposure to allicin. **Rev Soc Bras Med Trop** 44: 327 - 330. <https://doi.org/10.1590/S0037-86822011005000023>
- Lima KVS. 2014. **Conhecimento Etnobiológico Medicinal para saúde da mulher indígena Kantaruré, Bahia, Brasil**. Tesis, Universidade do Estado da Bahia, Brazil.
- Lima KVS, Araujo JF, Lopes RC. 2016. **Práticas agroecológicas de populações tradicionais como estratégia para mitigação da crise socioambiental**. In Marques J, Fraga J. Ecologia Humana e Agroecologia. SABEH, Brasil. http://www.sabeh.org.br/?mbddb_book=ecologia-humana-e-agroecologia
- Lin JY, Liu SY. 1986. Studies on the antitumor lectins isolated from the seeds of *Ricinus communis* (castor bean). **Toxicon** 24: 757 - 765.
- Liu T, Zhang J, Li K, Deng L, Wang H. 2020. Combination of an autophagy inducer and an autophagy inhibitor: A smarter strategy emerging in cancer therapy. **Front Pharmacol** 11: 1 - 14. <https://doi.org/10.3389/fphar.2020.00408>
- Lohiya NK, Mishra PK, Pathak N, Manivannan B, Bhande SS, Panneerdoss S, Sriram S. 2005. Efficacy trial on the purified compounds of the seeds of *Carica papaya* for male contraception in albino rat. **Reprod Toxicol** 20: 135 - 148. <https://doi.org/10.1016/j.reprotox.2004.11.015>
- Luciano PA. 2015. When quinine was King: A Note on the global ecology of health. **Pract Anthropol** 37: 31 - 34. <https://doi.org/10.17730/praa.37.2.r745608t35g36606>
- Luoga W, Mansur F, Buttle DJ, Duce IR, Garnett MC, Lowe A, Behnke JM. 2015. The relative anthelmintic efficacy of plant-derived cysteine proteinases on intestinal nematodes. **J Helminthol** 89: 165 - 174. <https://doi.org/10.1017/S0022149X13000692>
- Macedo ITF, Bevilacqua CML, Oliveira LMB, Camurça-Vasconcelos ALF, Vieira LS, Oliveira FR, Queiroz-Junior EM, Portela BG, Barros RS, Chagas ACS. 2009. Atividade ovicida e larvicida *in vitro* do óleo essencial de *Eucalyptus globulus* sobre *Haemonchus contortus*. **Rev Bras Parasitol Vet** 18: 62 - 66. <https://doi.org/10.4322/rbpv.01803011>
- Machado AC, Dezan E, Gomes-Filho JE, Cintra LTA, Ruvíere DB, Zoccal R, Damante CA, Gaetti Jardim E. 2012. Evaluation of tissue reaction to Aroeira (*Myracrodruon urundeuva*) extracts: A histologic and edemogenic study. **J Appl Oral Sci** 20: 414 - 418. <https://doi.org/10.1590/S1678-77572012000400005>
- Magalhães LG, Kapadia GJ, Silva Tonuci LR, Caixeta SC, Parreira NA, Rodrigues V, Silva Filho AA. 2010. *In vitro* schistosomicidal effects of some phloroglucinol derivatives from *Dryopteris* species against *Schistosoma mansoni* adult worms. **Parasitol Res** 106: 395 - 401. <https://doi.org/10.1007/s00436-009-1674-8>
- Mahata D, Mandal SM, Bharti R, Gupta VK, Mandal M, Nag A, Nando GB. 2014. Self-assembled cardanol azo derivatives as antifungal agent with chitin-binding ability. **Int J Biol Macromol** 69: 5 - 11. <https://doi.org/10.1016/j.ijbiomac.2014.05.017>
- Mahmood A, Omar MN, Ngah N. 2012. Galactagogue effects of *Musa x paradisiaca* flower extract on lactating rats. **Asian Pac J Trop Med** 5: 882 - 886. [https://doi.org/10.1016/S1995-7645\(12\)60164-3](https://doi.org/10.1016/S1995-7645(12)60164-3)
- Mannoji H, Yeger H, Becker LE. 1986. A specific histochemical marker (lectin *Ricinus communis* agglutinin-1) for normal human microglia, and application to routine histopathology. **Acta Neuropathol** 71: 341 - 343. <https://doi.org/10.1007/BF00688060>

- Marie-Magdeleine C, Boval M, Philibert L, Borde A, Archimède H. 2010. Effect of banana foliage (*Musa x paradisiaca*) on nutrition, parasite infection and growth of lambs. **Livest Sci** 131: 234 - 239. <https://doi.org/10.1016/j.livsci.2010.04.006>
- Marie-Magdeleine C, Udino L, Philibert L, Bocage B, Archimede H. 2014. In vitro effects of *Musa x paradisiaca* extracts on four developmental stages of *Haemonchus contortus*. **Res Vet Sci** 96: 127 - 132. <https://doi.org/10.1016/j.rvsc.2013.12.004>
- Mattalia G, Sôukand R, Corvo P, Pieroni A. 2021. “We became rich and we lost everything”: Ethnobotany of remote mountain villages of Abruzzo and Molise, central Italy. **Hum Ecol** 49: 217 - 224. <https://doi.org/10.1007/s10745-020-00209-6>
- Mbatchou VC, Kosoono I. 2012. Aphrodisiac activity of oils from *Anacardium occidentale* L. seeds and seed shells. **Phytopharmacology** 2: 81 - 91.
- Medeiros KCP, Monteiro JC, Diniz MFFM, Medeiros IA, Silva BA, Piuvezam MR. 2007. Effect of the activity of the Brazilian polyherbal formulation: *Eucalyptus globulus* Labill, *Peltodon radicans* Pohl and *Schinus terebinthifolius* Radd in inflammatory models. **Rev Bras Farmacogn** 17: 23 - 28. <https://doi.org/10.1590/S0102-695X2007000100006>
- Mello MJR, Leite JAD, Vasconcellos RJH, Morais HHA. 2013. Atividade anti-inflamatória, cicatrizante e antimicrobiana do extrato aquoso de aroeira-do-sertão a 20% (*Myracrodruon urundeuva* fr. All.), aplicado em fraturas expostas induzidas em mandíbula de coelho. **Rev Cir Traumatol Buco-Maxilo-Facial** 13: 97 - 104.
- Melo JIM, Alves IM, Sousa RTM, Barbosa LMMA, Andrade WM. 2010a. Verbenaceae sensu lato em um trecho da ESEC Raso da Catarina, Bahia, Brasil. **Revista Caatinga** 23: 41 - 47.
- Melo JG, Araújo TAS, Castro VTNA, Cabral DLV, Rodrigues MD, Nascimento SC, Amorim ELC, Albuquerque UP. 2010b. Antiproliferative activity, antioxidant capacity and tannin content in plants of semi-arid northeastern Brazil. **Molecules** 15: 8534 - 8542. <https://doi.org/10.3390/molecules15128534>
- Mengarda AC, Mendonça PS, Morais CS, Cogo RM, Mazloun SF, Salvadori MC, Teixeira FS, Morais TR, Antar GM, Lago JHG, Moraes J. 2020. Antiparasitic activity of piplartine (piperlongumine) in a mouse model of schistosomiasis. **Acta Trop** 205: 1 - 17. <https://doi.org/10.1016/j.actatropica.2020.105350>
- Mesoudi A. 2007. A darwinian theory of cultural evolution can promote an evolutionary synthesis for the social sciences. **Biol Theory** 2: 263 - 275. <https://doi.org/10.1162/biot.2007.2.3.263>
- Million M, Gautret P, Colson P, Roussel Y, Dubourg G, Chabriere E, Honore S, Rolain JM, Fenollar F, Fournier PE, Lagier JC, Parola P, Brouqui P, Raoult D. 2020. Clinical efficacy of chloroquine derivatives in COVID-19 infection: comparative meta-analysis between the big data and the real world. **New Microbes New Infect** 38: 100709. <https://doi.org/10.1016/j.nmni.2020.100709>
- Ministry of Health. 2009. **MS elabora relação de plantas medicinais de interesse ao SUS**. https://bvsms.saude.gov.br/bvs/sus/pdf/marco/ms_relacao_plantas_medicinais_sus_0603.pdf
- Mojica L, Mejía EG. 2016. Optimization of enzymatic production of anti-diabetic peptides from black bean (*Phaseolus vulgaris* L.) proteins, their characterization and biological potential. **Food Funct** 7: 713 - 727. <https://doi.org/10.1039/c5fo01204j>
- Monk JM, Zhang CP, Wu W, Zarepoor L, Lu JT, Liu R, Pauls KP, Wood GA, Tsao R, Robinson LE, Power KA. 2015. White and dark kidney beans reduce colonic mucosal damage and inflammation in response to dextran sodium sulfate. **J Nut Biochem** 26: 752 - 760. <https://doi.org/10.1016/j.jnutbio.2015.02.003>
- Moraes D, Levenhagen MA, Costa-Cruz JM, Costa-Netto AP, Rodrigues RM. 2017. In vitro efficacy of latex and purified papain from *Carica papaya* against *Strongyloides venezuelensis* eggs and larvae. **Rev Inst Med Trop Sao Paulo** 59: 1 - 6. <https://doi.org/10.1590/s1678-9946201759007>
- Moraes J, Nascimento C, Yamaguchi LF, Kato MJ, Nakano E. 2012. *Schistosoma mansoni*: In vitro schistosomicidal activity and tegumental alterations induced by piplartine on schistosomula. **Exp Parasitol** 132: 222 - 227. <https://doi.org/10.1016/j.exppara.2012.07.004>
- Morais SM, Dantas JDP, Silva ARA, Magalhães EF. 2005. Plantas medicinais usadas pelos índios Tapebas do Ceará. **Rev Bras Farmacogn** 15: 169 - 177. <https://doi.org/10.1590/s0102-695x2005000200017>
- Morais TC, Pinto NB, Carvalho KMMB, Rios JB, Ricardo NMPS, Trevisan MTS, Rao VS, Santos FA. 2010.

- Protective effect of anacardic acids from cashew (*Anacardium occidentale*) on ethanol-induced gastric damage in mice. **Chem Biol Interact** 183: 264 - 269. <https://doi.org/10.1016/j.cbi.2009.10.008>
- Mothé CG, Souza IA, Calazans GMT. 2008. Antitumor activity of cashew gum from *Anacardium Occidentale* L. **Agro Food Ind Hi Tech** 19: 50 - 52.
- Moura-Costa GF, Nocchi SR, Ceole LF, Mello JCPD, Nakamura CV, Dias Filho BP, Temponi LG, Ueda-Nakamura T. 2012. Antimicrobial activity of plants used as medicinals on an indigenous reserve in Rio das Cobras, Paraná, Brazil. **J Ethnopharmacol** 143: 631 - 638. <https://doi.org/10.1016/j.jep.2012.07.016>
- Muthumani P, Meera R, Venkatraman S, Ganapathy S, Devi P. 2010. Study of phyto chemical, analgesic and antiulcer activity of extracts of aerial parts of *Cardiospermum hallicacabum* Linn. **Int J Pharmaceut Sci Res** 1: 128 - 137.
- Nade VS, Kawale LA, Zambre SS, Kapure AB. 2015. Neuroprotective potential of *Beta vulgaris* L. in Parkinson's disease. **Indian J Pharmacol** 47: 403 - 408.
- Naithani HB. 2014. **Botany of genus Eucalyptus**, In Bhojvaid PP, Kaushik S, Singh YP, Kumar D, Thapliyal M, Barthwal S: Eucalyptis in India. ENVIS centre on forestry ICFRE, India.
- Napoleão TH, Belmonte BR, Pontual EV, Albuquerque LP, Sá RA, Paiva LM, Coelho LCBB, Paiva PMG. 2013. Deleterious effects of *Myracrodruon urundeuva* leaf extract and lectin on the maize weevil, *Sitophilus zeamais* (Coleoptera, Curculionidae). **J Stored Prod Res** 54: 26 - 33. <https://doi.org/10.1016/j.jspr.2013.04.002>
- Nath S, Choudhury MD, Roychoudhury S, Talukdar AD, Misro MM. 2013. Male contraceptive efficacy of *Ricinus communis* L. extract. **J Ethnopharmacol** 149: 328 - 334. <https://doi.org/10.1016/j.jep.2013.06.044>
- Naz R, Bano A. 2012. Antimicrobial potential of *Ricinus communis* leaf extracts in different solvents against pathogenic bacterial and fungal strains. **Asian Pac J Trop Biomed** 2: 944 - 947. [https://doi.org/10.1016/S2221-1691\(13\)60004-0](https://doi.org/10.1016/S2221-1691(13)60004-0)
- Neiva VA, Ribeiro MNS, Nascimento FRF, Cartágenes MSS, Coutinho-Moraes DF, Amaral FMM. 2014. Plant species used in giardiasis treatment: Ethnopharmacology and *in vitro* evaluation of anti-Giardia activity. **Rev Bras Farmacogn** 24: 215 - 224. <https://doi.org/10.1016/j.bjp.2014.04.004>
- Nepomuceno SC, Athiê-Souza SM, Buril MT. 2016. Convolvulaceae da microrregião do Alto Capibaribe, PE, Brasil. **Hoehnea** 43: 371 - 386. <https://doi.org/10.1590/2236-8906-31/2016>
- Nirmala M, Giriya K, Lakshman K, Divya T. 2012. Hepatoprotective activity of *Musa paradisiaca* on experimental animal models. **Asian Pac J Trop Biomed** 2: 11 - 15. [https://doi.org/10.1016/S2221-1691\(11\)60181-0](https://doi.org/10.1016/S2221-1691(11)60181-0)
- Nwaehujor CO, Ode JO, Ekwere MR, Udegbuma RI. 2014. Anti-fertility effects of fractions from *Carica papaya* (Pawpaw) Linn. methanol root extract in male Wistar rats. **Arab J Chem** 12: 1563 - 1568. <https://doi.org/10.1016/j.arabjc.2014.10.018>
- Oduola T, Adeniyi FAA, Ogunyemi EO, Bello IS, Idowu TO. 2006. Antisickling agent in an extract of unripe pawpaw (*Carica papaya*): Is it real? **Afr J Biotechnol** 5: 1947 - 1949. <https://doi.org/10.5897/AJB2006.000-5089>
- Okeniyi JAO, Ogunlesi TA, Oyelami OA, Adeyemi LA. 2007. Effectiveness of dried *Carica papaya* seeds against human intestinal parasitosis: A pilot study. **J Med Food** 10: 194 - 196. <https://doi.org/10.1089/jmf.2005.065>
- Olajide OA, Aderogba MA, Fiebich BL. 2013. Mechanisms of anti-inflammatory property of *Anacardium occidentale* stem bark: Inhibition of NF-κB and MAPK signalling in the microglia. **J Ethnopharmacol** 145: 42 - 49. <https://doi.org/10.1016/j.jep.2012.10.031>
- Oliveira DG, Prince KA, Higuchi CT, Santos ACB, Lopes LMX, Simões MJS, Leite CQF. 2007. Antimycobacterial activity of some Brazilian indigenous medicinal drinks. **Rev Ciênc Farm Básica Aplic** 28: 165 - 169.
- Oliveira LMB, Bevilacqua CML, Macedo ITF, Morais SM, Machado LKA, Campello CC, Mesquita MA. 2011. Effects of *Myracrodruon urundeuva* extracts on egg hatching and larval exsheathment of *Haemonchus contortus*. **Parasitol Res** 109: 893 - 898. <https://doi.org/10.1007/s00436-011-2331-6>
- Oliveira FC, Albuquerque UP, Fronseca-Kruel VS, Hanazaki N. 2009. Avanços nas pesquisas etnobotânicas no Brasil. **Acta Bot Bras** 23: 590 - 605. <https://doi.org/10.1590/S0102-33062009000200031>

- Oliveira LN, Duarte ER, Nogueira FA, Silva RB, Faria Filho DE, Geraseev LC. 2010. Eficácia de resíduos da bananicultura sobre a inibição do desenvolvimento larval em *Haemonchus spp.* provenientes de ovinos. **Cienc Rural** 40: 488 - 490. <https://doi.org/10.1590/s0103-84782009005000254>
- Oloyede HOB, Adaja MC, Ajiboye TO, Salawu MO. 2015. Human nature, the means-ends relationship, and alienation: Themes for potential EastWest collaboration. **J Integr Med** 13: 105 - 114. [https://doi.org/https://doi.org/10.1016/S2095-4964\(15\)60160-1](https://doi.org/https://doi.org/10.1016/S2095-4964(15)60160-1)
- Oloyede OI. 2005. Chemical profile of unripe pulp of *Carica papaya*. **Pak J Nut** 4: 379 - 381. <https://doi.org/10.3923/pjn.2005.379.381>
- Orafidiya LO, Agbani EO, Oyedele AO, Babalola OO, Onayemi O, Aiyedun FF. 2004. The effect of *Aloe vera* gel on the anti-acne properties of the essential oil of *Ocimum gratissimum* Linn leaf - A preliminary clinical investigation. **Int J Aromather** 14: 15 - 21. <https://doi.org/10.1016/j.ijat.2003.12.005>
- Oriakpono O, Aduabobo H, Awi-Waadu GDB, Nzeako S. 2012. Anti-parasitic effects of methanolic extracts of *Artemisia annua* L. against parasites of sarotherodon melanotheron. **Int J Modern Biol Med** 1: 108 - 116.
- Otsuki N, Dang NH, Kumagai E, Kondo A, Iwata S, Morimoto C. 2010. Aqueous extract of *Carica papaya* leaves exhibits anti-tumor activity and immunomodulatory effects. **J Ethnopharmacol** 127: 760 - 767. <https://doi.org/10.1016/j.jep.2009.11.024>
- Owoyele BV, Adebukola OM, Funmilayo AA, Soladoye AO. 2008. Anti-inflammatory activities of ethanolic extract of *Carica papaya* leaves. **Inflammopharmacology** 16: 168 - 173. <https://doi.org/10.1007/s10787-008-7008-0>
- Paganotte DM, Sannomiya M, Rinaldo D, Vilegas W, Salgado HRN. 2016. *Operculina macrocarpa*: Chemical and intestinal motility effect in mice. **Rev Bras Farmacogn** 26: 1 - 6. <https://doi.org/10.1016/j.bjp.2016.03.003>
- Paiva PMG, Napoleão TH, Santos NDL, Correia MTS, Navarro DMAF, Coelho LCB. 2011. **Plant compounds with *Aedes aegypti* larvicidal activity and other biological properties**. In: Liong MT. Bioprocess sciences and technology. Nova Science Publishers Inc, New York, USA.
- Panahi Y, Khedmat H, Valizadegan G, Mohtashami R, Sahebkar A. 2015. Efficacy and safety of *Aloe vera* syrup for the treatment of gastroesophageal reflux disease: a pilot randomized positive-controlled trial. **J Tradit Chin Med** 35: 632 - 636. [https://doi.org/10.1016/s0254-6272\(15\)30151-5](https://doi.org/10.1016/s0254-6272(15)30151-5)
- Patil CD, Borase HP, Salunkhe RB, Suryawanshi RK, Narkhade CP, Salunke BK, Patil SV. 2014. Mosquito larvicidal potential of *Gossypium hirsutum* (Bt cotton) Leaves extracts against *Aedes aegypti* and *Anopheles stephensi* larvae. **J Arthropod Borne Dis** 8: 91 - 101.
- Paull RE, Duarte O. 2011. **Tropical Fruits**. Ed. II, CAB International, Wallingford, UK.
- Peiris LDC, Dhanushka MAT, Jayathilake TAHDG. 2015. Evaluation of aqueous leaf extract of *Cardiospermum halicacabum* (L.) on fertility of male rats. **Biomed Res Int** 2015: 1 - 6. <https://doi.org/10.1155/2015/175726>
- Pereira LA, Amorim BS, Alves M, Somner GV, Baborsa MRV. 2016. Flora da usina São José, Igarassu, Pernambuco: Sapindaceae. **Rodriguésia** 67: 1047 - 1059. <https://doi.org/10.1590/2175-7860201667414>
- Pierdoná TM, Lima NR, Rodrigues RCM, Teixeira JP, Gonçalves RP, Fontenele JB, Vasconcelos SMM, Viana GSDB, Leal LKAM. 2014. The *Operculina macrocarpa* (L.) urb. (jalapa) tincture modulates human blood platelet aggregation. **J Ethnopharmacol** 151: 151 - 157. <https://doi.org/10.1016/j.jep.2013.10.008>
- Pillai NR, Vijayamma N. 1985. Some pharmacological studies on *Cardiospermum halicacabum* linn. **Anc Sci Life** 5: 32 - 36.
- Pinto CDP, Rodrigues VD, Pinto FDP, Pinto RDP, Uetanabaro APT, Pinheiro CSR, Gadea SFM, Silva TRDS, Lucchese AM. 2013. Antimicrobial activity of *Lippia* species from the Brazilian semiarid region traditionally used as antiseptic and anti-infective agents. **Evid Based Complement Alternat Med** 2013: 1 - 5. <https://doi.org/10.1155/2013/614501>
- Porto BC, Cristianini M. 2014. Evaluation of cashew tree gum (*Anacardium occidentale* L.) emulsifying properties. **LWT - Food Sci Technol** 59: 1 - 7. <https://doi.org/10.1016/j.lwt.2014.03.033>
- Queiroz LP. 2009. **Leguminosas da Caatinga**. Editora da Universidade Estadual de Feira de Santana, Feira de Santana, Brasil.

- Queiroz MMF, Queiroz EF, Zeraik ML, Marti G, Favre-Godal Q, Simões-Pires C, Marcourt L, Carrupt PA, Cuendet M, Paulo MQ, Bolzani VS, Wolfender JL. 2014. Antifungals and acetylcholinesterase inhibitors from the stem bark of *Croton heliotropiifolius*. **Phytochem Lett** 10: lxxxviii - xciii. <https://doi.org/10.1016/j.phytol.2014.08.013>
- Quinlan MB, Quinlan RJ, Nolan JM. 2002. Ethnopharmacology and herbal treatments of intestinal worms in Dominica, West Indies. **J Ethnopharmacol** 80: 75 - 83. [https://doi.org/10.1016/S0378-8741\(02\)00002-8](https://doi.org/10.1016/S0378-8741(02)00002-8)
- Quintans-Júnior LJ, Almeida RN, Falcão ACGM, Agra MF, Sousa MFV, Barbosa-Filho JM. 2002. Avaliação da atividade anticonvulsivante de plantas do Nordeste Brasileiro. **Acta Farm Bonaerense** 21: 179 - 184.
- Raj P, Dave A. 2019. Indigenous knowledge of antidiabetic medicinal plants in Madhya Pradesh. **Int J Recent Scient Res** 10: 31235 - 31237. <https://doi.org/10.24327/IJRSR>
- Ramos JMO, Santos CA, Santana DG, Santos DA, Alves PB, Thomazzi SM. 2013. Chemical constituents and potential anti-inflammatory activity of the essential oil from the leaves of *Croton argyrophyllus*. **Braz J Pharmacogn** 23: 644 - 650. <https://doi.org/10.1590/S0102-695X2013005000045>
- Rampadarath S, Puchooa D. 2016. *In vitro* antimicrobial and larvicidal properties of wild *Ricinus communis* L. in Mauritius. **Asian Pac J Trop Biomed** 6: 100 - 107. <https://doi.org/10.1016/j.apjtb.2015.10.011>
- Rampadarath S, Puchooa D, Ranghoo-Sanmukhiya VM. 2014. A comparison of polyphenolic content, antioxidant activity and insecticidal properties of *Jatropha* species and wild *Ricinus communis* L. found in Mauritius. **Asian Pac J Trop Biomed** 7: 384 - 390. [https://doi.org/10.1016/S1995-7645\(14\)60263-7](https://doi.org/10.1016/S1995-7645(14)60263-7)
- Rana D, Bhatt A, Lal B, Parkash O, Kumar A, Uniyal SK. 2021. Use of medicinal plants for treating different ailments by the indigenous people of Churah subdivision of district Chamba, Himachal Pradesh, India. **Environ Develop Sustainability** 23: 1162 - 1241. <https://doi.org/10.1007/s10668-020-00617-0>
- Rao NV, Prakash KC, Kumar SMS. 2006. Pharmacological investigation of *Cardiospermum halicacabum* (Linn) in different animal models of diarrhoea. **Indian J Pharmacol** 38: 346 - 349. <https://doi.org/10.4103/0253-7613.27703>
- Rapini A, Andrade MJG, Giulietti AM, Queiro LP, Silva JMC. 2009. **Introdução**. In Giulietti AM, Rapini A, Andrade MJG, Queiroz LP, Silva JMC. Plantas raras do Brasil. Conservação Internacional/Universidade Estadual de Feira de Santana, Belo Horizonte, Brasil.
- Rathor N, Arora T, Manocha S, Patil AN, Mediratta PK, Sharma KK. 2014. Anticonvulsant activity of *Aloe vera* leaf extract in acute and chronic models of epilepsy in mice. **J Pharm Pharmacol** 66: 477 - 485. <https://doi.org/10.1111/jphp.12181>
- Rehder VLG, Rodrigues MVN, Sartoratto A, Foglio MA. 2002. Dosagem de artemisinina em *Artemisia annua* L. por cromatografia líquida de alta eficiência com detecção por índice de refração. **Rev Bras Farmacogn** 12: 116 - 118. <https://doi.org/10.1590/s0102-695x2002000300055>
- Remali J, Aizat WM. 2021. A Review on plant bioactive compounds and their modes of action against Coronavirus infection. **Front Pharmacol** 11: 1 - 13. <https://doi.org/10.3389/fphar.2020.589044>
- Renslo AR. 2013. Antimalarial drug discovery: From quinine to the dream of eradication. **ACS Med Chem Lett** 4: 1126 - 1128. <https://doi.org/10.1021/ml4004414>
- Ribeiro ARC, Andrade FD, Medeiros MC, Camboim AS, Pereira FA, Athayde ACR, Rodrigues OG, Silva WW. 2014. Estudo da atividade anti-helmíntica do extrato etanólico de *Jatropha mollissima* (Pohl) Baill. (Euphorbiaceae) sob *Haemonchus contortus* em ovinos no semiárido paraibano. **Pesq Vet Bras** 34: 1051 - 1055. <https://doi.org/10.1590/s0100-736x2014001100002>
- Riondato I, Donno D, Roman A, Razafintsalama VE, Petit T, Mellano MG, Torti V, De Biaggi M, Rakotoniaina EN, Giacomini C, Beccaro GL. 2019. First ethnobotanical inventory and phytochemical analysis of plant species used by indigenous people living in the Maromizaha forest, Madagascar. **J Ethnopharmacol** 232: 73 - 89. <https://doi.org/10.1016/j.jep.2018.12.002>
- Ríos-de Álvarez L, Jackson F, Greer AW, Grant G, Jackson E, Morrison AA, Huntley JF. 2012. Direct anthelmintic and immunostimulatory effects of oral dosing semi-purified phytohaemagglutinin lectin in sheep infected with *Teladorsagia circumcincta* and *Trichostrongylus colubriformis*. **Vet Parasitol** 187: 267 - 274. <https://doi.org/10.1016/j.vetpar.2012.01.005>
- Rivas MR, Parets MH, Gallardo AIA, Guerra RLL, Chaviano YM. 2004. Acción antiasmática del *Aloe vera* en

- pacientes. **Rev Cub Plant Med** 9 (1).
- Rocha-Guzmán NE, Herzog A, González-Laredo RF, Ibarra-Pérez FJ, Zambrano-Galván G, Gallegos-Infante JA. 2007. Antioxidant and antimutagenic activity of phenolic compounds in three different colour groups of common bean cultivars (*Phaseolus vulgaris*). **Food Chem** 103: 521 - 527.
<https://doi.org/10.1016/j.foodchem.2006.08.021>
- Rupeshkumar M, Kavitha K, Basu S. 2012. Antioxidant and hepatoprotective effect of flavanone from *Cardiospermum halicacabum* N. against acetaminophen induced hepatotoxicity in rats. **J Pharm Res** 5: 544 - 547.
- Sacan O, Yanardag R. 2010. Antioxidant and antiacetylcholinesterase activities of chard (*Beta vulgaris* L. var. cicla). **Food Chem Toxicol** 48: 1275 - 1280. <https://doi.org/10.1016/j.fct.2010.02.022>
- Sacande M, Voutier H. 2006. *Ximenia americana* L. **Forest Landscape Denmark** 112: 1 - 2.
- Sadiq A, Hayat MQ, Ashraf M. 2014. *Artemisia annua* - **Pharmacology and biotechnology**, In Aftab T, Ferreira JFS, Khan MMA, Naeem M. *Artemisia annua* - Pharmacology and Biotechnology. Springer, Berlin, Germany. <https://doi.org/10.1007/978-3-642-41027-7>
- Saini MR, Goyal PK, Chaudhary G. 2010. Anti-tumor activity of *Aloe vera* against DMBA/croton oil-induced skin papillomagenesis in swiss albino mice. **J Environ Pathol Toxicol** 29: 127 - 135.
<https://doi.org/10.1615/JEnvironPatholToxicolOncol.v29.i2.60>
- Salehi B, Sharifi-Rad J, Quispe C, Llaque H, Villalobos M, Smeriglio A, Trombetta D, Ezzat SM, Salem MA, Zayed A, Salgado Castillo CM, Yazdi SE, Sen S, Acharya K, Sharopov F, Martins N. 2019. Insights into *Eucalyptus* genus chemical constituents, biological activities and health-promoting effects. **Trends Food Sci Technol** 91: 609 - 624. <https://doi.org/10.1016/j.tifs.2019.08.003>
- Salles HO, Braga ACL, Nascimento MTSC, Sousa AMP, Lima AR, Vieira LS, Cavalcante ACR, Egito AS, Andrade LBS. 2014. Lectin, hemolysin and protease inhibitors in seed fractions with ovicidal activity against *Haemonchus contortus*. **Rev Bras Parasitol Vet** 23: 136 - 143.
<https://doi.org/10.1590/s1984-29612014050>
- Santos FCC, Vogel FSF, Monteiro SG. 2012. Extrato aquoso de alho (*Allium sativum*) sobre nematóides gastrintestinais de ovinos. **Rev Bras Agroecol** 7: 139 - 144.
- Santos JM. 2008. **Cultura material e etnicidade dos povos indígenas do são francisco afetados por barragens: Um Estudo de Caso dos Tuxá de Rodelas, Bahia, Brasil**. Thesis, Universidade Federal da Bahia, Brazil.
- Saraswathi NT, Gnanam FD. 1997. Effect of medicinal plants on the crystallization of cholesterol. **J Cryst Growth** 179: 611 - 617. [https://doi.org/10.1016/S0022-0248\(97\)00172-3](https://doi.org/10.1016/S0022-0248(97)00172-3)
- Sátiro LN, Roque N. 2008. A família Euphorbiaceae nas caatingas arenosas do médio rio São Francisco, BA, Brasil. **Acta Bot Bras** 22: 99 - 118. <https://doi.org/10.1590/S0102-33062008000100013>
- Satrija F, Nansen P, Murtini S, He S. 1995. Anthelmintic activity of papaya latex against patent *Heligmosomoides polygyrus* infections in mice. **J Ethnopharmacol** 48: 161 - 164.
[https://doi.org/10.1016/0378-8741\(95\)01298-R](https://doi.org/10.1016/0378-8741(95)01298-R)
- Shettar AK, Kotresha K, Kaliwal BB, Vedamurthy AB. 2015. Evaluation of *in vitro* antioxidant and anti-inflammatory activities of *Ximenia americana* extracts. **Asian Pac J Trop Dis** 5: 918 - 923.
[https://doi.org/10.1016/S2222-1808\(15\)60957-4](https://doi.org/10.1016/S2222-1808(15)60957-4)
- Shokeen P, Anand P, Murali YK, Tandon V. 2008. Antidiabetic activity of 50% ethanolic extract of *Ricinus communis* and its purified fractions. **Food Chem Toxicol** 46: 3458 - 3466.
<https://doi.org/10.1016/j.fct.2008.08.020>
- Silva-Luz CL. 2011. **Anacardiaceae R.Br. na Flora Fanerogâmica do Estado de São Paulo**. Thesis, Universidade de São Paulo, Brazil. <https://doi.org/10.11606/D.41.2012.tde-27042012-123436>
- Silva AAS, Moraes SM, Falcão MJC, Vieira IGP, Ribeiro LM, Viana SM, Teixeira MJ, Barreto FS, Carvalho CA, Cardoso RPA, Andrade-Junior HF. 2014. Activity of cycloartane-type triterpenes and sterols isolated from *Musa paradisiaca* fruit peel against *Leishmania infantum chagasi*. **Phytomedicine** 21: 1419 - 1423.
<https://doi.org/10.1016/j.phymed.2014.05.005>
- Silva CG, Marinho MGV, Lucena MFA, Costa JGM. 2015a. Levantamento etnobotânico de plantas medicinais em área de Caatinga na comunidade do Sítio Nazaré, município de Milagres, Ceará, Brasil. **Rev Bras Plant**

- Med 17: 133 - 142. https://doi.org/10.1590/1983-084X/12_055
- Silva AG, Silva LCN, Filho CMB, Araújo DRC, Silva JFV, Arruda IR, Araújo JM, Correia MTS, Macedo AJ, Silva MV. 2006. Antimicrobial activity of medicinal plants of the Caatinga (semi-arid) vegetation of NE Brazil. **Phytochemistry** 1: 1 - 15.
- Silva HM. 2014. Nematodioses gastrointestinais de caprinos: uma revisão. **Rev Ciênc Agrovet** 13: 199 - 208.
- Silva FFS, Dantas BF. 2017. *Sideroxylon obtusifolium* (Humb. ex Roem. & Schult.) T.D. Penn. Quixabeira. Embrapa Semiárido – Nota Técnica/Nota Científica (ALICE), Comitê Técnico de Sementes Florestais (CTSF), Associação Brasileira de Tecnologia em Sementes (ABRATES). Londrina, Brasil.
- Silva HT, Costa AO. 2003. **Caracterização botânica de espécies silvestres do gênero Phaseolus L. (Leguminosae)**. Embrapa Arroz e Feijão, Santo Antonio de Goiás, Brasil.
- Silva JS, Sales MF, Gomes APS, Carneiro-Torres DS. 2010. Sinopse das espécies de *Croton* L. (Euphorbiaceae) no estado de Pernambuco, Brasil. **Acta Bot Bras** 24: 441 - 453. <https://doi.org/10.1590/s0102-33062010000200015>
- Silva LB, Torres EB., Silva KF, Souza JSN, Lopes MS, Andrade LH, Xavier ZF. 2012. Toxicity of ethanolic extract of *Croton heliotropiifolius* in weevil populations of stored maize grains. **J Entomol** 9: 1 - 9.
- Silva LN, Trentin DDS, Zimmer KR, Treter J, Brandelli CLC, Frasson AP, Tasca T, Silva AG, Silva MV, Macedo AJ. 2015b. Anti-infective effects of Brazilian Caatinga plants against pathogenic bacterial biofilm formation. **Pharm Biol** 53: 464 - 468. <https://doi.org/10.3109/13880209.2014.922587>
- Silva TLL, Ferreira Junior WS, Albuquerque UP. 2020. Is there a biological basis in the selection of medicinal plants in the human species? An initial approach based on chemosensory perception of taste. **Ethnobiol Conserv** 9: 1 - 15. <https://doi.org/10.15451/ec2020-01-9.03-1-15>
- Silva AV. 2003. **Etnobotânica dos índios Fulni-ô (Pernambuco, Nordeste do Brasil)**. Thesis, Universidade Federal de Pernambuco, Brazil.
- Singh A, Mishra A, Chaudhary R, Kumar V. 2020. Dear role of herbal plants in prevention and treatment of parasitic diseases. **J Scient Res** 64: 50 - 58. <https://doi.org/10.37398/jsr.2020.640106>
- Siqueira Filho JA, Santos APB, Nascimento MFS, Espírito Santo FS. 2009. **Guia de campo de árvores da caatinga**. Editora e gráfica Franciscana Ltda. Petrolina, Brasil.
- Somanadhan B, Varughese G, Palpu P, Sreedharan R, Gudiksen L, Wagner Smitt U, Nyman U. 1999. An ethnopharmacological survey for potential angiotensin converting enzyme inhibitors from Indian medicinal plants. **J Ethnopharmacol** 65: 103 - 112. [https://doi.org/10.1016/S0378-8741\(98\)00201-3](https://doi.org/10.1016/S0378-8741(98)00201-3)
- Soro TY, Traoré F, Datte JY, Nene-Bi AS. 2009. Activité antipyrétique de l'extrait aqueux de *Ximenia americana*. **Phytotherapie** 7: 297 - 303: <https://doi.org/10.1007/s10298-009-0507-3>
- Sousa FCF, Melo CTV, Citó MCO, Félix FHC, Vasconcelos SMM, Fonteles MMF, Barbosa Filho JM, Viana GSB. 2008. Plantas medicinais e seus constituintes bioativos: Uma revisão da bioatividade e potenciais benefícios nos distúrbios da ansiedade em modelos animais. **Rev Bras Farmacogn** 18: 642 - 654. <https://doi.org/10.1590/S0102-695X2008000400023>
- Sousa RG, Falcão HS, Barbosa Filho JM, Melo DMFF, Batista LM. 2013. Atividade anti-helmíntica de plantas nativas do continente americano: Uma revisão. **Rev Bras Plant Med** 15: 287 - 292. <https://doi.org/10.1590/S1516-05722013000200018>
- Souza CP, Mendes NM, Jannotti-Passos LK, Pereira JP. 1992. O uso da casca da castanha do caju, *Anacardium occidentale*, como moluscicida alternativo. **Rev Inst Med Trop Sao Paulo** 34: 459 - 466. <https://doi.org/10.1590/S0036-46651992000500014>
- Souza TM, Menezes ESB, Oliveira RV, Almeida Filho LCP, Martins JM, Moreno FB, Monteiro-Moreira ACO, Moura AAA, Carvalho AFU. 2015. Further evidences for the mode of action of the larvicidal m-pentadecadienyl-phenol isolated from *Myracrodruon urundeuva* seeds against *Aedes aegypti*. **Acta Trop** 152: 49 - 55. <https://doi.org/10.1016/j.actatropica.2015.08.010>
- Sprenger LK. 2015. Atividade ovicida e larvicida do extrato etanólico de *Aloe vera* L. sobre *Haemonchus contortus*. **Rev Bras Cienc Vet** 22: 152 - 156. <https://doi.org/10.4322/rbcv.2016.005>
- Subramaniam J, Kovendan K, Mahesh Kumar P, Murugan K, Walton W. 2012. Mosquito larvicidal activity of *Aloe vera* (Family: Liliaceae) leaf extract and *Bacillus sphaericus*, against Chikungunya vector, *Aedes aegypti*.

- Saudi J *Biol Sci* 19: 503 - 509. <https://doi.org/10.1016/j.sjbs.2012.07.003>
- Sueth-Santiago V, Moraes JBB, Alves ESS, Vannier-Santos MA, Freire-De-lima CG, Castro RN, Mendes-Silva GP, De Nigris Del Cistia C, Magalhães LG, Andricopulo AD, Santánnia CMR, Decote-Ricardo D, Edilsonfreiredelima M. 2016. The effectiveness of natural diarylheptanoids against *Trypanosoma cruzi*: Cytotoxicity, ultrastructural alterations and molecular modeling studies. **Plos One** 11: 1 - 20. <https://doi.org/10.1371/journal.pone.0162926>
- Syed TA, Ahmad SA, Holt AH, Ahmad SA, Ahmad SH, Afzal M. 1996. Management of psoriasis with *Aloe vera* extract in a hydrophilic cream: A placebo-controlled, double-blind study. **Trop Med Int Health** 1: 505 - 509. <https://doi.org/10.1046/j.1365-3156.1996.d01-91.x>
- Tabolacci C, Cordella M, Turcano L, Rossi S, Lentini A, Mariotti S, Nisini R, Sette G, Eramo A, Piredda L, De Maria R, Facchiano F, Beninati S. 2015. Aloe-emodin exerts a potent anticancer and immunomodulatory activity on BRAF-mutated human melanoma cells. **Eur J Pharmacol** 762: 283 - 292. <https://doi.org/10.1016/j.ejphar.2015.05.057>
- Taiwo K, Makinde S, Ojekale A, Sowunmi M, Oyetunji O, Lasisi M. 2018. Ethno-botanical survey , review and informatics of medicinal plants used by indigenous people of Lekki, Ibeju-Lekki Local Government Area of Lagos. **J Res Rev Sci** 5: 30 - 50. [https://doi.org/10.36108/jrrslasu/8102/50\(0150\)](https://doi.org/10.36108/jrrslasu/8102/50(0150))
- Taleghani A, Emami SA, Tayarani-Najaran Z. 2020. Artemisia: a promising plant for the treatment of cancer. **Bioorg Med Chem** 28: 115180. <https://doi.org/10.1016/j.bmc.2019.115180>
- Tang D, Li J, Zhang R, Kang R, Klionsky DJ. 2020. Chloroquine in fighting COVID-19: good, bad, or both? **Autophagy** 16: 2273 - 2275. <https://doi.org/10.1080/15548627.2020.1796014>
- Taranto AG, Carneiro JWM, Araujo MT, Silva BM. 2006. Revisão de literatura sobre o mecanismo de ação da artemisinina e dos endoperóxidos antimaláricos - parte I. **Sitientibus** 34: 47 - 58.
- Tardío J, Arnal A, Lázaro A. 2021. Ethnobotany of the crab apple tree (*Malus sylvestris* (L.) Mill., Rosaceae) in Spain. **Genet Resour Crop Evol** 68: 795 - 808. <https://doi.org/10.1007/s10722-020-01026-y>
- Tarkang PA, Okalebo FA, Ayong LS, Agbor GA, Guantai AN. 2014. Anti-malarial activity of a polyherbal product (Nefang) during early and established *Plasmodium infection* in rodent models. **Malar J** 13: 1 - 11. <https://doi.org/10.1186/1475-2875-13-456>
- Taur DJ, Patil RY. 2011. Antiasthmatic activity of *Ricinus communis* L. roots. **Asian Pac J Trop Biomed** 1: 13 - 16. [https://doi.org/10.1016/S2221-1691\(11\)60113-5](https://doi.org/10.1016/S2221-1691(11)60113-5)
- Taur DJ, Waghmare MG, Bandal RS, Patil RY. 2011. Antinociceptive activity of *Ricinus communis* L. leaves. **Asian Pac J Trop Biomed** 1: 139 - 141. [https://doi.org/10.1016/S2221-1691\(11\)60012-9](https://doi.org/10.1016/S2221-1691(11)60012-9)
- Tedong L, Dimo T, Dzeufiet PDD, Asongalem AE, Sokeng DS, Callard P, Flejou JF, Kamtchouing P. 2006. Antihyperglycemic and renal protective activities of *Anacardium occidentale* (Anacardiaceae) leaves in streptozotocin induced diabetic rats. **Afr J Tradit Complement Altern Med** 3: 23 - 35. <https://doi.org/10.4314/ajtcam.v3i1.31136>
- Teske M, Trentini AMM. 1994. **Herbarium - Compêndio de Fitoterapia**. Herbarium Laboratório Botânico, Curitiba, Brasil.
- Thomás GE, Rodolfo HG, Juan MD, Georgina SF, Luis CG, Ingrid RB, Santiago GT. 2009. Proteolytic activity in enzymatic extracts from *Carica papaya* L. cv. Maradol harvest by-products. **Process Biochem** 44: 77 - 82. <https://doi.org/10.1016/j.procbio.2008.09.013>
- Tirloni CAS, Palozi RAC, Tomazetto TA, Vasconcelos PCP, Souza RIC, Santos AC, Almeida VP, Budel JM, Souza LM, Gasparotto Junior A. 2018. Ethnopharmacological approaches to kidney disease-prospecting an indigenous species from Brazilian Pantanal. **J Ethnopharmacol** 211: 47 - 57. <https://doi.org/10.1016/j.jep.2017.09.020>
- Toliopoulos I, Simos Y, Verginadis I, Oikonomidis S, Karkabounas S. 2012. NK cell stimulation by administration of vitamin C and *Aloe vera* juice *in vitro* and *in vivo*: A pilot study. **J Herb Med** 2: 29 - 33. <https://doi.org/10.1016/j.hermed.2012.04.002>
- Torres RC, Garbo AG, Walde RZML. 2015. Characterization and bioassay for larvicidal activity of *Anacardium occidentale* (cashew) shell waste fractions against dengue vector *Aedes aegypti*. **Parasitol Res** 114: 3699 - 3702. <https://doi.org/10.1007/s00436-015-4598-5>

- Tounekti T, Mahdhi M, Khemira H. 2019. Ethnobotanical study of indigenous medicinal plants of Jazan region, Saudi Arabia. **Evid Based Complement Alternat Med**. 2019: 1 - 44.
<https://doi.org/10.1155/2019/3190670>
- Traoré MS, Baldé MA, Camara A, Baldé ES, Diané S, Diallo MST, Maes L, Keita A, Cos P, Pieters L, Baldé AM. 2015. The malaria co-infection challenge: An investigation into the antimicrobial activity of selected Guinean medicinal plants. **J Ethnopharmacol** 174: 576 - 581. <https://doi.org/10.1016/j.jep.2015.03.008>
- Upreti Y, Asselin H, Boon E, Yadav S, Shrestha K. 2010. Indigenous use and bio-efficacy of Rasuwa district, central Nepal. **J Ethnobiol Ethnomed** 6: 1 - 10.
- Van Der Hoogte AR, Pieters T. 2016. Quinine, malaria, and the cinchona bureau: Marketing practices and knowledge circulation in a Dutch transoceanic cinchona-quinine enterprise (1920s-30s). **J Hist Med Allied Sci** 71: 197 - 225. <https://doi.org/10.1093/jhmas/jrv009>
- Vannier-Santos MA, De Castro SL. 2009. Electron microscopy in antiparasitic chemotherapy: A (close) view to a Kill. **Current Drug Targets** 10: 246 - 260. <https://doi.org/10.2174/138945009787581168>
- Vasco-dos-Santos DR, Santos JV, Andrade WM, Santos-Lima TM, Lima LN, Dias-Lima AG, Andrade MJG, Vannier-Santos MA, Moura GJB, Nunes ES. 2018. Antiparasitic plants used by the Kantaruré-Batida indigenous community (NE-Brazil): Ethnobotany and local knowledge-erosion risks. **Ambient Soc** 21: 1 - 20. <https://doi.org/10.1590/1809-4422asoc0011r1vu1811ao>
- Vasco-dos-Santos, DR, Pereira VC, Valões RMP, Dias-Lima AG, Armstrong AC, Santos CAB. 2019. Farmacopeia odontológica dos indígenas brasileiros: Uma revisão sistemática acerca do uso e bioatividade. **Espaço Ameríndio** 13: 278 - 298. <https://doi.org/10.22456/1982-6524.83781>
- Vasco-dos-Santos DR, Armstrong AC, Dias-Lima AG. 2020. Água, Saúde e Doença: Uma revisão sistemática sobre doenças de veiculação hídrica em comunidades indígenas brasileiras. **Revista Científica do UniRios** 2020: 226 - 246.
- Veeramani C, Pushpavalli G, Pugalendi KV. 2010. *In vivo* antioxidant and hypolipidemic effect of *Cardiospermum halicacabum* leaf extract in streptozotocin-induced diabetic rats. **J Basic Clin Physiol Pharmacol** 21: 107 - 125.
- Veeramani C, Al-Numair KS, Alsaif MA, Chandramohan G, Al-Numair NS, Pugalendi KV. 2012. Protective effect of *Cardiospermum halicacabum* leaf extract on glycoprotein components on STZ-induced hyperglycemic rats. **Asian Pac J Trop Med** 5: 939 - 944. [https://doi.org/10.1016/S1995-7645\(12\)60178-3](https://doi.org/10.1016/S1995-7645(12)60178-3)
- Veiga Junior VF, Pinto AC, Maciel MAM. 2005. Plantas medicinais: Cura segura? **Quím Nova** 28: 519 - 528.
<https://doi.org/10.1590/S0100-40422005000300026>
- Viana GSB, Bandeira MAM, Matos FJA. 2003. Analgesic and antiinflammatory effects of chalcones isolated from *Myracrodruon urundeuva* Allemão. **Phytomedicine** 10: 189 - 195.
<https://doi.org/10.1078/094471103321659924>
- Vieira LM, Castro CFS, Dias ALB, Silva AR. 2015. Fenóis totais, atividade antioxidante e inibição da enzima tirosinase de extratos de *Myracrodruon urundeuva* Fr. All. (Anacardiaceae). **Rev Bras Plant Med** 17: 521 - 527. https://doi.org/10.1590/1983-084X/13_033
- Voss C, Eyol E, Berger MR. 2006. Identification of potent anticancer activity in *Ximenia americana* aqueous extracts used by African traditional medicine. **Toxicol Appl Pharmacol** 211: 177 - 187.
<https://doi.org/10.1016/j.taap.2005.05.016>
- Waako PJ, Gumede B, Smith P, Folb PI. 2005. The *in vitro* and *in vivo* antimalarial activity of *Cardiospermum halicacabum* L. and *Momordica foetida* Schumch. Et Thonn. **J Ethnopharmacol** 99: 137 - 143.
<https://doi.org/10.1016/j.jep.2005.02.017>
- Wafa G, Amadou D, Larbi KM, Héla EFO. 2014. Larvicidal activity, phytochemical composition, and antioxidant properties of different parts of five populations of *Ricinus communis* L. **Ind Crops Prod** 56: 43 - 51.
<https://doi.org/10.1016/j.indcrop.2014.02.036>
- Watt G. 1907. **The wild and cultivated cotton plants of the world**. Longmans, London, UK.
<https://doi.org/10.5962/bhl.title.17182>
- Weremfo A, Adinortey MB, Pappoe ANM. 2011. Haemostatic effect of the stem juice of *Musa paradisiaca* L. (Musaceae) in guinea pigs. **Advan Biol Res** 5: 190 - 192.

- Whittemore R, Knafl K. 2005. The integrative review: Updated methodology. **J Advan Nurs** 52: 546 - 553. <https://doi.org/10.1111/j.1365-2648.2005.03621.x>
- Wijaya CH, Chen F. 2013. Flavour of papaya (*Carica papaya* L.) Fruit. **Biotropia** 20: 50 - 71. <https://doi.org/10.1159/btb.2013.20.1.288>
- Wink M. 2012. Medicinal plants: A source of anti-parasitic secondary metabolites. **Molecules** 17: 12771 - 12791. <https://doi.org/10.3390/molecules171112771>
- World Health Organization. 2020. **Traditional, Complementary and Integrative Medicine**. https://www.who.int/health-topics/traditionalcomplementary-and-integrative-medicine#tab=tab_1
- Yakubu MT, Nurudeen QO, Salimon SS, Yakubu MO, Jimoh RO, Nafiu MO, Akanji MA, Oladiji A. T, Williams FE. 2015. Antidiarrhoeal activity of *Musa paradisiaca* sap in wistar rats. **Evid Based Complement Alternat Med** 2015: 1 - 9. <https://doi.org/10.1155/2015/683726>
- Yeung AWK, Heinrich M, Kijjoa A, Tzvetkov NT, Atanasov AG. 2020. The ethnopharmacological literature: An analysis of the scientific landscape. **J Ethnopharmacol** 250: 1 - 21. <https://doi.org/10.1016/j.jep.2019.112414>
- Yoshikawa M, Murakami T, Kadoya M, Matsuda H, Muraoka O, Yamahara J, Murakami N. 1996. Medicinal foodstuffs. III. Sugar beet. (1): Hypoglycemic oleanolic acid oligoglycosides, betavulgarosides I, II, III and IV, from the root of *Beta vulgaris* L. (Chenopodiaceae). **Chem Pharm Bull** 44: 1212 - 1217.
- Yun N, Lee CH, Lee SM. 2009. Protective effect of *Aloe vera* on polymicrobial sepsis in mice. **Food Chem Toxicol** 47: 1341 - 1348. <https://doi.org/10.1016/j.fct.2009.03.013>
- Zahir AA, Rahuman AA, Bagavan A, Geetha K, Kamaraj C, Elango G. (2012). Evaluation of medicinal plant extracts and isolated compound epicatchin from *Ricinus communis* against *Paramphistomum cervi*. **Parasitol Res** 111: 1629 - 1635. <https://doi.org/10.1007/s00436-011-2589-8>
- Zhou H. 2014. **Treatment methods of traditional chinese medicine for infection with Ascaris lumbricoides and other nematodes**, In Mehlhorn H, Wu Z, Ye B. Treatment of human parasitosis in traditional chinese medicine. Springer-Verlag Berlin Heidelberg, Germany. <https://doi.org/10.1007/978-3-642-39824-7>
- Ziaei A, Sahranavard S, Gharagozlou MJ, Faizi M. 2016. Preliminary investigation of the effects of topical mixture of *Lawsonia inermis* L. and *Ricinus communis* L. leaves extract in treatment of osteoarthritis using MIA model in rats. **Daru** 24: 1 - 10. <https://doi.org/10.1186/s40199-016-0152-y>
- Zunjar V, Dash RP, Jivrajani M, Trivedi B, Nivsarkar M. 2016. Antithrombocytopenic activity of carpaine and alkaloidal extract of *Carica papaya* Linn. leaves in busulfan induced thrombocytopenic Wistar rats. **J Ethnopharmacol** 181: 20 - 25. <https://doi.org/10.1016/j.jep.2016.01.035>