

Revisión / Review

Ecotoxicity assays in the evaluation of molluskicidal agents from natural products in freshwater mollusks of medical importance: Lack of information and the choice of tested organisms

[Ensayos de ecotoxicidad en la evaluación de agentes molusquicidas derivados de productos naturales en moluscos de agua dulce de importancia médica: falta de información y elección de organismos evaluados]

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Abstract: Several natural products have been tested on mollusks of medical importance aiming to control these animals. Diverse scientific groups have demonstrated interest in finding new molluskicides from natural origin. In the literature, many promising studies have examined the molluskicidal activity of natural products on freshwater mollusks with medical importance. We used the keywords "natural products" and "molluskicidal activity" in diverse scientific databanks. A total of 301 articles were found, 25 of which were used in this letter. Most of these articles did not present ecotoxicity test reports. Some studies have reported ecotoxicity assays using nontarget organisms that do not represent freshwater ecosystems. Few articles showed ecotoxicity test reports using appropriate nontarget organisms, and only 1 presented test with more than 1 representative organism. The present work demonstrates the need to realize ecotoxicity tests using representative organisms in studies with natural products on freshwater mollusks of medical importance.

Keywords: Environmental toxicity; Ecotoxicology; Ecotoxicity models; Plants; Molluscicide

Resumen: Varios productos naturales han sido evaluados en moluscos de importancia médica con el objetivo de controlar estos animales. Diversos grupos científicos han mostrado interés en encontrar nuevos molusquicidas de origen natural. En la literatura, muchos estudios prometedores han examinado la actividad molusquicida de productos naturales en moluscos de agua dulce con importancia médica. Se utilizaron las palabras clave "productos naturales" y "actividad molusquicida" en diversas bases de datos científicas. Se encontraron un total de 301 artículos, de los cuales 25 fueron utilizados en esta comunicación. La mayoría de estos artículos no presentaron informes de pruebas de ecotoxicidad. Algunos estudios reportaron ensayos de ecotoxicidad utilizando organismos no objetivo que no representan los ecosistemas de agua dulce. Pocos artículos mostraron informes de pruebas de ecotoxicidad utilizando organismos no objetivo apropiados, y solo uno presentó pruebas con más de un organismo representativo. El presente trabajo demuestra la necesidad de realizar pruebas de ecotoxicidad usando organismos representativos en estudios con productos naturales sobre moluscos de agua dulce de importancia médica.

Palabras clave: Toxicidad ambiental; Ecotoxicología; Modelos de ecotoxicidad; Plantas; Molusquicida

INTRODUCTION

Several natural products have been tested on mollusks of medical importance with the aim of controlling the populations of these animals. A large number of human and animal parasites have freshwater snails as intermediate hosts (Chontanarith et al., 2017; Lee et al., 2017). In this context, many methods have been used to control snail populations (Abd El-Ghany & Abd El Ghany 2017), with the use of chemical molluskicidal products being one of these methods (Abdel-Ghaffar et al., 2016). However, these products have some problems reported in the literature, such as environmental toxicity and the need for repeated applications, among others (Rey 1987; Coura-Filho et al., 1992; Giovanelli et al., 2002; Dai et al., 2010; Oliveira-Filho et al., 2010; Lopes et al., 2011; Famakinde, 2018), which has raised interest in natural molluscicides (Elsareh et al., 2016). Several promising works in the literature have analyzed natural products with molluskicidal activity in freshwater mollusks of medical importance. The keywords "natural products" and "molluskicidal activity" were used. A total of 301 articles were found. Of these, 25 works were used in this letter.

MAIN TEXT

Shen et al. (2018), tested extracts of aerial parts of *Solidago canadensis* L. on the snail *Pomacea canaliculata* Lam, an intermediate host of the worm *Angiostrongylus cantonensis* (Song et al., 2016), which causes eosinophilic meningitis (Kliks & Palumbo, 1992). The petroleum ether extract of *S. canadensis* caused molluskicidal effect with an LC₅₀ value of 180.0 mg/L in 48 h, showing greater activity compared to the positive control, the molluskicidal Mie Luo Yu Kang (Shen et al., 2018). In another study involving the snail *P. caniculata*, Yang et al. (2022), tested arecoline (1), a major biologically active compound of areca nut, on these snails (Coppola & Mondola, 2012; Yuan et al., 2012). Arecoline obtained LC₅₀ values of 1.05, 2.12, and 3.54 mg/L in 72 h for snails with 15 ± 2, 25 ± 2 and 35 ± 2 mm shell diameters, respectively (Yang et al., 2022).

Despite the promising results, the work by Shen et al. (2018) and Yang et al. (2022), did not present ecotoxicity tests to assess the effects of their evaluated products on nontarget species. Other promising studies have investigated the molluskicidal

activity of natural products on *P. caniculata*. However, they did not exhibit reports of ecotoxicity tests (Yang et al., 2017; Yang et al., 2019; Li & Zou, 2019; Chen et al., 2020; Zhang & Zou, 2020).

Ke et al. (2019), investigated the molluskicidal activity of benzo[c]phenanthridine quaternary alkaloids (QBAs) (2) from fruits of the *Macleaya cordata* species on *Oncomelania hupensis*. This snail is the only intermediate host of *Schistosoma japonica*, a helminth that transmits schistosomiasis (Ross et al., 2001; Ke et al., 2017). The authors used QBAs at concentrations of 2.5 mg/L and 10.0 mg/L and observed mortality of more than 90% of snails in 48 h, a result similar to that observed for niclosamide (Ke et al., 2019).

Ke et al. (2019), also did not report ecotoxicity tests after assays using *P. caniculata*, as well as other studies testing the molluskicidal activity of natural products such as *O. hupensis* (Ke et al., 2017; Xing et al., 2021).

Ibrahim & Ghoname (2018), evaluated the molluskicidal activity of the aqueous extract of leaves of *Anagallis arvensis* on the snail *Biomphalaria alexandrina*. Several species of the genus *Biomphalaria* act as intermediate hosts for helminths of the genus *Schistosoma*, which cause schistosomiasis (WHO, 2017a). The aqueous extract of *A. arvensis* showed LC₅₀ and LC₉₀ values of 37.9 and 48.3 mg/L, respectively (Ibrahim & Ghoname, 2018).

This study also did not present ecotoxicity tests or other studies reported in the literature involving *Biomphalaria* spp. snails (Mandefro et al., 2018; Mendes et al., 2018; de Carvalho Augusto et al., 2020; Matos et al., 2020; Silva et al., 2020).

Natural products are not exempt from risks to the environment. Therefore, the WHO Pesticide Evaluation Scheme (WHOPES), aiming to ensure the use of compounds in aquatic environments in tolerable and nontoxic concentrations, recommends checking the toxicity of these substances (Canesi et al., 2015; Schiavo et al., 2017; Schiffer & Liber 2017; WHO, 2017b).

Some works observed in the literature proposed testing the molluskicidal activity of natural products on freshwater snails of medical importance using the microcrustacean *Artemia salina* as a test organism (Rocha-Filho et al., 2015; Martins et al., 2017; Araújo et al., 2018; Silva et al., 2019; Aguiar et al., 2022; Batista et al., 2022). Although this

organism is indicated as an experimental model because of its high sensitivity in ecotoxicological monitoring of new products (Lima *et al.*, 2002; Albuquerque *et al.*, 2014; Martins *et al.*, 2014), the *A. salina* habitat is a saltwater environment. Therefore, this species does not live in the same habitat as the freshwater snails mentioned in this work.

A recommended organism for ecotoxicological testing is the fish *Danio rerio*. Thus,

some scientific groups are using this model for evaluating ambient toxicity (Pereira *et al.*, 2017; Jia *et al.*, 2019). In another work, a species of water bug, *Diplonychus rusticus* Fabricius, was used to evaluate ecotoxicity (Hung *et al.*, 2021).

The information analyzed above was included in Table No. 1 to enable a better analysis of the researched works.

Table No. 1
Mollusk species, ecotoxicity assays used, and animal model

Publications	Mollusk	Ecotoxicity Assays	Model
Shen <i>et al.</i> , 2018	<i>P. canaliculata</i>	X	-
Yang <i>et al.</i> , 2022	<i>P. canaliculata</i>	X	-
Chen <i>et al.</i> , 2020	<i>P. canaliculata</i>	X	-
Zhang & Zou, 2020	<i>P. canaliculata</i>	X	-
Yang <i>et al.</i> , 2017	<i>P. canaliculata</i>	X	-
Yang <i>et al.</i> , 2019	<i>P. canaliculata</i>	X	-
Li & Zou, 2019	<i>P. canaliculata</i>	X	-
Ke <i>et al.</i> , 2019	<i>O. hupensis</i>	X	-
Ke <i>et al.</i> , 2017	<i>O. hupensis</i>	X	-
Xing <i>et al.</i> , 2021	<i>O. hupensis</i>	X	-
Ibrahim & Ghoname, 2018	<i>B. alexandrina</i>	X	-
Mandefro <i>et al.</i> , 2018	<i>B. pfeifferi</i>	X	-
Mendes <i>et al.</i> , 2018	<i>B. glabrata</i>	X	-
de Carvalho Augusto <i>et al.</i> , 2020	<i>B. glabrata</i>	X	-
Matos <i>et al.</i> , 2020	<i>B. glabrata</i>	X	-
Silva <i>et al.</i> , 2020	<i>B. glabrata</i>	X	-
Rocha-Filho <i>et al.</i> , 2015	<i>B. glabrata</i>	✓	<i>A. salina</i>
Martins <i>et al.</i> , 2017	<i>B. glabrata</i>	✓	<i>A. salina</i>
Araújo <i>et al.</i> , 2018	<i>B. glabrata</i>	✓	<i>A. salina</i>
Silva <i>et al.</i> , 2019	<i>B. glabrata</i>	✓	<i>A. salina</i>
Aguiar <i>et al.</i> , 2022	<i>B. glabrata</i>	✓	<i>A. salina</i>
Batista <i>et al.</i> , 2022	<i>B. glabrata</i>	✓	<i>A. salina</i>
Pereira <i>et al.</i> , 2017	<i>B. glabrata</i>	✓	<i>D. rerio</i>
Jia <i>et al.</i> , 2019	<i>O. hupensis</i>	✓	<i>D. rerio</i>
	<i>B. alexandrina</i>		<i>C. japonica</i>
	<i>Bulinus truncatus</i>		<i>M. nipponense</i>
Hung <i>et al.</i> , 2021	<i>Pomacea canaliculata</i>	✓	<i>D. rusticus</i>
	<i>Gyraulus convexiusculus</i>		
	<i>Tarebia granifera</i>		

Jia *et al.* (2019), used more than one species to evaluate ecotoxicity. In addition to the already mentioned *D. rerio*, the authors also tested their product on the bird *Coturnix japonica* and on the freshwater shrimp *Macrobrachium nipponense*. Silva *et al.* (2015), indicated the use of more than one organism for ecotoxicological tests, providing greater reliability for the research. The authors realized tests with creatures of different trophic levels since different organisms may exhibit different sensitivities to substances. Therefore, Jia *et al.* (2019), is interesting since the authors evaluated more than one trophic level and representative organisms of the local fauna, which may live in the same habitat as the tested snail.

CONCLUSION

Among a quantity of 25 works used in this letter, 16 did not present reports of ecotoxicity tests, and 6 presented ecotoxicity tests. However, they did not use a freshwater ecosystem organism as a nontarget species. Only 3 publications presented reports of ecotoxicity tests with a representative model, and only 1 used more than one organism to assess ecotoxicity (Table No. 2). Therefore, the present work demonstrates the need to perform ecotoxicity tests in studies investigating the possible molluscicidal action of natural products on freshwater snails with medical importance. Furthermore, ecotoxicity tests using organisms of different trophic levels that are present in the natural habitat of studied snails are strongly recommended.

Table No. 2

Number of studies with ecotoxicity assays; representative models; and number of tested organisms

Number of Works	Works that presented ecotoxicity assays	Works that used a representative model	Works that used more than one organism
25	6	3	1

REFERENCES

- Abd El-Ghany AM, Abd El-Ghany NM. 2017. Molluscicidal activity of *Bacillus thuringiensis* strains against *Biomphalaria alexandrina* snails. *Beni Suef Univ J Basic Appl Sci* 6: 391 - 393. <https://doi.org/10.1016/j.bjbas.2017.05.003>
- Abdel-Ghafar F, Ahmed AK, Bakry F, Rabei I, Ibrahim AM. 2016. The impact of three herbicides on biological and histological aspects of *Biomphalaria alexandrina*, intermediate host of *Schistosoma mansoni*. *Malacologia* 59: 197 - 210. <https://doi.org/10.4002/040.059.0201>
- Aguiar TWA, Batista JJ, Ferreira SAO, Sampaio MVL, Pereira DR, Ferreira MRA, Soares LAL, Melo AMMA, Albuquerque MCPA, Aires AL, Araújo HDA, Coelho LCBB. 2022. Effect of *Bauhinia monandra* Kurz leaf preparations on embryonic stages and adult snails of *Biomphalaria glabrata* (Say, 1818), *Schistosoma mansoni* Cercariae and toxicity in *Artemia salina*. *Molecules* 5: 4993. <https://doi.org/10.3390/molecules27154993>
- Albuquerque LP, Pontual EV, Santana GMS, Silva LRS, Aguiar JS, Coelho LCBB, Rêgo MJBM, Pitta MGR, Silva TG, Melo AMMA, Napoleão TH, Paiva PMG. 2014. Toxic effects of *Microgramma vacciniifolia* rhizome lectin on *Artemia salina*, human cells, and the schistosomiasis vector *Biomphalaria glabrata*. *Acta Trop* 138: 23 - 27. <https://doi.org/10.1016/j.actatropica.2014.06.005>
- Araújo HDA, Silva LRS, Siqueira WN, Fonseca CSM, Silva NH, Melo AMMA, Martins MCB, Lima VLM. 2018. Toxicity of usnic acid from *Cladonia substellata* (Lichen) to embryos and adults of *Biomphalaria glabrata*. *Acta Trop* 179: 39 - 43. <https://doi.org/10.1016/j.actatropica.2017.11.007>
- Batista JJ, de Araújo HDA, Aguiar TWA, Ferreira SAO, Lima MV, Pereira DR, Ferreira MRA, Soares LAL, Melo AMMA, Albuquerque MCPA, Aires AL, Coelho LCBB. 2022. Toxic, cytotoxic and genotoxic effect of saline extract and fraction of *Parkia pendula* seeds in the developmental stages of *Biomphalaria glabrata* (Say 1818 - intermediate host) and cercaricide activity against the infectious agent of schistosomiasis. *Acta Trop* 228: 106312. <https://doi.org/10.1016/j.actatropica.2022.106312>

- Canesi L, Ciacci C, Bergami E, Monopoli MP, Dawson KA, Papa S, Canonico B, Corsi I. 2015. Evidence for immunomodulation and apoptotic processes induced by cationic polystyrene nanoparticles in the hemocytes of the marine bivalve *Mytilus*. **Mar Environ Res** 111: 34 - 40.
<https://doi.org/10.1016/j.marenvres.2015.06.008>
- Chen HB, Lv TX, Zhang M, Liao JX, Chang XL, Yue GZ, Li PL, Zhao XM, Qiu XY, Qian Y, Yang CP. 2020. Two new compounds from the roots of *Pueraria peduncularis* and their molluscicidal effects on *Pomacea canaliculata*. **J Asian Nat Prod Res** 22: 144 - 152. <https://doi.org/10.1080/10286020.2018.1540597>
- Chontanarith T, Tejangkura T, Wetchasart N, Chimburut C. 2017. Morphological characteristics and phylogenetic trends of trematode cercariae in freshwater snails from Nakhon Nayok Province, Thailand. **Korean J Parasitol** 55: 47 - 54. <https://doi.org/10.3347/kjp.2017.55.1.47>
- Coppola M, Mondola R. 2012. Potential action of betel alkaloids on positive and negative symptoms of schizophrenia: a review. **Nord J Psychiatry** 662: 73 - 78. <https://doi.org/10.3109/08039488.2011.605172>
- Coura-Filho P, Rocha RS, Costa MFFL, Katz N. 1992. A municipal level approach to the management of schistosomiasis control in Peri-Peri, MG, Brasil. **Rev Inst Med Trop São Paulo** 34: 543 - 548.
<https://doi.org/10.1590/s0036-46651992000600008>
- Dai JR, Coles GC, Wang W, Liang YS. 2010. Toxicity of a novel suspension concentrate of niclosamide against *Biomphalaria glabrata*. **Trans Royal Soc Trop Med Hyg** 104: 304 - 306.
<https://doi.org/10.1016/j.trstmh.2009.07.015>
- de Carvalho Augusto R, Merad N, Rognon A, Gourbal B, Bertrand C, Djabou N, Duval D. 2020. Molluscicidal and parasitocidal activities of *Eryngium triquetrum* essential oil on *Schistosoma mansoni* and its intermediate snail host *Biomphalaria glabrata*, a double impact. **Parasit Vectors** 13: 486.
<https://doi.org/10.1186/s13071-020-04367-w>
- Famakinde DO. 2018. Treading the path towards genetic control of snail resistance to schistosome infection. **Trop Med Infect Dis** 3: 1 - 15. <https://doi.org/10.3390/tropicalmed3030086>
- Giovanelli A, Silva CLPAC, Medeiros L, Vaconcellos MC. 2002. The molluscicidal activity of niclosamide (Bayluscide WP70®) on *Melanoides tuberculata* (Thiaridae), a snail associated with habitats of *Biomphalaria glabrata* (Planorbidae). **Memórias do Instituto Oswaldo Cruz** 97: 743 - 745.
- Hung NH, Dai DN, Satyal P, Huong LT, Chinh BT, Hung DQ, Tai TA, Setzer WN. 2021. *Lantana camara* essential oils from Vietnam: Chemical composition, molluscicidal, and mosquito larvicidal activity. **Chem Biodivers** 18: e2100145. <https://doi.org/10.1002/cbdv.202100145>
- Ibrahim AM, Ghoname SI. 2018. Molluscicidal impacts of *Anagallis arvensis* aqueous extract on biological, hormonal, histological and molecular aspects of *Biomphalaria alexandrina* snails. **Exp Parasitol** 192: 36 - 41. <https://doi.org/10.1016/j.exppara.2018.07.014>
- Jia TW, Wang W, Sun LP, Lv S, Yang K, Zhang NM, Huang XB, Liu JB, Liu HC, Liu RH, Gawish FA, Habib MR, El-Emam MA, King CH, Zhou XN. 2019. Molluscicidal effectiveness of Luo-Wei, a novel plant-derived molluscicide, against *Oncomelania hupensis*, *Biomphalaria alexandrina* and *Bulinus truncatus*. **Infect Dis Poverty** 8: 42. <https://doi.org/10.1186/s40249-019-0535-7>
- Ke W, Lin X, Yu Z, Sun Q, Zhang Q. 2017. Molluscicidal activity and physiological toxicity of *Macleaya cordata* alkaloids components on snail *Oncomelania hupensis*. **Pestic Biochem Physiol** 143: 111 - 115.
<https://doi.org/10.1016/j.pestbp.2017.08.016>
- Ke W, Tu C, Cao D, Lin X, Sun Q, Zhang Q. 2019. Molluscicidal activity and physiological toxicity of quaternary benzo[c]phenanthridine alkaloids (QBAs) from *Macleaya cordata* fruits on *Oncomelania hupensis*. **Plos Negl Trop Dis** 13: e0007740. <https://doi.org/10.1371/journal.pntd.0007740>
- Kliks MM, Palumbo NE. 1992. Eosinophilic meningitis beyond the Pacific Basin: The global dispersal of a peridomestic zoonosis caused by *Angiostrongylus cantonensis*, the nematode lungworm of rats. **Soc Sci Med** 34: 199 - 212. [https://doi.org/10.1016/0277-9536\(92\)90097-a](https://doi.org/10.1016/0277-9536(92)90097-a)
- Lee JH, Quan JH, Choi IW, Park GM, Cha GH, Kim HJ, Yuk JM, Lee YH. 2017. Fasciola hepatica: infection status of freshwater snails collected from Gangwon-do (Province), Korea. **Korean J Parasitol** 55: 95.
<https://doi.org/10.3347/kjp.2017.55.1.95>

- Li S, Zou Z. 2019. Toxicity of *Chimonanthus nitens* flower extracts to the golden apple snail, *Pomacea canaliculata*. **Pestic Biochem Physiol** 160: 136 - 145. <https://doi.org/10.1016/j.pestbp.2019.07.015>
- Lima NMF, Santos AF, Porfírio Z, Goulart MO, Sant'Ana AEG. 2002. Toxicity of lapachol and isolapachol and their potassium salts against *Biomphalaria glabrata*, *Schistosoma mansoni* cercariae, *Artemia salina* and *Tilapia nilotica*. **Acta Trop** 83: 43 - 47. [https://doi.org/10.1016/S0001-706X\(02\)00055-4](https://doi.org/10.1016/S0001-706X(02)00055-4)
- Lopes TC, Gonçalves JRS, Silva-Souza N, Coutinho-Moraes DF, Amaral FMM, Rosa IG. 2011. Avaliação moluscicida e perfil fitoquímico das folhas de *Caryocar brasiliense* Camb. **Revista Cadernos de Pesquisa** 18: 23 - 30.
- Mandefro B, Mereta ST, Ambelu A. 2018. Efficacy of *Achyranthes áspera* (L.) as a molluscicidal bait formulation against fresh water snail *Biomphalaria pfeifferi*. **Evid Based Complement Alternat Med** 2018: 2718585. <https://doi.org/10.1155/2018/2718585>
- Martins MBC, Silva MC, Silva LRS, Lima VLM, Pereira EC, Falcão EPS, Melo AMMA, Silva NH. 2014. Usnic acid potassium salt: an alternative for the control of *Biomphalaria glabrata* (Say, 1818). **Plos One** 9. <https://doi.org/10.1371/journal.pone.0111102>
- Martins MC, Silva MC, Silva HA, Silva LR, Albuquerque MC, Aires AL, Falcão EP, Pereira EC, de Melo AM, da Silva NH. 2017. Barbatic acid offers a new possibility for control of *Biomphalaria glabrata* and schistosomiasis. **Molecules** 22: 568. <https://doi.org/10.3390/molecules22040568>
- Matos JL, da Silva KR, de Lima Paula LA, Cunha WR, Ramos SB, Rodrigues V, Cabral FJ, Magalhães LG. 2020. Molluscicidal and cercaricidal activities of curcumin on *Biomphalaria glabrata* and *Schistosoma mansoni* cercariae. **Pest Manag Sci** 76: 1228 - 1234. <https://doi.org/10.1002/ps.5631>
- Mendes RJA, Pereira Filho AA, Nogueira AJL, Araújo KRF, França CRC, Carvalho IB, Silva NMLD, Azevedo AS, Rosa IG. 2018. Evaluation of molluskicidal activity of three mangrove species (*Avicennia schaueriana*, *Laguncularia racemosa* and *Rhizophora mangle*) and their effects on the bioactivity of *Biomphalaria glabrata* Say, 1818. **Rev Inst Med Trop Sao Paulo** 60: e7. <https://doi.org/10.1590/s1678-9946201860007>
- Oliveira-Filho EC, Geraldino BR, Coelho DR, De-Carvalho RR, Paumgarten FJ. 2010. Comparative toxicity of *Euphorbia milii* latex and synthetic molluscicides to *Biomphalaria glabrata* embryos. **Chemosphere** 81: 218 - 227. <https://doi.org/10.1016/j.chemosphere.2010.06.038>
- Pereira LPLA, Dias CN, Miranda MV, Firmo WDCA, Rosa CDS, Santos PF, Brito MCA, Araruna FOS, Araruna FB, Silva-Souza N, Coutinho DF. 2017. Molluscicidal effect of *Euphorbia umbellata* (Pax) Bruyns latex on *Biomphalaria glabrata*, *Schistosoma mansoni* host snail. **Rev Inst Med Trop Sao Paulo** 59: e85. <https://doi.org/10.1590/S1678-9946201759085>
- Rey L. 1987. Estratégias e Métodos de Controle da Esquistossomose. **Cadernos de Saúde Pública** 3: 38 - 55.
- Rocha-Filho CA, Albuquerque LP, Silva LR, Silva PC, Coelho LC, Navarro DM, Albuquerque MC, Melo AM, Napoleão TH, Pontual EV, Paiva PM. 2015. Assessment of toxicity of *Moringa oleifera* flower extract to *Biomphalaria glabrata*, *Schistosoma mansoni* and *Artemia salina*. **Chemosphere** 132: 188 - 192. <https://doi.org/10.1016/j.chemosphere.2015.03.041>
- Ross AG, Sleight AC, Li Y, Davis GM, Williams GM, Jiang Z, Feng Z, McManus DP. 2001. Schistosomiasis in the People's Republic of China: Prospects and challenges for the 21st Century. **Clin Microbiol Ver** 14: 270 - 295. <https://doi.org/10.1128/CMR.14.2.270-295.2001>
- Schiavo S, Duroudier N, Bilbao E, Mikolaczyk M, Schäfer J, Cajaraville MP, Manzo S. 2017. Effects of PVP/PEI coated and uncoated silver NPs and PVP/PEI coating agent on three species of marine microalgae. **Sci Total Environ** 577: 45 - 53. <https://doi.org/10.1016/j.scitotenv.2016.10.051>
- Schiffer S, Liber K. 2017. Toxicity of aqueous vanadium to zooplankton and phyto- plankton species of relevance to the athabasca oil sands region. **Ecotoxicol Environ Saf** 137: 1 - 11. <https://doi.org/10.1016/j.ecoenv.2016.10.040>
- Shen X, Wang Z, Liu L, Zou Z. 2018. Molluscicidal activity of *Solidago canadensis* L. extracts on the snail *Pomacea canaliculata* Lam. **Pestic Biochem Physiol** 149: 104 - 112. <https://doi.org/10.1016/j.pestbp.2018.06.009>
- Silva DCVR, Pompeo MLM, Paiva TCB. 2015. **A ecotoxicologia no contexto atual no Brasil**. In: Pompeo M, Nishimura PY, da Silva SC, Doval JCL. (Ed.). *Ecologia de reservatórios e interfaces*. São Paulo, Brasil.

- Silva HAMF, Sá JLF, Siqueira WN, Lima MV, Martins MCB, Aires AL, Albuquerque MCPA, Falcão EPDS, Buril MLL, Pereira EC, Melo AMMA, Silva NHD. 2019. Toxicological effects of *Ramalina aspera* (lichen) on *Biomphalaria glabrata* snails and *Schistosoma mansoni* cercariae. **Acta Trop** 196: 172 - 179. <https://doi.org/10.1016/j.actatropica.2019.05.010>
- Silva YRR, Silva LD, Rocha TL, Santos DBD, Bezerra JCB, Machado KB, Paula JAM, Amaral VCS. 2020. Molluscicidal activity of *Persea americana* Mill. (Lauraceae) stem bark ethanolic extract against the snail *Biomphalaria glabrata* (Say, 1818): a novel plant-derived molluscicide? **An Acad Bras Cienc** 92: e20200715. <https://doi.org/10.1590/0001-37652020200715>
- Song L, Wang X, Yang Z, Lv Z, Wu Z. 2016. *Angiostrongylus cantonensis* in the vector snails *Pomacea canaliculata* and *Achatina fulica* in China: A meta-analysis. **Parasitol Res** 115: 913 - 923. <https://doi.org/10.1007/s00436-015-4849-5>
- Xing Y, Zhang S, Qu G, Dai J, Yao J, Feng B. 2021. Discovery and validation of a novel target of molluscicides against *Oncomelania hupensis*, the intermediate host of *Schistosoma japonicum*. **Acta Trop** 221: 106003. <https://doi.org/10.1016/j.actatropica.2021.106003>
- WHO. (World Health Organization). 2017a. Field use of molluscicides in schistosomiasis control programmes: An operational manual for programme managers. <https://apps.who.int/iris/bitstream/handle/10665/254641/9789241511995-eng.pdf;jsessionid=3610D943F651F342A48502E779AF9A4F?sequence=1>
- WHO. (World Health Organization). 2017b. **Field use of molluscicides in schistosomiasis control programmes: An operational manual for programme managers**. Geneva. Switzerland.
- Yang C, Zhang M, Lei B, Gong G, Yue G, Chang X, Sun X, Tian Y, Chen H. 2017. Active saponins from root of *Pueraria peduncularis* (Grah. ex Benth.) Benth. and their molluscicidal effects on *Pomacea canaliculata*. **Pest Manag Sci** 73: 1143 - 1147. <https://doi.org/10.1002/ps.4432>
- Yang C, Lv T, Wang B, Qiu X, Luo L, Zhang M, Yue G, Qin G, Xie D, Chen H. 2019. The damaging effects of pedunsaponin a on *Pomacea canaliculata* hemocytes. **Toxins** 11: 390. <https://doi.org/10.3390/toxins11070390>
- Yang C, Wang Y, Ma Y, Liu J, Zhou Y, Yan X, Zhang M, Qiu X, Luo L, Gong G, Chen H. 2022. Research on the molluscicidal activity and molecular mechanisms of arecoline against *Pomacea canaliculata*. **Ecotoxicol Environ Saf** 246: 114198. <https://doi.org/10.1016/j.ecoenv.2022.114198>
- Yuan J, Yang D, Liang Y, Gao W, Ren Z, Zeng W, Wang B, Han J, Guo D. 2012. Alkaloids from areca (betel) nuts and their effects on human sperm motility *in vitro*. **J Food Sci** 774: 70 - 78. <https://doi.org/10.1111/j.1750-3841.2012.02653.x>
- Zhang L, Zou Z. 2020. Molluscicidal activity of fatty acids in the kernel of *Chimonanthus praecox* cv. Luteus against the golden apple snail *Pomacea canaliculata*. **Pestic Biochem Physiol** 167: 104620. <https://doi.org/10.1016/j.pestbp.2020.104620>